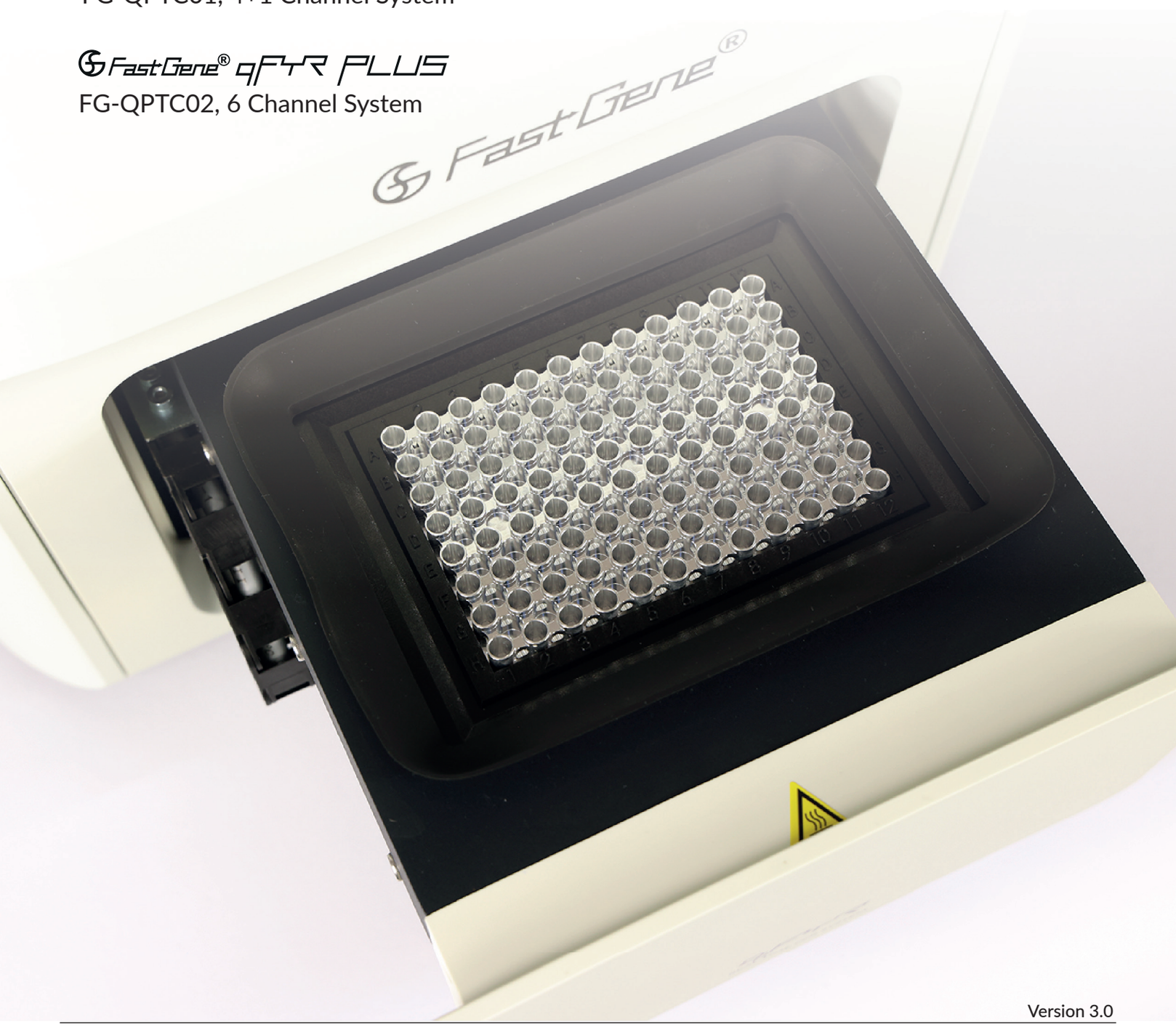
NIPPON Genetics EUROPE
INNOVATION FOR YOU

FastGene® qPTR

96-well Real-Time PCR Systems

FastGene® qPTR
FG-QPTC01, 4+1 Channel System

FastGene® qPTR PLUS
FG-QPTC02, 6 Channel System



Version 3.0

+49 2421 55496 0
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User manual version number:	V3.0
Type of instrument applicable:	FastGene® qFYR Real-Time PCR System (FG-QPTC01) FastGene® qFYR Plus Real-Time PCR System (FG-QPTC02)
Software release number:	V 2.0
Instrument production date:	See production label

Introduction

Welcome to the FastGene® qFYR 96-well Real-Time PCR Systems. Please read this instruction carefully so that you can use the instrument(s) correctly. After careful reading, please keep it for future inquiry when necessary.

Product information

Description	Real-Time PCR System
Product model	FastGene® qFYR Real-Time PCR System (FG-QPTC01) FastGene® qFYR Plus Real-Time PCR System (FG-QPTC02)
Software Name	FastGene®qFYR Analysis Studio
Software Release Number	V2.0
Product Size	350 x 520 x 370 mm (length x wide x height)
Net Weight	25 Kg

Enterprise information

Statement of intended use

FastGene® qFYR Real-Time PCR Systems are used to run real-time fluorescence PCR experiments (Real-time PCR) and to analyze the experimental data. The instrument(s) is/are operated in a laboratory with the corresponding reagents, to carry out rapid and accurate quantitative and qualitative detection of target nucleic acids sequences from human and animal samples (such as blood, oral swabs, nasopharyngeal swabs, body fluids, etc.) or other analytes, or to conduct melting curves, genotype analysis, etc.

Special statement

Before installing and using the instrument(s) for the first time, please read the instructions in this manual carefully. Please take into consideration the possible scope of maloperation or non-recommended usage and pay special attention to the possible consequences.

Intended usage

This product is for Research-Use-Only (RUO).

User requirements

FastGene® qFYR Real-Time PCR Systems must be operated by trained professionals, who have studied this user manual.

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1. Security and considerations

The operation, maintenance, repair, and transportation of FastGene® qFYR Real-Time PCR Systems must strictly comply with the safety precautions listed below. The design of the FastGene® qFYR Real-Time PCR Systems prevents biological pollution, ensures electrical safety protection and mechanical movement protection. Any operation that fails to comply with safety precautions or other tips stated in this manual may cause protection failure. It may also destroy the safety standards of design and manufacture and the expected scope of use of the instrument.
NIPPON Genetics EUROPE GmbH does not bear any responsibility for any loss caused by the user's failure to read this manual or to operate in accordance with the contents of this manual!

Please read this manual carefully before using the instrument. Wrong understanding or operation may lead to instrument damage, laboratory hazards, operator injuries, and lower instrument efficiency.

1.1. Security symbols and markings

Safety markings used on product packaging

Symbol	Heading	Description
	Fragile	The transport package is fragile, so handle it carefully.
	Upwards identification	Packaging should be kept facing up during transportation.
	Keep dry	The package should be away from rain.
	No heap height	It is indicated that the packing of this instrument is more than four layers during storage and transportation.
	Storage temperature	Storage environment temperature range: -20°C - 55°C.
	Storage humidity	Storage relative humidity: maximum 85%.

Safety markings used on product packaging

Symbol	Heading	Description
	Caution	This symbol is used to indicate that non- compliance with instructions or procedures may lead to physical injury or even death or could cause damage to the instrument. Consult the Operator‘s Manual.
	Biohazard	This symbol is used to indicate that certain precautions must be taken when working with potentially infectious material.
	Hot surface	Indicates the presence of a hot surface or other high-temperature hazard and to proceed with appropriate caution.
	Electricity hazard	Indicates the presence of high voltage at the instrument.
	Appliance class 1	Indicates that insulation of voltage-carrying parts (= basic insulation) and connection of touchable metal parts to the protective conductor (= grounding) protects against excessive touch voltages.
CE	CE symbol	This symbol indicates European conformity.
SN	Serial number	Identification of the serial number of the instrument.
	Date of manufacturing	The date of manufacturing will appear next to this symbol.
	Manufacturer	The name and the address of the manufacturer will appear next to this symbol.
	Disposal	Do not dispose of this product as unsorted municipal waste. Follow local municipal waste ordinances for proper disposal provisions to reduce the environmental impact of waste electrical and electronic equipment (WEEE).

1.2. General instrument safety and precautions

The operator shall not disassemble the instrument, replace components or debug the instrument(s) without the authorization of our company. If one of the above is needed, it must previously be approved by our company, and completed by trained professionals, otherwise unexpected consequences will occur.

Please avoid collision, damage the instrument(s) and store carefully. The user should immediately unplug the instrument(s) from the power outlet and contact the supplier or ask our professional maintenance staff if one of the following is to happen:

- The instrument(s) is subjected to rain or flooding.
- Abnormal sound or smell in the instrument(s).

- Collision or damage to the enclosure.
- The function of the instrument(s) changed.

It is forbidden to carry or move the instrument during the operation of the instrument(s).

The openings on this instrument(s) are ventilated. In order to avoid excessive temperature, **do not block or cover these vents during the operation of the instrument(s)**, or use dust shields and any other substances to cover the surface of the instrument(s).

If you need to install or transport this instrument(s), please contact our company to request professional personnel or professional guidance. Otherwise, we will not bear any responsibility for the damage caused by the instrument(s).

After the installation of the instrument(s), the transport lock should be properly retained in accordance with the requirements of the technical personnel.

During the operation of the instrument(s), do not forcibly open the sample draw, otherwise it will destroy the biological safety of the instrument(s), its electromagnetic radiation, and other protective measures.

Do not force consumables (8 tubes / test tubes /96 well plates, etc.) not matched with the instrument(s) into the sample draw.

Failure to use the instrument(s) in accordance with the manufacturer‘s prescribed method may damage the protection provided by the instrument(s).

1.3. Personal safety and precautions

The weight of the instrument(s) is heavy. Moving or lifting the instrument(s) requires appropriate tools, methods, or cooperation with others to complete. Please, avoid moving the instrument(s) alone without the cooperation of others. Improper lifting of the instrument(s) may cause physical injury, pain and/or instrument(s) damage.

Do not touch the power plug, power cord or switch with wet hands.

High surface temperature

During the operation of the instrument,(s) the heating module of the sample draw may become very hot! During and after the experiment, do not touch the heating block directly with your hand or any part of the body to avoid scalding. Please, open the sample draw after the heating block temperature drops to standby temperature and take out the sample.

1.4. Electrical safety and precautions

Prohibition
FastGene® qFYR Real-Time PCR Systems hve a voltage harmful to the human body! At any time, before disassembling the instrument(s) shell, please cut off the instrument(s) power supply. It is forbidden to replace components when power lines are connected.

The shell of the FastGene® qFYR Real-Time PCR Systems should be properly grounded through the power ground wire. Any damage to the internal or external grounding path of the astGene® qFYR Real-Time PCR Systems may be dangerous.

Note: If leakage is found, please cut off the power immediately and stop using the instrument. Please cut off the power supply before moving the instrument(s).

Use the power cord provided with FastGene® qFYR Real-Time PCR Systems. If the original power cord is broken, worn, or disconnected, the same power cord should be replaced as soon as possible.

In order to avoid the danger of electric shock, the power cord of astGene® qFYR Real-Time PCR Systems must be reliably grounded. The power cord of FastGene® qFYR Real-Time PCR Systems is a standard plug power cord. Please insert the power plug into the appropriate three-wire grounding socket (the external network power supply connected by the socket must be reliably grounded) to ensure the safe use of the FastGene® qFYR Real-Time PCR Systems.

The electrical supply in which astGene® qFYR Real-Time PCR Systems is located must be connected with ground wire. Please carefully check whether the power connection is firm. Do not hard pull the power cord. When the plug is plugged into the power supply, make sure the plug is fully plugged into the socket.

Please keep the power plug and the power cable away from high temperature objects such as heater. When using FastGene® qFYR Real-Time PCR Systems, please do not place any items on the power cord, do not put the power cord in the place where people often walk.

The fuse type of FastGene® qFYR Real-Time PCR Systems is T10AH250VP, placed in the backup box at the power outlet at the back of the instrument(s). The use of improper fuses may damage the instrument‘s line system and cause fire. Before turning on the instrument(s) power switch, check and make sure the fuse is properly installed.

In order to prevent fire hazards, use FastGene® qFYR Real-Time PCR Systemsonly with the indicated fuse. When replacing the fuse, please cut off the power supply, pull out the power cord, pry open the fuse box with a word screwdriver, and replace the old fuse with the same new fuse.

1.5. Environmental safety and precautions

FastGene® qFYR Real-Time PCR Systems is only suitable for indoor use, indoor ventilation should be good, and free of corrosive gas.

EDM may cause explosion hazard and is prohibiting the use of this instrument in the presence or possible presence of flammable and explosive gases.

The working environment temperature of FastGene® qFYR Real-Time PCR Systems should be between 10°C-30°C and the relative humidity should ≤85%.

The working environment pressure of FastGene® qFYR Real-Time PCR Systems should be normal atmospheric pressure (altitude less than 2000 meters).

1.6. Biosafety and precautions

Biological hazard

The sample object of FastGene® qFYR Real-Time PCR Systems is nucleic acid. In practice, please regard it as a biological sample with potential biological hazard. When handling and operating samples, generally applicable safety precautions shall be taken and appropriate protective goggles, clothing and gloves shall be worn.

Users are requested to comply with all applicable local or national regulations to complete the disposal of waste samples and contaminated materials.

FastGene® qFYR Real-Time PCR Systems shall be treated as waste with biological pollution hazard. Before reuse, recycling, or disposal, the instrument needs to be purified (including cleaning, disinfection and/or sterilization). FastGene® qFYR Real-Time PCR Systems shall be handled in accordance with relevant local or local regulations.

During any operation, if liquid samples overflow, appropriate disinfectants should be quickly used for disinfection, to avoid the spread of pollutants to laboratory personnel or contaminated instruments.

1.7. Electromagnetic Compatibility (EMC) standards

FastGene® qFYR Real-Time PCR Systems should be installed and used according to the electromagnetic compatibility information in this chapter.

The electromagnetic compatibility of FastGene® qFYR Real-Time PCR Systems is classified into categories A and groups I.

1. FastGene® qFYR Real-Time PCR Systems meet the emission and immunity requirements specified in the EN 61326-1:2013 and EN 61326-2-6:2013.

2. FastGene® qFYR Real-Time PCR Systems are designed and tested according to A kinds of instruments in the CISPR 11:2016.
3. Portable and mobile RF communication instruments may affect the use of FastGene® qFYR Real-Time PCR Systems. It is recommended to stay away from or close the portable and mobile RF communication instruments when using FastGene® qFYR Real-Time PCR Systems normally.
4. FastGene® qFYR Real-Time PCR Systems must be used with the power cord provided by our company, the length of the power cord is not more than 1.5 m.

Except the accessories provided by our company, the use of other manufacturer accessories may lead to an increase in the emission of FastGene® qFYR Real-Time PCR Systems or a decrease in immunity.

1. Please evaluate the electromagnetic environment before FastGene® qFYR Real-Time PCR Systems are used.

2. Do not use FastGene® qFYR Real-Time PCR Systems next to strong radiation sources (such as unshielded RF sources), otherwise it may interfere with the normal operation of the instrument(s).

Note: It is the responsibility of the user to ensure the EMC environment of the instrument so that the instrument can work properly.

2. Product Overview

2.1. Scope of application

2.1.1. Intended use

FastGene® qFYR Real-Time PCR Systems are used to run real-time fluorescence PCR experiments (Real-time PCR) and to analyze the experimental data. The instruments are operated in a laboratory, in conjunction with the corresponding reagents, for rapid and accurate quantitative and qualitative detection of target nucleic acids from human samples (e.g., blood, oral swabs, nasopharyngeal swabs, body fluids, etc.) or other analytes in the patient's body, or for melting curves, genotype analysis.

2.1.2. Product intended use environment

Institutions such as medical institutions, laboratories, and companies that meet the requirements of the product operating environment:

1. diagnostic laboratory use
2. general laboratory research use. Its users are professional and technical personnel of the hospital laboratory or general laboratory and need specialized training for PCR laboratory technology and instrument, software operation, and have skilled relevant operation skills.

2.2. Application areas

- Basic scientific research
- Clinical detection of pathogens
- Genetic screening
- Cancer diagnosis
- Diagnosis and treatment of cardiovascular diseases
- Drug genome
- Public health and safety surveillance
- Food hygiene and quarantine
- Customs import and export quarantine
- Transgenic and pathogen detection in animals and plants
- Others

2.3. Basic principles

PCR principle is similar to the natural replication process of DNA. Its specificity depends on oligonucleotide primers complementary to both ends of the target sequence. PCR consists of three basic reaction steps: denaturation-annealing-extension. Repeated cyclic denaturation-annealing-extending processes can amplify the target gene by millions of times in 1~2 hours, for a high detection sensitivity.

The principle of real-time fluorescence PCR is to add a fluorescent reporter (probe or dye) to the PCR reaction system. The target nucleic acid derived from DNA samples is qualitatively or quantitatively detected by fluorescence detection system and real-time monitoring of the fluorescence value. Real-time fluorescence PCR detection has higher specificity and sensitivity than end-point PCR detection.

2.4. Main functions

FastGene® qFYR Real-Time PCR Systems are special instruments for real-time fluorescence detection. It integrates PCR amplification, fluorescence detection and data analysis. It can monitor the increase of fluorescence in each test tube in real time as the amplification increases. After the amplification, the software automatically processes the experimental data, carries on the quantitative/qualitative, melting curve or genotyping analysis to the sample, displays and prints the experimental results.

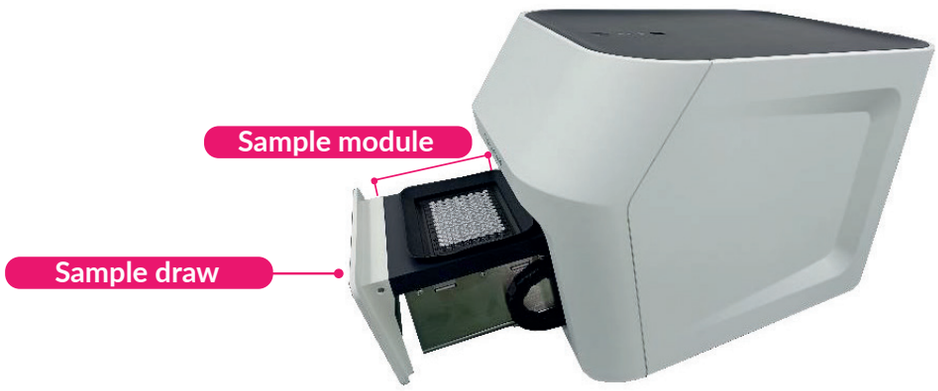
2.5. Product composition

FastGene® qFYR Real-Time PCR Systems mainly consist of electronic control system, power module, temperature control system, detection system, shell components and software (V2.0).

Product structure

1. Diagram of the structure of the FastGene® qFYR Real-Time PCR Systems

a)



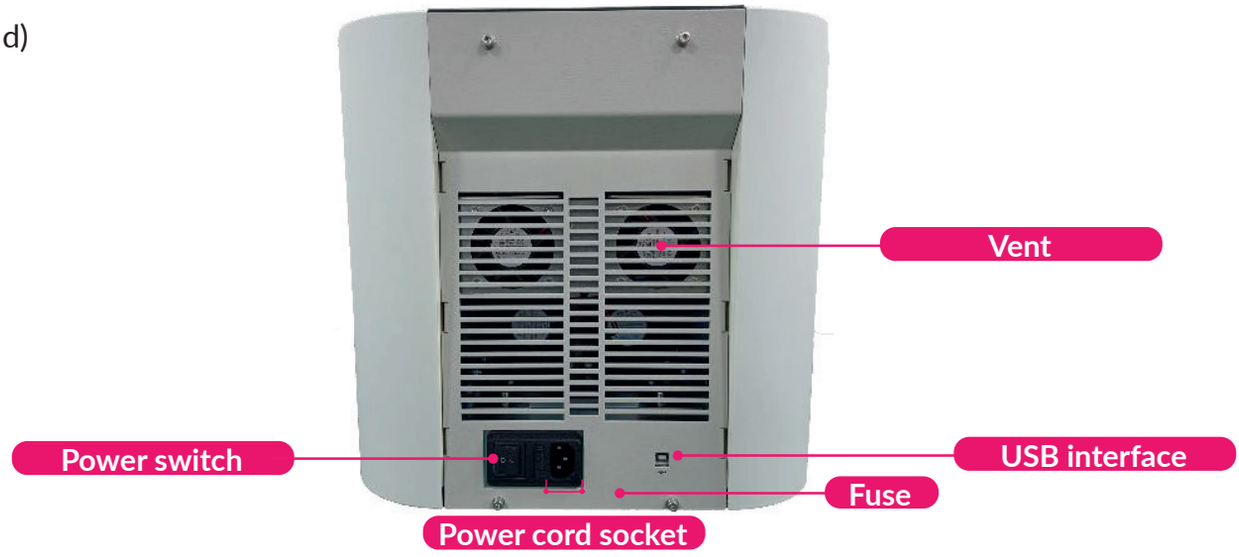
b)



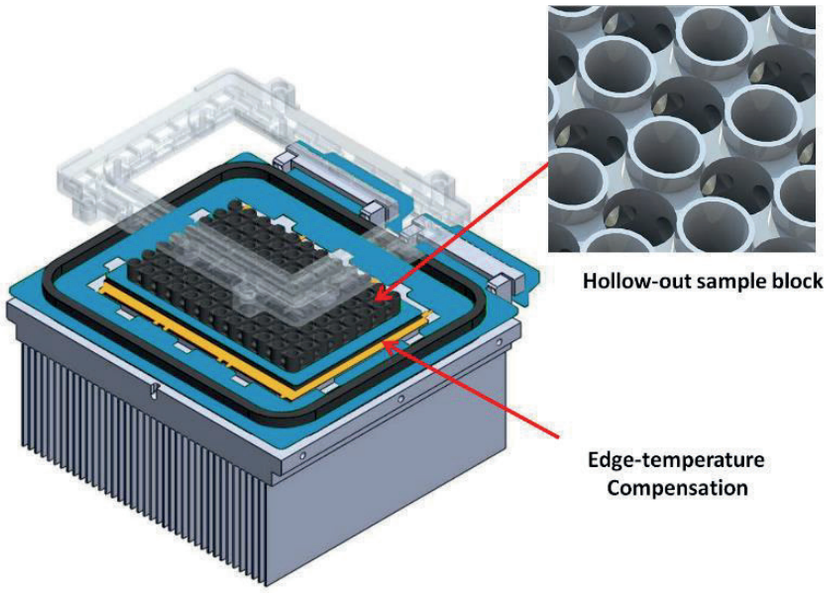
c)


Power indicator
In/Out button
Transport lock
Status indicator

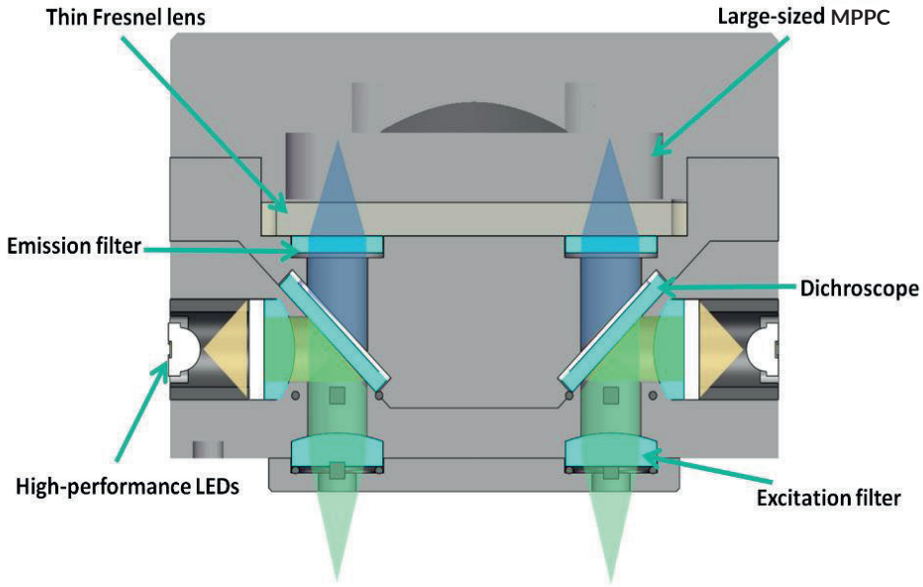
d)


Vent
Power switch
USB interface
Fuse
Power cord socket

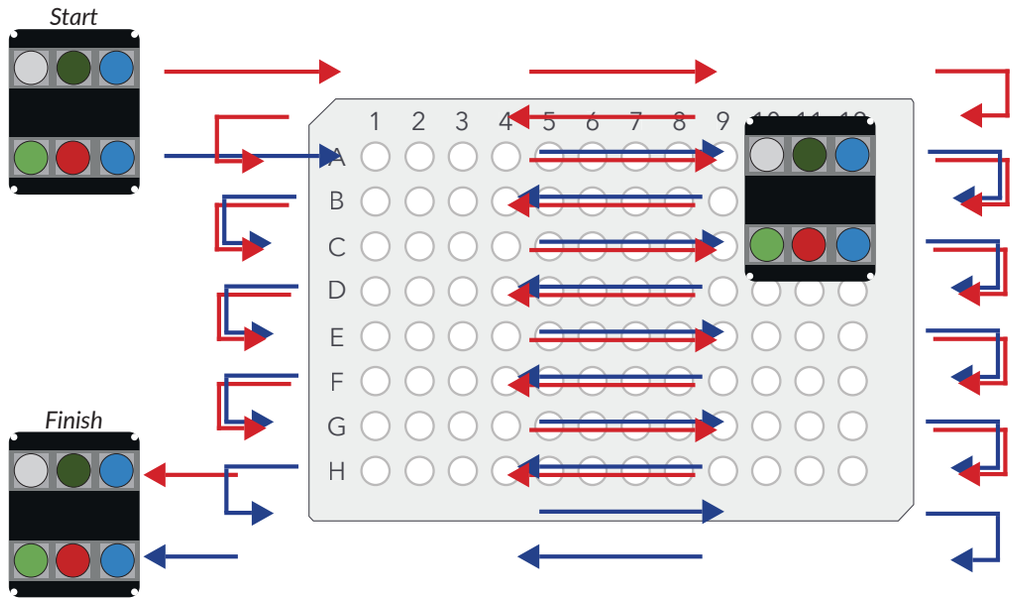
2. Schematic diagram of temperature control module.



3. Schematic diagram of optical path module.



4. Schematic illustration of optical scanning patterns.



2.6. Parameters and characteristics

2.6.1. Physical specifications

Instrument(s) specifications	Size: 350 mm(L) × 520 mm (W) × 370 mm (H). Weight: 25 kg
Packing specifications	Size: 710 mm(L) × 540 mm (W) × 580 mm (H). Weight: 36 kg
Power supply specification	Power supply voltage: AC 110-230 V Frequency: 50 Hz. Power: 750 w
Communication specifications	USB ports
Environmental conditions	Temperature: 10 °C - 30 °C. Humidity: ≤85% RH no condensation. Atmospheric pressure: 85.0 kPa ~ 106.0 kPa. Altitude: < 2000m
Storage and transportation environment	Temperature: -20 °C - 55 °C in the transport box. Humidity: ≤85% RH
Running noise	Maximum noise when running instrument does not exceed 60dB (A)

2.6.2. Technical parameters

	Optical detection system	
	FastGene® qFYR (FG-QPTC01)	**FastGene® qFYR Plus (FG-QPTC02)**
Excitation source	**4 single-color** high-efficiency LEDs (maintenance-free, working life >100,000 hours)	**6 single-color** high-efficiency LEDs (maintenance-free, working life >100,000 hours)
Detector	High-sensitivity solid-state photomultiplier (MPPC / SiPM)	High-sensitivity solid-state photomultiplier (MPPC / SiPM)
Detector position	Top of the block	Top of the block
Detection sensitivity	1 copy of the target sequence	1 copy of the target sequence
Scanning principle	Time-resolved scanning technology	Time-resolved scanning technology
Detection time	• Standard mode (full channel): 8.5 seconds/96-well plate; • **Fast mode (dual FAM): 4 seconds/96-well plate**	Standard mode (full channel): 8.5 seconds/96-well plate
Excitation/ detection	• Excitation range: 455–650 nm; • Scope of test: 510–715 nm	• **Excitation range: 415–685 nm;** • **Scope of test: 455–745 nm**
Channel number	4 channels (2x FAM)	**6 channels**
Dye compatibility	• FAM/SYBR Green • VIC/HEX/TET/JOE • ROX/Texas Red, Mustang Purple • Cy5/LIZ	• FAM/SYBR Green • VIC/HEX/TET/JOE • ROX/Texas Red, Mustang Purple • Cy5/LIZ • **Cy5.5/Quasar 705/Alexa Fluor 680** • **ATTO 425**
Repeatability of fluorescence intensity detection	CV ≤3%	CV ≤3%
Detection of fluorescence intensity	CV ≤5%	CV ≤5%
Sample repeatability	CV ≤ 3%	CV ≤ 3%
Linear fluorescence intensity detection	A linear regression coefficient | r | ≥0.990 in 5 concentration gradients	A linear regression coefficient | r | ≥0.990 in 5 concentration gradients
Sample linear correlation	A linear regression coefficient | ≥0.980 in 5 concentration gradients	A linear regression coefficient | ≥0.980 in 5 concentration gradients

	Thermal block	
	FastGene® qFYR (FG-QPTC01)	FastGene® qFYR Plus (FG-QPTC02)
Block capacity	96	96
Sample volume	5-100 µl	5-100 µl
Heating/ cooling method	Peltier (6 temperature control modules)	Peltier (6 temperature control modules)
Temperature control technology	Hollow-out module combined with edge temperature compensation technology	Hollow-out module combined with edge temperature compensation technology
Maximum ramp rate	• 6 °C/s (thermal block) • 4 °C/s (sample)	• 6 °C/s (thermal block) • 4 °C/s (sample)
Temperature setting range	4-100 °C	4-100 °C
Heated lid	Electronic automatic lid	Electronic automatic lid
Temperature accuracy	± 0.2 °C	± 0.2 °C
Temperature uniformity	± 0.2 °C	± 0.2 °C
Gradient zone	12 columns	12 columns
Gradient range	1-36 °C	**1-40 °C**
Linear dynamic range	10 orders of magnitude: 1-10^{10} copies	10 orders of magnitude: 1-10^{10} copies

2.6.3. Software features

- **Software interface:** Wizard interface, intuitive sample plate layout and program settings, easy to use operation editing.
- **Software functions**: Absolute quantitation, relative quantitation, genotyping, HRM-analysis, and other analytical functions to meet the requirements of a variety of experiments.
- **Language support:** English.
- **Data transmission:** FastGene® qFYR Real-Time PCR Systems can transmit the experimental data to the connected computer in real time during the experiment.
- **Overlong program settings:** Each stage may include up to 100 steps and up to 10 cycles.

2.6.4. Chemistry reagent features

- **PCR reagent:** Open reagent platform, all quantitative, qualitative PCR reagents are applicable.
- **Fluorescent dyes:** Suitable fluorescent dyes are, depending on the system;

FastGene® qFYR (FG-QPTC01)
- FAM/SYBR Green/FastGene® IC Green
- VIC/HEX/TET/JOE
- ROX/Texas Red
- Cy5

FastGene® qFYR Plus (FG-QPTC02)
- FAM/SYBR Green/FastGene® IC Green
- VIC/HEX/TET/JOE
- ROX/Texas Red
- Cy5
- Cy5.5/Quasar 705/Alexa Fluor 680
- ATTO 425

2.6.5. Consumables specifications

0.1 ml clear/white PCR thin-walled single tube, flat-topped transparent optical tube cover.
0.1 ml clear/white PCR thin-walled 8-well tubes, flat-top transparent optical tube cover.
0.1 ml clear/white PCR thin-walled 96-well plate with no hemline or hemline.

Note: FastGene® qFYR Real-Time PCR Systems is compatible with low 0.1 mL PCR flat top transparent tube cover reaction tube /8-well joint tube, no skirt or half skirt edge 96-well PCR reaction plate.

Do not use 0.2 mL PCR reaction tube and convex cover.

If the sample tubes are just a few, please use 8-well tubes for support on both sides, or use empty single tubes in the four corners to prevent uneven force on the plate slot.

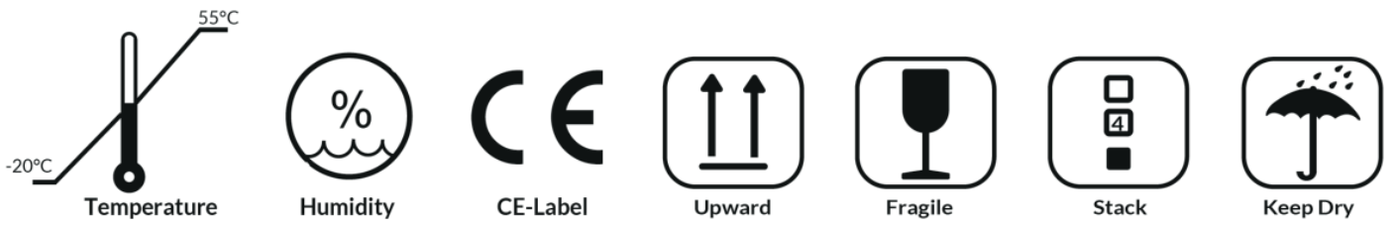
3. Installation and transportation

3.1. Transport and storage

Product transportation is carried out in accordance with the requirements of the product. Original packaging must be used in transportation to avoid damage to the instrument. FastGene® qFYR Real-Time PCR Systems in the packaging and transport state, should be stored in the ambient temperature of -20 °C-55 °C, relative humidity less than 85%, and the air cannot contain corrosive agents. The outer packaging of the product specifies the transport instructions allowed for this product, which are:

- **Packaged products are only allowed to be placed in the direction indicated by the arrow.**
- Please hold the handling process lightly.
- Maximum stack number is limited to four layers, do not exceed four layers stack.

Do not transport or store this product in rain or wet environment.



3.2. Unboxing

The outer packing of the product is cardboard box, filled with shock absorber foam. Please do not rush to open the packaging after receiving the FastGene® qFYR Real-Time PCR System(s). Carefully check the packaging´s appearance for damage. Please check the content you received if parts are missing and damaged after unpacking. If the product is damaged during transportation, please do not use it, and contact NIPPON Genetics EUROPE or the distributor!

3.3. Packing list

	Name	Unit
1	FastGene® qFYR Real-Time PCR System (FG-QPTC01) or FastGene® qFYR Plus Real-Time PCR System (FG-QPTC02)	1
2	Instrument power cable	1
3	Instrument data cable	1
4	Fuse	2
5	Hexagon spanner	1
6	Packing list	1
7	USB Stick (including software and electronic user manual)	1

3.4. Installation requirements

3.4.1. Instrument working environment requirements

- **Location:** The main unit must be placed on a stable, horizontal working table, avoiding direct sunlight, not near heating equipment. The instrument´s environment must be at ambient temperature of 10 - 30 °C, with a relative humidity less than 85%. For indoor use only.

- **Surrounding environment:** Do not install the main unit next to instruments with strong electromagnetic interference or high induction factors, such as refrigerators, high-speed centrifuges, oscillators, etc. The mainframe must be placed at least 15 cm from the surrounding object or wall for heat dissipation and ventilation of the FastGene® qFYR Real-Time PCR Systems, and for the convenience of users to turn on and off the instrument power supply.

Do not place the mainframe in a narrow space where it is difficult to operate and disconnect the power supply!

- **Power requirements:** 110-220 VAC,50/60Hz, voltage fluctuations <10 V.

- **Power outlet:** To prevent electrical shock, FastGene® qFYR Real-Time PCR Systems mainframe must be connected to a three-core grounding socket that meets the safety standards. The voltage is ~110-220 V (50-60 Hz) and the zero-ground voltage ≤5 V. If conditions permit, connect the instrument to a dedicated power outlet.

- **Warning:** Incorrect grounding may cause electrical shock or instrument damage. Confirm that the input voltage meets the requirements of the instrument and that the fuse has been installed in accordance with the specifications.

3.4.2. Computer configuration requirements

The computer installed with the FastGene® qFYR Analysis Studio must have the following configuration:

Processor	Intel or AMD dual-core CPU, 2.8GHz
Memory	4 GB
Hard disk	500 GB
Network card	10 M/100M adaptive network card (optional)
Display resolution	1440 × 900 or higher
Operating system	Windows 10 or above & Office Word/Excel 2007 or above

Note: The control computer of FastGene® qFYR Real-Time PCR Systems is not designed for online use. Connecting it to the network will have the risk of acquiring viruses and becoming the target of hacker attacks. NIPPON Genetics EUROPE is not responsible for any damage caused by an internet connection. It is not recommended to install other software in the control computer. There will be a potential risk of conflict with the instrument software module and may affect the reliability of the

results. NIPPON Genetics EUROPE does not provide antivirus software. Therefore, if necessary, take preventive measures to ensure that there is no virus on the computer used with any FastGene® qFYR Real-Time PCR System.

3.5. Equipment installation

The following contents are convenient for users to quickly complete the installation of the FastGene® qFYR Real-Time PCR Systems. Please be sure to operate according to the following contents. If you have any questions during the installation operation, please contact our support.

3.5.1. Equipment appearance



Status lamp

There are power status light “POW” and system status light “STU” on the equipment´s top part. The meaning of status lights is shown in the following:

Name	Status	Meaning
POW	Blue light	Ready
STU	Green light slowly flashing	Self-checking
	Green light quickly flashing	Working
	Green light on	In idle condition
	Yellow	Caution, able to continue working
	Red	Malfunction, stop working

Equipment switch

FastGene® qFYR Real-Time PCR Systems have a power switch in the back. Turn on the instrument by turning the switch to the „I“ position and switch off at the „O“ position.

Accessing the plate

The draw of the FastGene® qFYR Real-Time PCR Systems is controlled by the top button ⏏. The plate holder draw will open as soon as you press the top button. The in-and-out operation needs to be carried out in the idle state. It will not work during a PCR and during the self-inspection. The system does not respond to the in-and-out request.



Data communication

The interface on the back panel of FastGene® qFYR Real-Time PCR Systems is the USB, which is used for communication between the computer and the device.

Air inlet

The lower gap of the front panel of the FastGene® qFYR Real-Time PCR Systems is a set of air intakes. Blocking must be prevented when running. Pay special attention to this gap for air inlet, **it is strictly prohibited to access this area with your hands, as severe injuries can happen!**

3.5.2. Release instrument transport lock

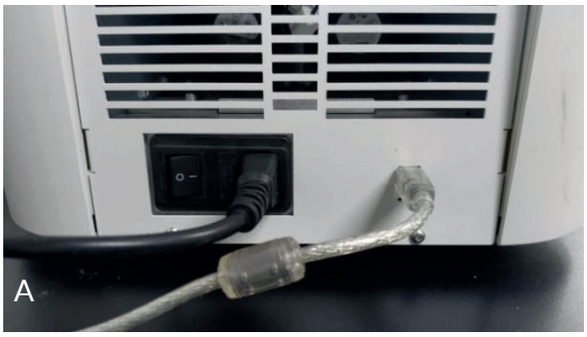
In order to avoid the damage of important parts of the FastGene® qFYR Real-Time PCR Systems during transportation, a transport lock is added. While the instrument(s) is(are) switched off, find the locking hole at the top of the instrument(s) and remove the soft rubber cap. Use the M4 hexagonal wrench to the inner bolt, press down slightly to the bottom and rotate 90 degrees counterclockwise. This will loosen the wrench, and the transport lock will pop up due to the of an internal spring mechanism of the instrument(s) (as shown below).

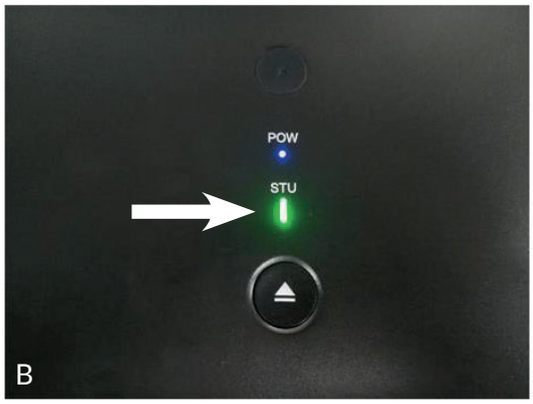


STU light status will change from slow flicker to green light on, unlocked successfully.

3.5.3. First installation and run protocol

Insert the USB data cable and the power cable (as shown in figure A below) and connect data cable to the computer. After turning on the power supply, press the power switch at the back end of the instrument(s) to turn on the instrument(s), and wait until the power indicator light (POW) is constantly on, and the system begins self-checking process. When the system self-test is completed, the instrument(s)´s normal state lamp (STU) is green (as shown in figure B). Press the plate remove button and remove the support plate in the sample platform (as shown in figure C). The installation steps are completed.

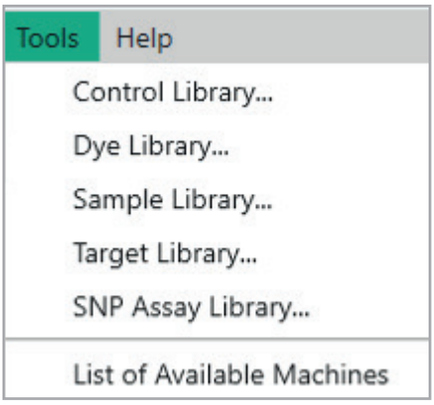


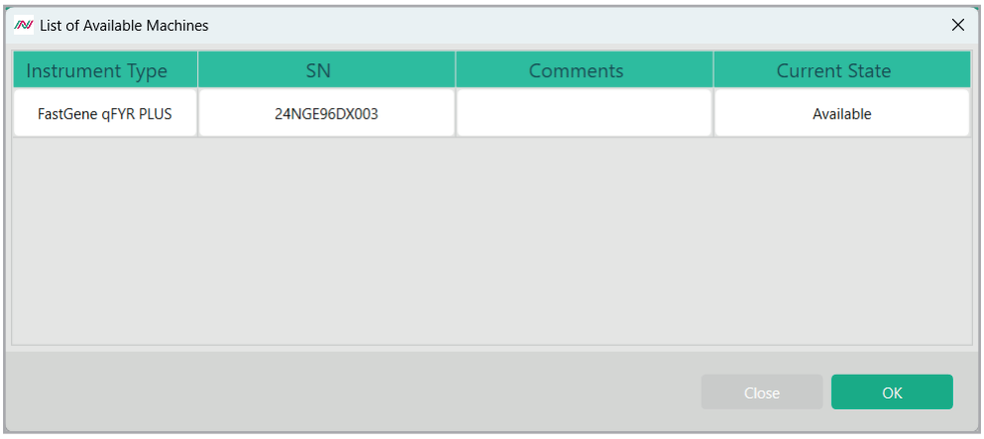




It is possible to connect multiple qFYR (Plus) Real-Time PCR systems to one PC. The connection of several USB data cables to one PC is necessary for that.

To view the connected devices, click on „Tools“ and then on „List of Available Machines“.





3.6. Instrument software installation

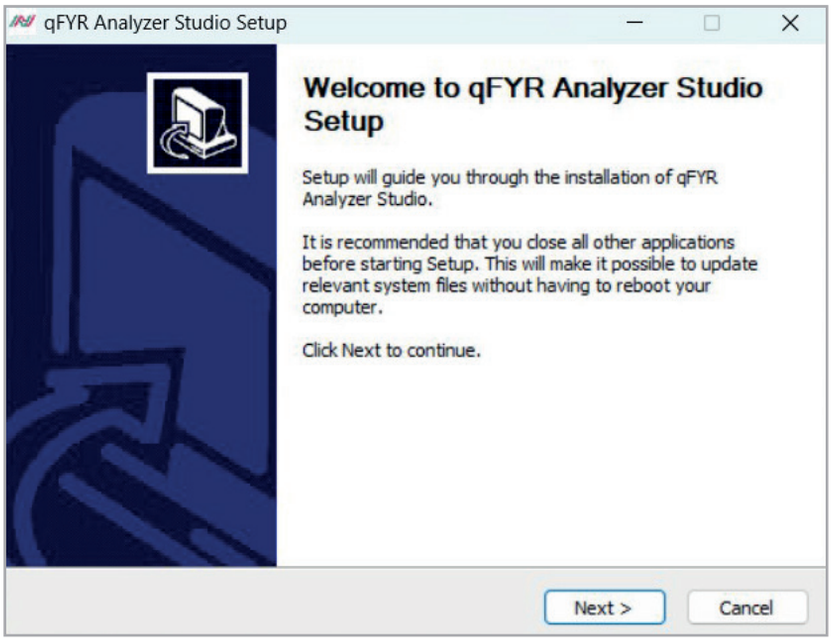
3.6.1. FastGene® qFYR Analysis Studio Software installation

You need to install the FastGene® qFYR Analysis Studio software on your computer before using the FastGene® qFYR Real-Time PCR Systems. You can find the installation software on the provided USB-drive.

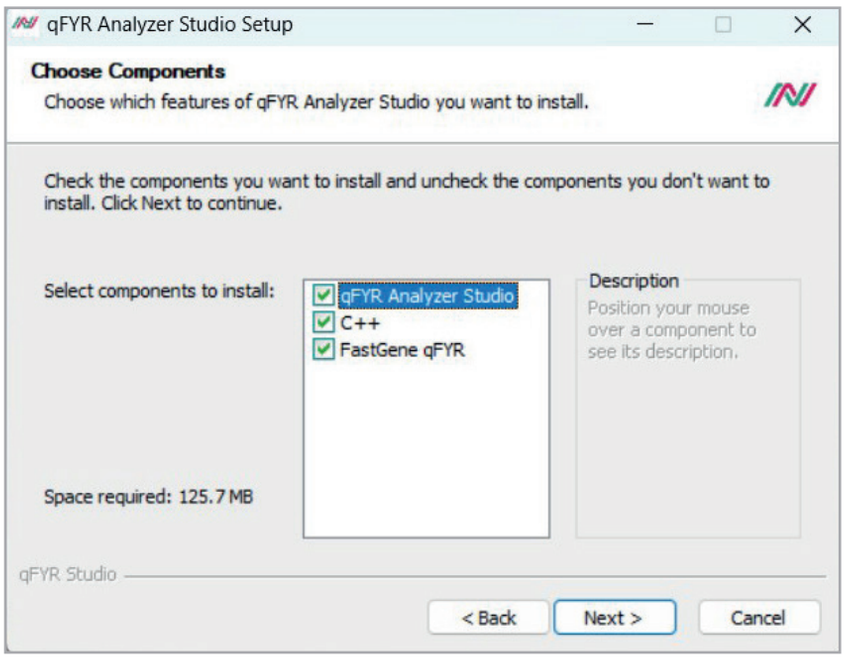
Please refer to the following steps to complete the installation process:

1. Click on the installion file.

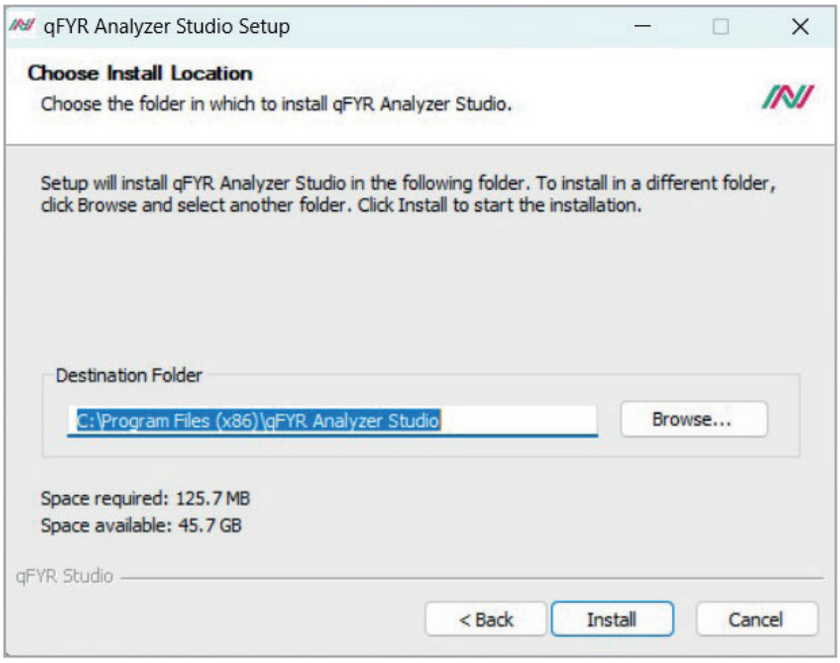
2. At the pop-up installation interface, click on next to continue.



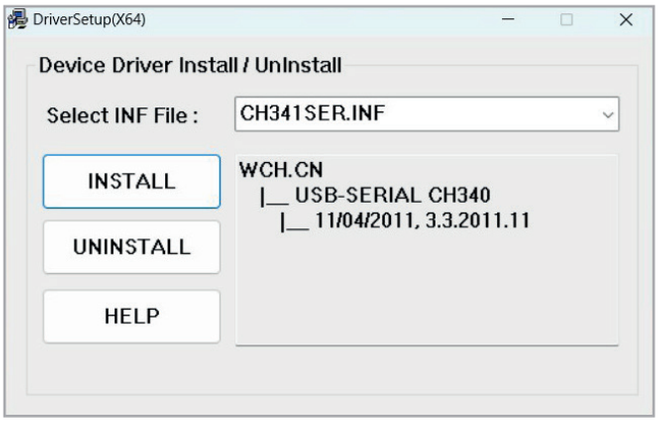
3. Select wich features of the FastGene® qFYR Analysis Studio software you want to install.



4. Choose the folder in which to install FastGene® qFYR Analysis Studio software

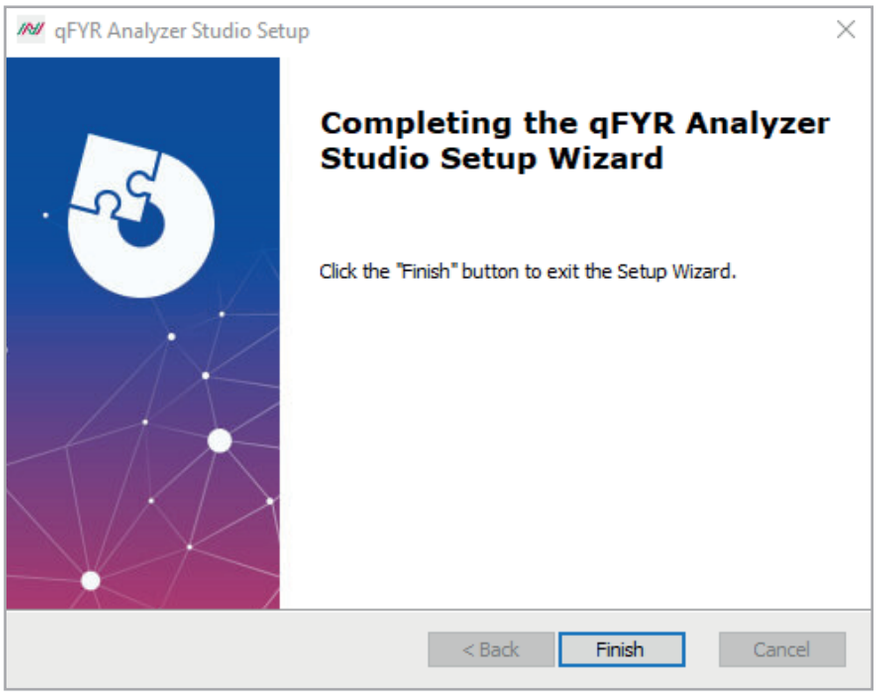


5. You need to pre-install drivers in the system before using FastGene® qFYR Real-Time PCR Systems and the FastGene® qFYR Analysis Studio software. After installation of the FastGene® qFYR Analysis Studio software, please click on the install button in the window below:



6. After installing, close the window.

7. Please click on Finish:



3.7. Pre-preparation for boot

3.7.1. Instrument self-inspection

The FastGene® qFYR Real-Time PCR Systems have the function of self-inspection. Before use, the user should run the instrument(s)´s self-inspection to ensure that the astGene® qFYR Real-Time PCR Systems can work normally during the experiment.

1. Turn the power switch on at the back of the astGene® qFYR Real-Time PCR Systems and turn on.
2. The astGene® qFYR Real-Time PCR Systems automatically enter the self-inspection mode: the automatic inspection includes the astGene® qFYR Real-Time PCR Systems‘ version, whether the electricity is in normal working condition, whether the power supply is normal or not, and the position of the initializing motor, etc.
3. After the self-inspection, the top light will constantly show a green color. The instrument(s) will be on standby until used.

Note: Make sure that astGene® qFYR Real-Time PCR Systems transport lock has been removed, the control computer and other external instruments have been correctly connected and that instrument(s)´s power supply has been connected correctly.

Note: In case of not passing the self-inspection, please turn the instrument(s) off and again on for self-inspection. If the instrument(s) still cannot pass self-inspection, please contact the support.

Do not shake the instrument(s).

3.7.2. System start-up

a) Start the computer.
b) Turn on the power switch of the backboard of the instrument(s), and wait until the power status light POW is blue and always on.



c) Wait for the system to self-check. During the self-check, the status light STU is green flashing state. After the successful self-check, green light will shine constantly.

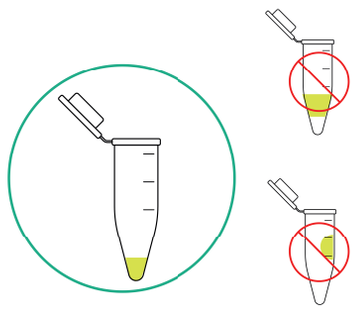
3.7.3. Experimental preparation

Good laboratory practice for PCR and RT-PCR

- ☑ When preparing PCR or RT-PCR amplification samples:
- ☑ Use only clean personal protection equipment and lab coat;
- ☑ Whenever you suspect that the gloves are contaminated, it is recommended to change them;
- ☑ Set up separate areas and special equipment and supplies for the following purposes:
  - sample preparation and reaction settings,
  - product amplification and analysis,
  - to avoid contamination, do not bring amplified products into the reaction setting area;
- ☑ Carefully open and cap all sample tubes to avoid splashing or spraying samples;
- ☑ Regularly clean the experimental table with proper disinfectants;
- ☑ Use calibrated pipettes and anti-aerosol tips;
- ☑ Prepare the reaction mixture according to the manufacturer's recommendations for the main mixture and the mixture to be tested;
- ☑ Include excess volume in the calculation to provide excess volume of master mixes for losses during reagent transfer;
- ☑ Dilute the sample and any standards with TE buffer or water;
- ☑ Be careful when diluting samples and standards, errors or inaccuracies in the dilution process directly affect the accuracy of the data;
- ☑ Place the diluent and assay mixture in the refrigerator away from light until they are ready for use. Overexposure may affect fluorescent probes or dyes;
- ☑ Before use:
  - thoroughly mix the master mix by rotating the bottle,
  - resuspend the assay mixture by vortexing and then centrifuge briefly,
  - thaw any frozen samples on ice. After thawing, resuspend the sample by vortexing, and then centrifuge the tube briefly.

Guide to set reaction in plate or tube

- ☑ Make sure that the liquid in each tube is at the bottom of the tube without bubbles. Otherwise centrifuge the well plate again;
- ☑ Ensure that the plate or tube is properly sealed;
- ☑ Keep the reaction plate or reaction tube in a dark environment at 4° C until it is ready to be loaded into the instrument;
- ☑ Keep the bottom of the plate clean. Liquid and other contaminants on the bottom of the plate will contaminate the sample and cause abnormally high background signals;
- ☑ If necessary, use permanent markers or pens to mark on the side of the test tube and plate, do not use highlighters.



4. Operation to run the experiment

This section will describe in detail how to set up and run experiments using FastGene® qFYR Analysis Studio software.

Note: The qFYR Analysis Studio software only accepts a dot as a decimal separator.

4.1. Software start

1. Double-click the desktop software icon to launch the software.

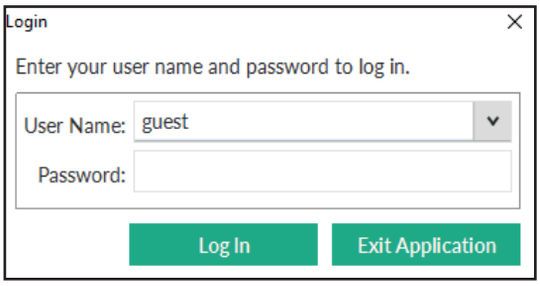


2. Interface of the software startup.



3. User login.

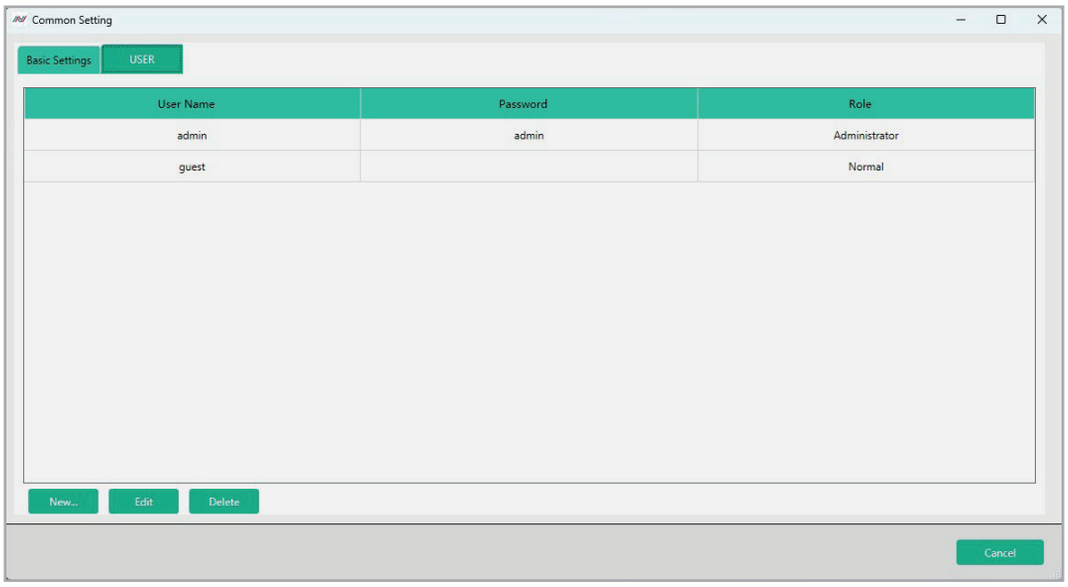
After the software starts, the system Login dialog box will automatically pop up (see below). Select the user by changing the “User Name”. If no user has been defined, you can proceed with “guest”. Click on the „Log In“ button to start the software.



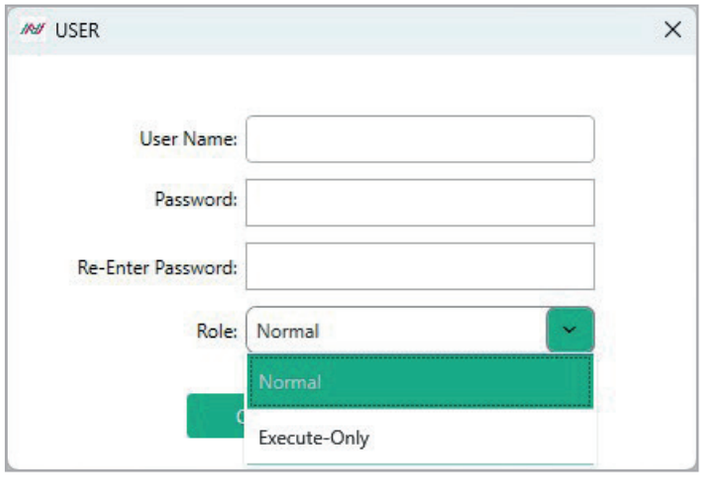
4. Adding a user.

Log in as Administrator. Select “admin” in the User Name and enter the Administrator password. The default password is ***admin*** (Please let the administrator of the instrument change the password as soon as possible).

Click on File in the top left corner and then click on settings. The next window (USER) will let you add a new user, edit or delete users.



5. When adding a new user, different roles can be defined and you can either choose a “normal” or an “execute-only” user role.



User	Functions allowed
Guest	No login with password required Create, save, open, import, and run template files Create and modify run settings View, modify, analyze and export data files
Normal	Login with password required Create, save, open, import, and run template files Create and modify run settings View, modify, analyze and export data files
Execute-only	Login with password required Open and run template files View and export data files
Administrator	Login with password required Same as normal user Manage, add, delete users

6. Startup interface will be shown:

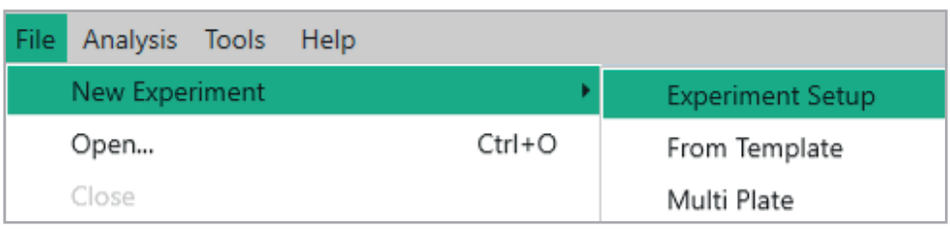


4.2. Create new experiment

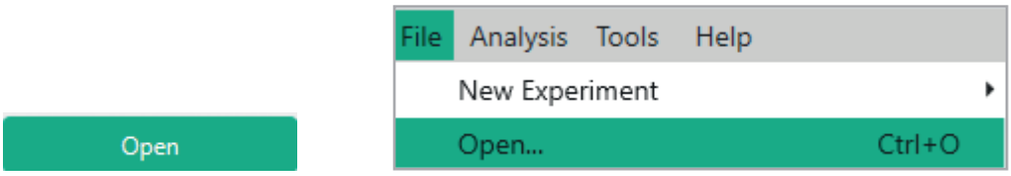
The Startup interface is used to create a new experiment, run, or open previously saved templates to create a new experiment.

4 2.1. New experiment or loading existing files





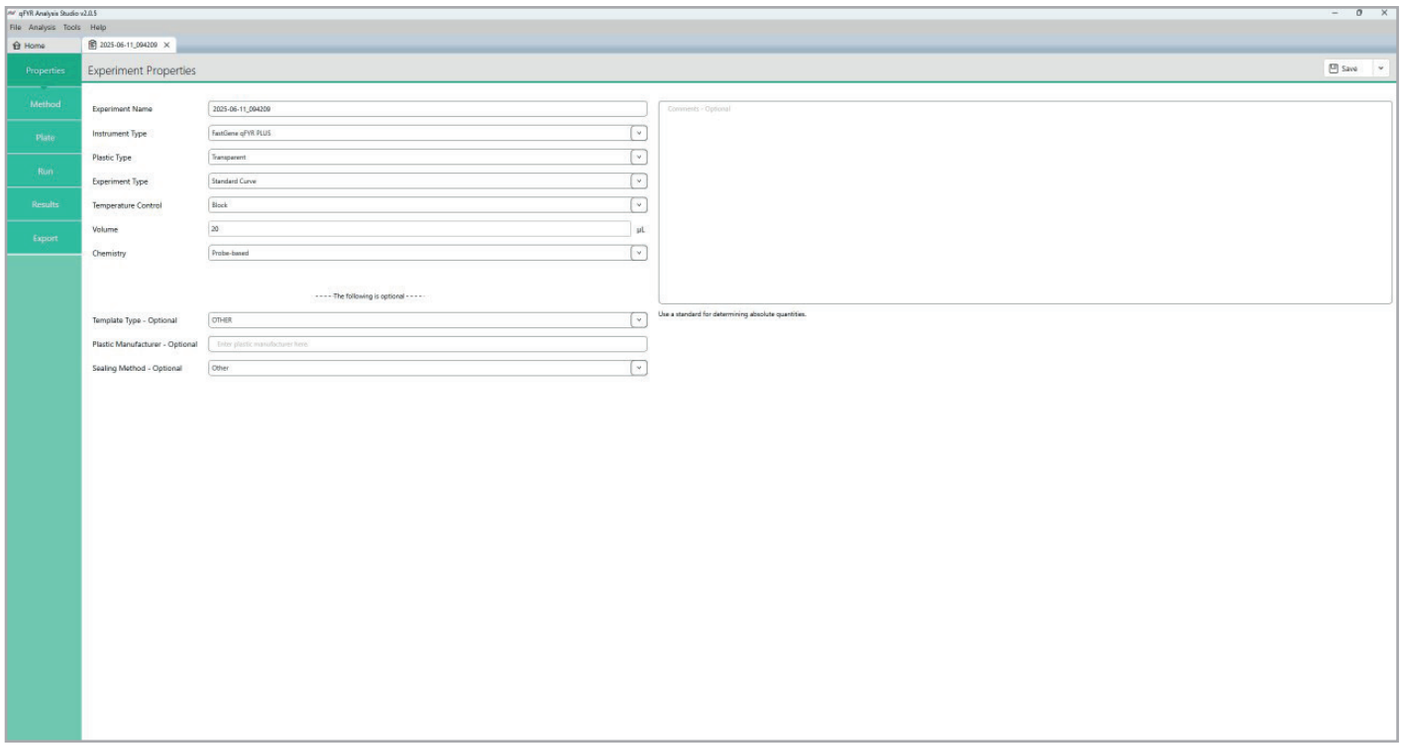
1. Click the „Experiment Setup“ button at the bottom right of the startup interface or select „New Experiment“ from the menu bar to start a new experiment setup.
2. Click „From template“ in the menu bar to create a new experiment by loading a previously saved experiment template.
3. Click the „Open“ button at the bottom right of the startup interface or select „Open“ from the menu bar to open previously saved experiment or template files.



The file types are as follows:
a) “*.qptx” file is a template file for experiment operation and contains information about operating conditions and reaction plate settings.
b) “*.qpsx” file is a complete raw data file, including operating conditions, reaction plate settings and experimental results.

4.3. Experiment properties

The „Experiment Properties“ interface is used to define the relevant properties of the experiment according to the actual experiment. This includes the name and type of the experiment, as well as the selection of the reaction tube and PCR reagent.

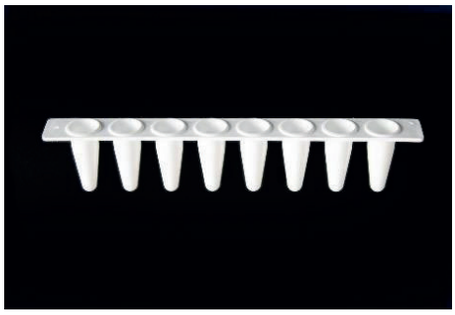


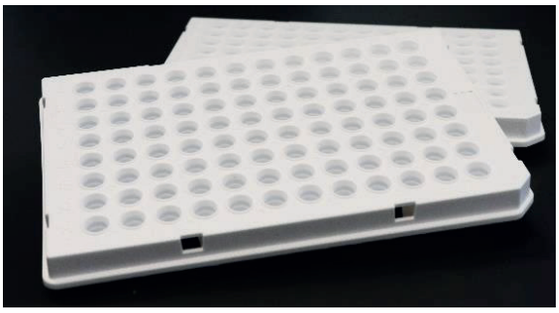
4.3.1. Name

The default setting for the experiment name is the current time and date, which can be changed as needed. Special characters are not supported.

4.3.2. Plastic type

The instrument is compatible with the common fluorescent quantitative reaction tube types, including 0.1 mL PCR single tube, 8-well strip, and 96-well plates. White tube or transparent tube (including frosted tubes) can be selected according to the reaction tube material. Reading of the wells will be adjusted according to the selected reaction tube type. Compared with transparent tubes, white tubes can reduce the loss of optical signal and crosstalk. Therefore, we recommend to use white opaque reaction tubes and plates to obtain better experimental data.





4.3.3. Experiment type

Select the experiment type according to the actual purpose of the experiment. The software will associate the subsequent experiment settings according to the selected experiment type. For the experiment type, please refer to the following table:

Type of experiment	Explanation
Standard Curve (Absolute quantification)	Absolute quantification based on a standard curve is a real-time quantitative PCR analysis method suitable for researchers who need to determine the actual concentration or copy number of target genes. Forabsolute quantification, a template solution with a known concentration needs to be serially diluted several times. The obtained data is used to generate a standard curve. The standard curve is plotted with each concentration and corresponding Cq value. The Cq value of the unknown sample is then compared with this standard curve to determine its quantity.

Type of experiment	Explanation
Relative Standard Curve or Comparative Cq (ΔΔCq) (Relative quantification)	Relative quantification is suitable for most gene expression studies. It can analyze the up-regulation or down-regulation of target gene expression levels in control samples and one or more experimental samples. With this technique, there is no need to accurately determine the copy number, but to focus on the fold change compared to the control. The ΔΔCq method is a very common method of relative quantitative analysis. It compares the results of experimental samples including controls (such as untreated or wild-type samples) and normalization targets (such as housekeeping gene expression). Using this method, the Cq of the target gene in the same two samples can be compared based on the Cq of the normalization gene in the test sample and the control sample. The resulting ΔΔCq value can be used to determine the fold difference in expression.
Melt curve or High Resolution Melt	The melt curve experiment is used to determine the melting temperature (Tm) of the PCR product detected by an intercalating dye, such as SYBR™ Green dye. Gene sequences with different base compositions or lengths have different Tm, so different reaction products can be determined according to the Tm value, including non-specific product. During a high-resolution melt analysis, the change in fluorescence is detected at very small temperature increase steps, allowing the identification of SNPs and mutations. For the detection of unknown or new mutations, saturated DNA combined with dyes is usually used to genotyping by high resolution melting curve.
Genotyping	Real-time fluorescence quantitative PCR can provide different methods for genotyping analysis. For the detection of known mutations, the most commonly used basic method is the end point genotyping using enzyme cleavage hydrolysis probe, and the more advanced method is the fusion curve genotyping using hybrid probe. The method can be easily improved and applied to the detection of various other SNP.
Presence or absence	Protocol using a probe-based assay to control the presence or the absence of a target.

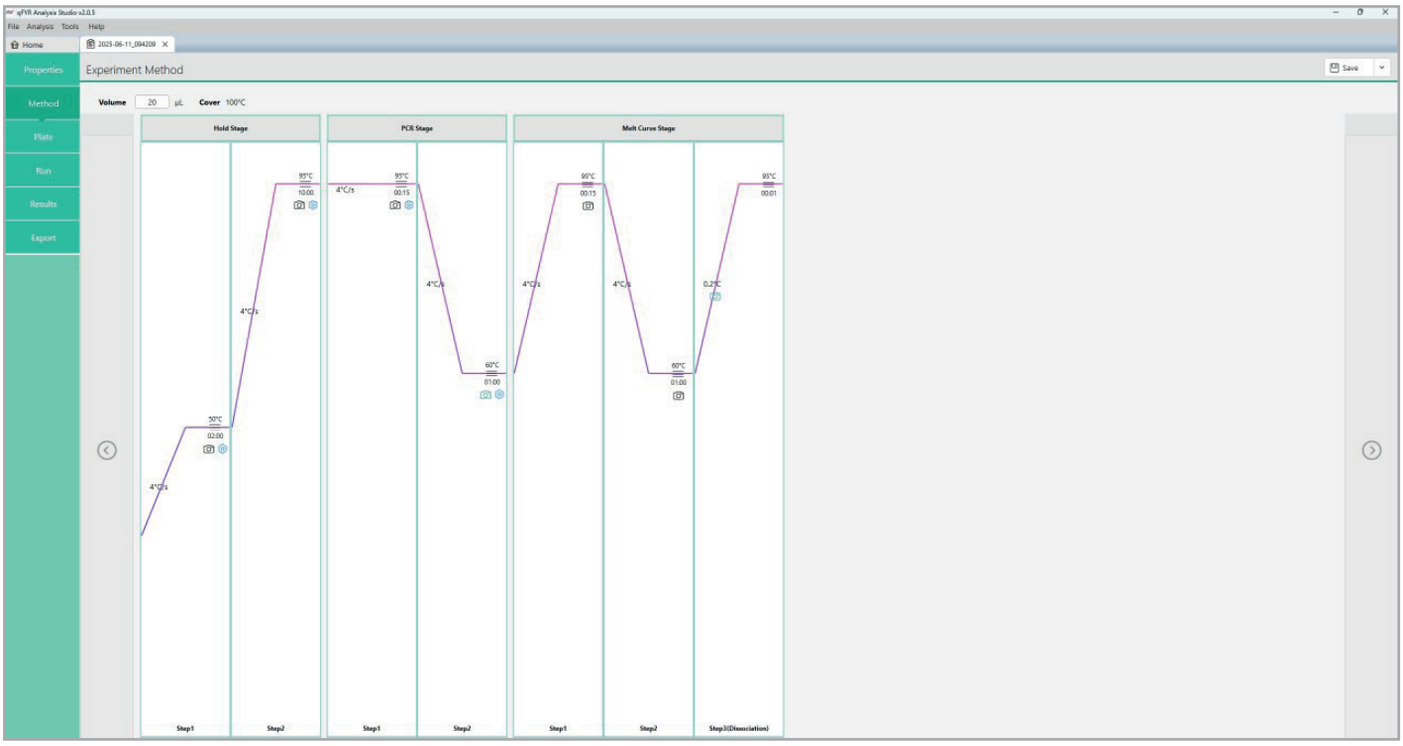
4.3.4. Chemistry

The instrument is compatible with most of the fluorescent quantitative PCR reagent consumables in the market. You can choose the probe method (TaqMan™ reagent) or the dye method (SYBR™ Green reagent) to carry out subsequent experiments according to the specific experimental design. For the comparison of the probe method and the dye method, please refer to the following table:

Methods	Advantages	Disadvantages	Scope of application
Dye-based method	• Widely used • Sensitive • Convenience • Cheap	• Difficult-todesign primers • Non-specific binding	Suitable for quantitative analysis of various target genes in scientific research, research of gene expression, research of transgenic animals
Probe-based method	• High specificity • Good repeatability	• Expensive • Only suitable for specific targets	Pathogen detection, disease resistance gene research, drug efficacy assessment, diagnosis of genetic diseases

4.4. Experiment method

The „Method“ interface is used to set the specific parameters of a quantitative PCR, including reaction volume, stage, step, temperature, time, and cycle number, etc. The above parameters can be carried out according to specific experimental needs and corresponding reaction reagent requirements.



4.4.1. Reaction volume

The default volume is 20 µL. Volumes ranging from 5 to 100 µL are supported.



4.4.2. Thermal lid temperature

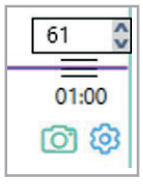
Thermal lid temperature is fixed to 100°C and cannot be changed.



4.4.3. Basic setting

The values of time, temperature, heating rate (melting curve mode) can be set by entering a value in the value area or clicking the arrows . Temperature can additionally be changed by clicking and dragging in the temperature curve.

Note: *The minimum value of the time setting is 00:01 minutes.*

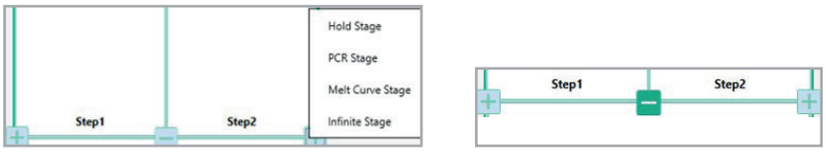


The temperature is selected.

2. Light up the camera icon to select the data collection point.
3. The camera is turned on by default to collect data after the extension is completed.
4. If you need to add a reaction step in a certain stage of PCR, hover the mouse to the bottom left or right step area and click „+“ to add a step before or after the current step. The „-“ button removes the selected step.



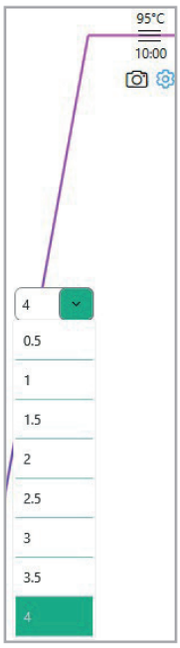
5. If you need to add a reaction stage of the PCR, hover the mouse to the bottom left or right stage area and click „+“ to add a stage before or after the current stage and select the type of stage (Hold, PCR, Melt Curve or Infinite) that should be added. The „-“ button removes the selected stage.



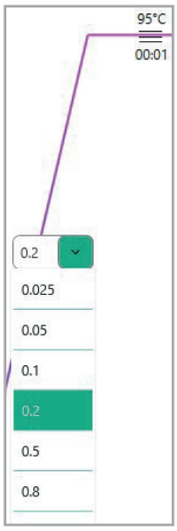
6. The number of cycles can be changed by entering a value in the value field or clicking the arrows. The standard number of cycles is 40.



7. The ramp rate can be set in the drop-down list at the beginning of each step. Ramp rate is given at temperature change within the sample volume and can be set between 0.5 and 4 °C/s.

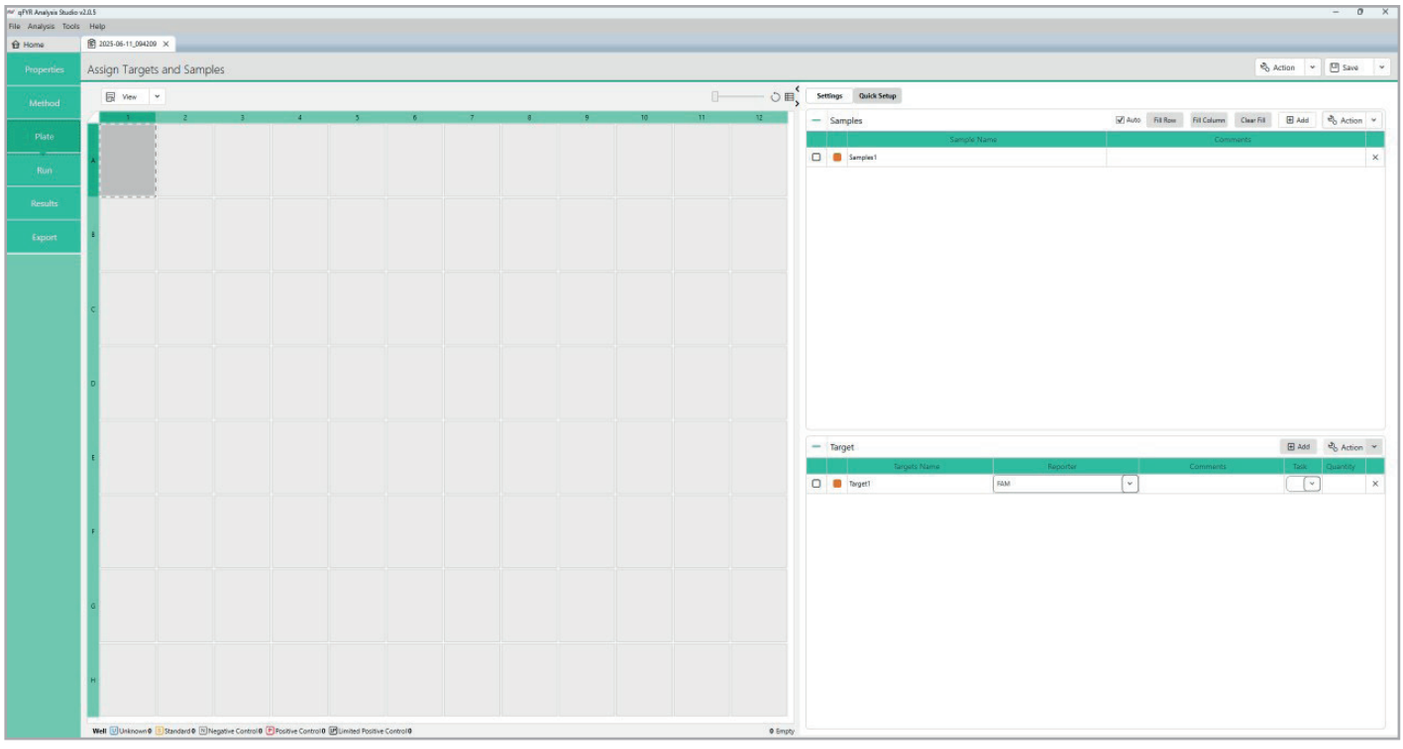


8. The temperature interval can be set in the drop-down list on the camera icon during a melt curve analysis in which the fluorescence signal is measured. The smaller the value, the more data points are collected. Standard value is 0.2 °C/detection step.



4.5. Plate Setup

The “Plate” interface is used to set the properties of the reaction wells, including assigning samples, test targets, reaction well tasks and other properties to each reaction well. This will make the identification of the sample and the subsequent analysis possible.

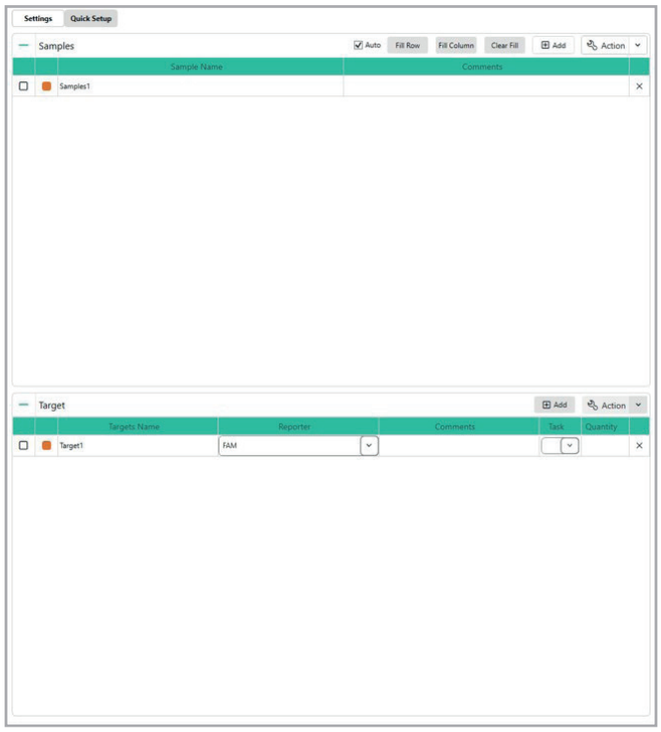


Note: The plate setup can be performed before, during and after the experiment.

4.5.1. Setup

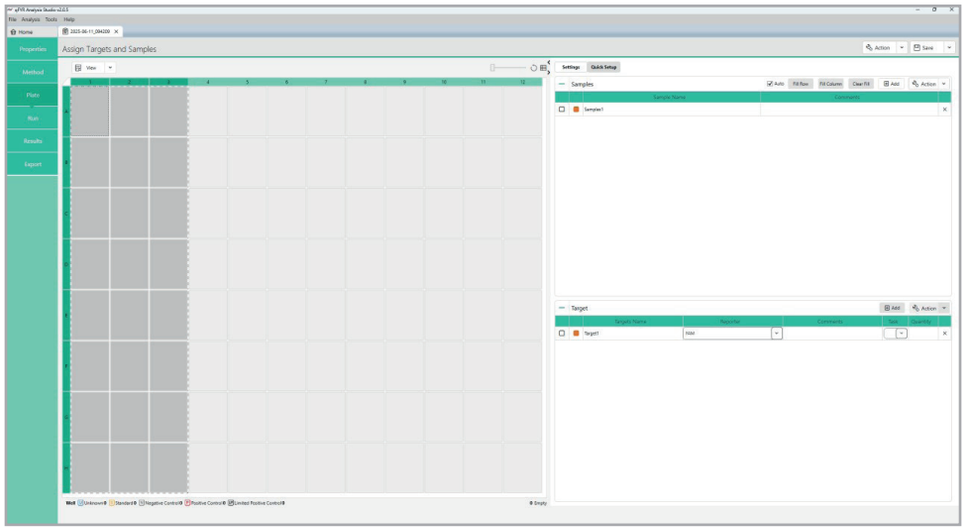
Define and assign well properties in the “Setup” window of the plate setup:

1. Click „Add“, to add samples and target items.



2. Click × on the right to remove sample and target items.

3. In (well plate view), select the reaction well according to the corresponding well position of the actual reaction tube.



4. Assign the sample (nucleic acid source) and the target item (gene of interest) to the selected well position and click the text box of each field to name the sample and target gene for actual test.

5. When you enter a new sample or target item in the “Setup” window, the software will automatically fill the Reporter with default values (FAM/SYBR). Change the dye or probe used accordingly. After the setting is completed, check the corresponding samples and target to assign the set attributes to the corresponding well positions selected previously.
6. Assign tasks in the window of the “Plate Setup” tab:



7. Select the reaction well in ⊞ (well plate view).

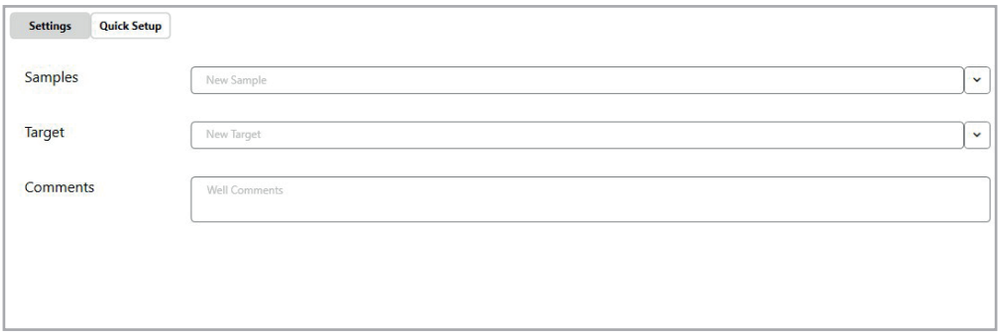
8. In the inspection item, select the check box of the inspection item, and then select the task from the drop-down list.

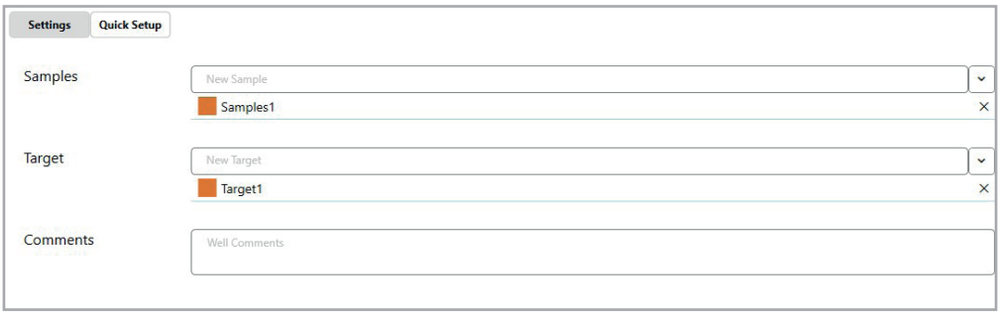
Reaction type	Duty
Unknown (sample to be tested)	U
Negative control	N
Positive control	P
Standard	S
Positive Control Allele 1/Allele 1	1 1
Positive Control Allele 2/Allele 2	2 2
Positive Control Allele 1/Allele 2	1 2
Positive Control - IPC	I
Blocked IPC control	

After the above settings are completed, assign the corresponding samples and targets items to the corresponding selected wells.

4.5.2. Quick Setup

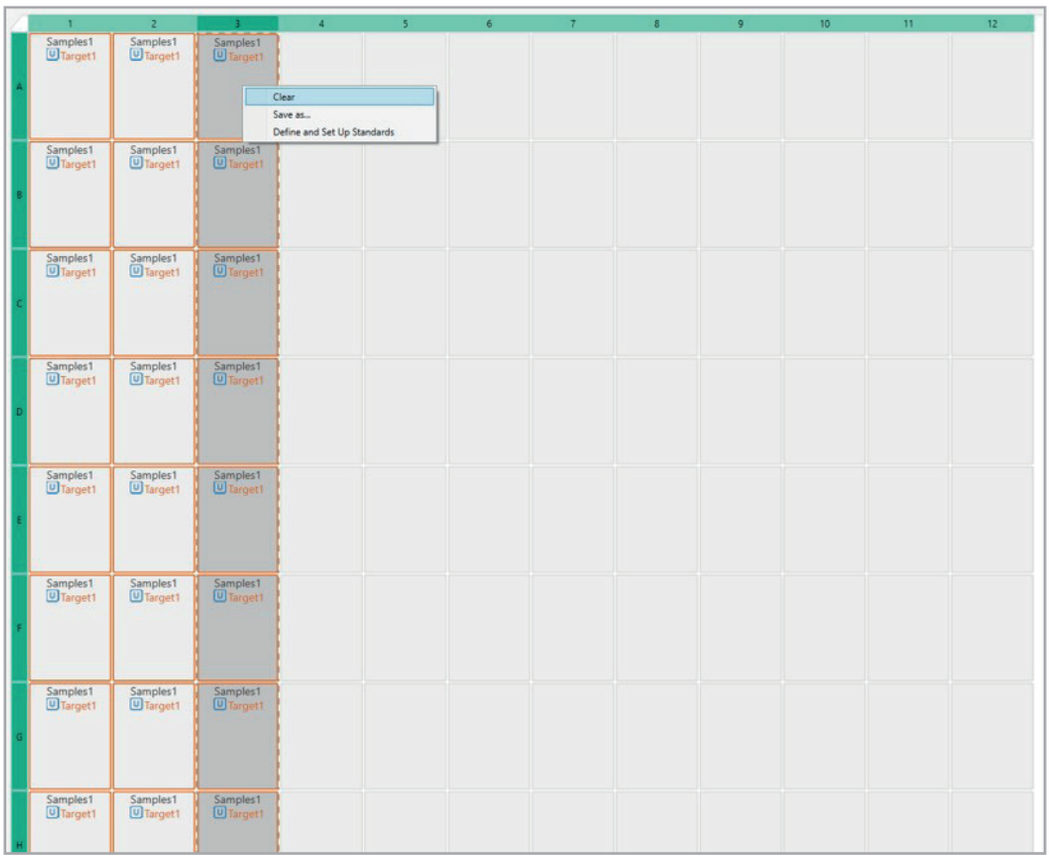
„Quick Setup“ is a function module for quick and easy setup of samples and targets. After selecting the well position to be set, you can click the sample text box and target text box to manually enter the name. If settings have been made in the “Setup”, you can select the options defined in the “Setup” through the drop-down menu to assign attributes to the corresponding well positions.





4.5.3. Remove Wells from the Reaction

Select the wells to be removed, click the right mouse button, and select “Clear”. For multiple wells, drag select these wells, then click the right mouse button to select “Clear”.



4.5.4. Data Import

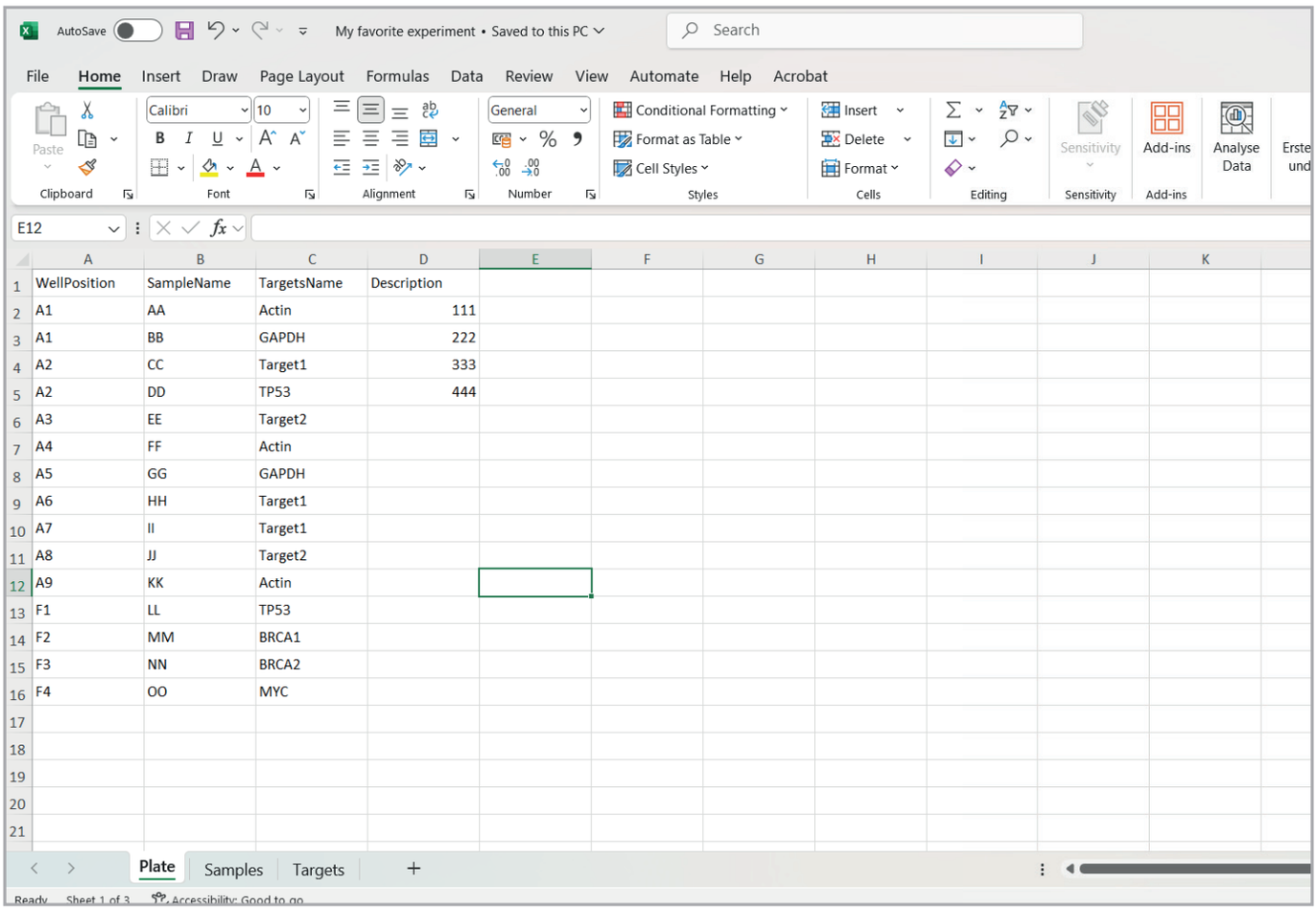
You can import a full plate layout into the qFYR Analysis Studio.
The plate layout file must be Excel file format.
Open Excel and create a file with the following sheets: “Plate”, “Samples” and “Targets”.

1. Plate Sheet

In the „Plate“ sheet, label the columns as follows:

Column	Header
A	WellPosition
B	SampleName
C	TargetName
D	Description

Enter your experiment information accordingly.
For multiplex experiments, you can assign multiple samples to the same well by repeating the well position in multiple rows (see Figure XXX for wells A1 and A2).



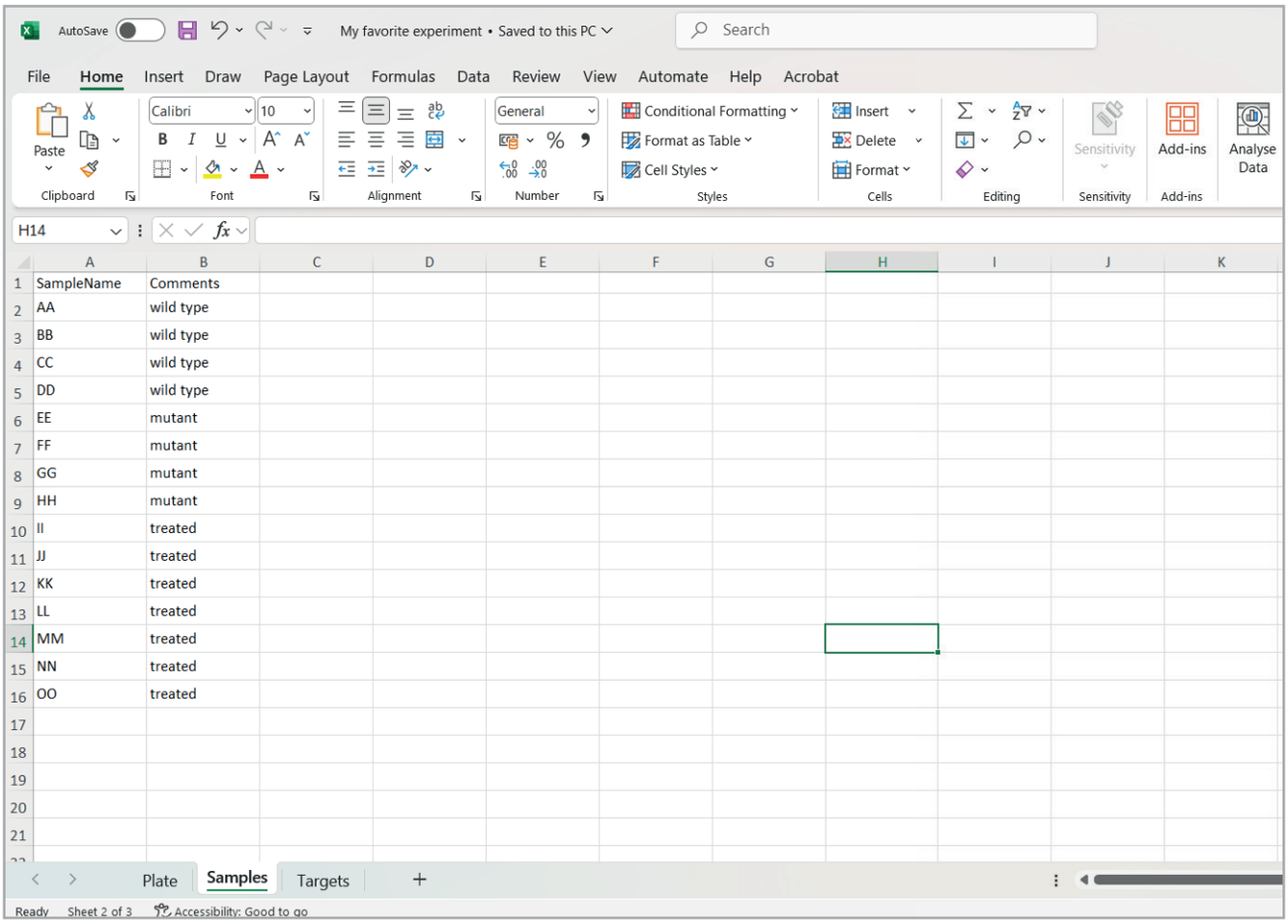
	A	B	C	D	E	F	G	H	I	J	K
1	WellPosition	SampleName	TargetsName	Description							
2	A1	AA	Actin	111							
3	A1	BB	GAPDH	222							
4	A2	CC	Target1	333							
5	A2	DD	TP53	444							
6	A3	EE	Target2								
7	A4	FF	Actin								
8	A5	GG	GAPDH								
9	A6	HH	Target1								
10	A7	II	Target1								
11	A8	JJ	Target2								
12	A9	KK	Actin								
13	F1	LL	TP53								
14	F2	MM	BRCA1								
15	F3	NN	BRCA2								
16	F4	OO	MYC								
17											
18											
19											
20											
21											

2. Samples Sheet

In the „Samples“ sheet, label the columns as follows:

Column	Header
A	SampleName
B	Comments

Enter relevant sample details in the respective columns.



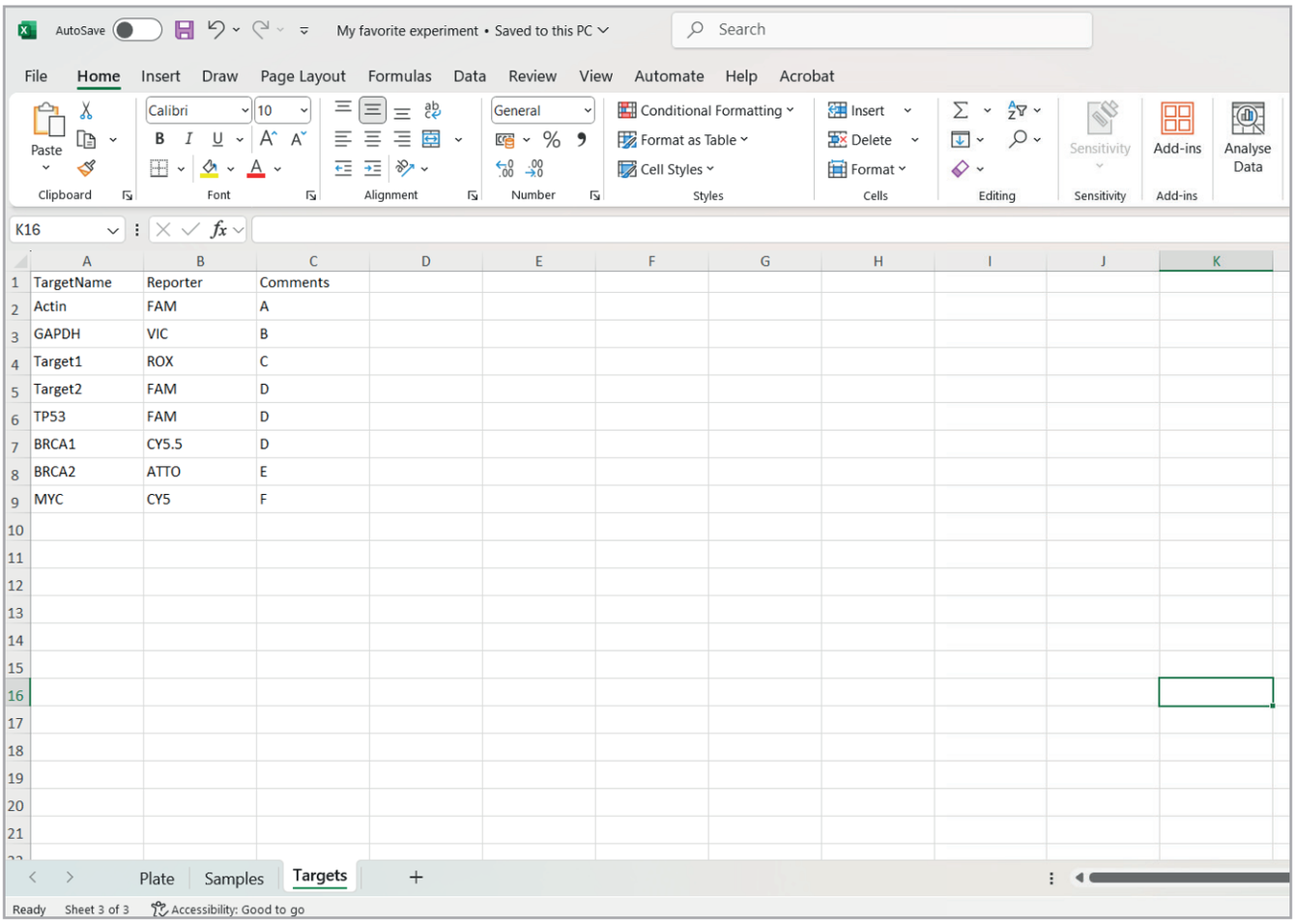
	A	B	C	D	E	F	G	H	I	J	K
1	SampleName	Comments									
2	AA	wild type									
3	BB	wild type									
4	CC	wild type									
5	DD	wild type									
6	EE	mutant									
7	FF	mutant									
8	GG	mutant									
9	HH	mutant									
10	II	treated									
11	JJ	treated									
12	KK	treated									
13	LL	treated									
14	MM	treated									
15	NN	treated									
16	OO	treated									
17											
18											
19											
20											
21											

3. Targets Sheet

In the „Targets“ sheet, label the columns as follows:

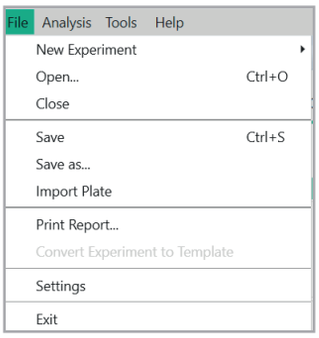
Column	Header
A	TargetName
B	Reporter
C	Comments

Input the target information as needed.

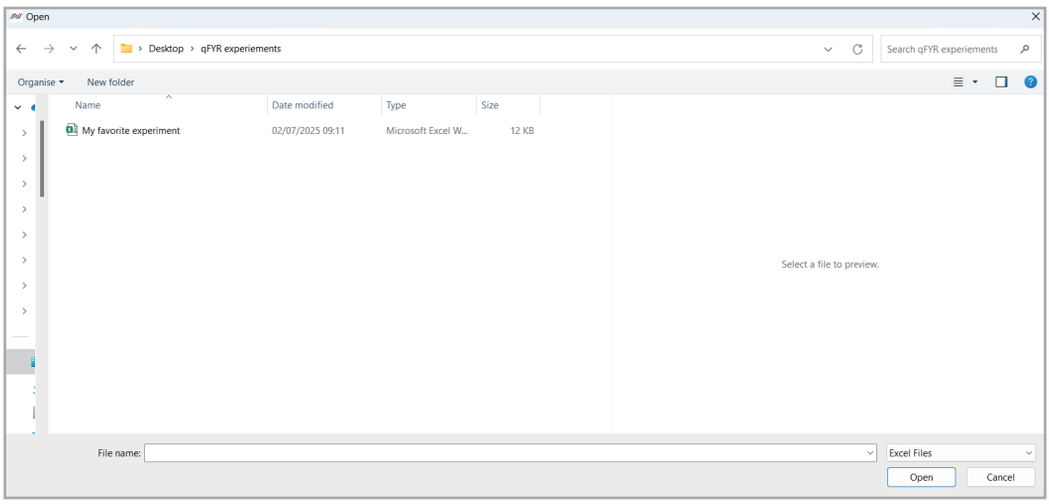


Save the Excel layout file.

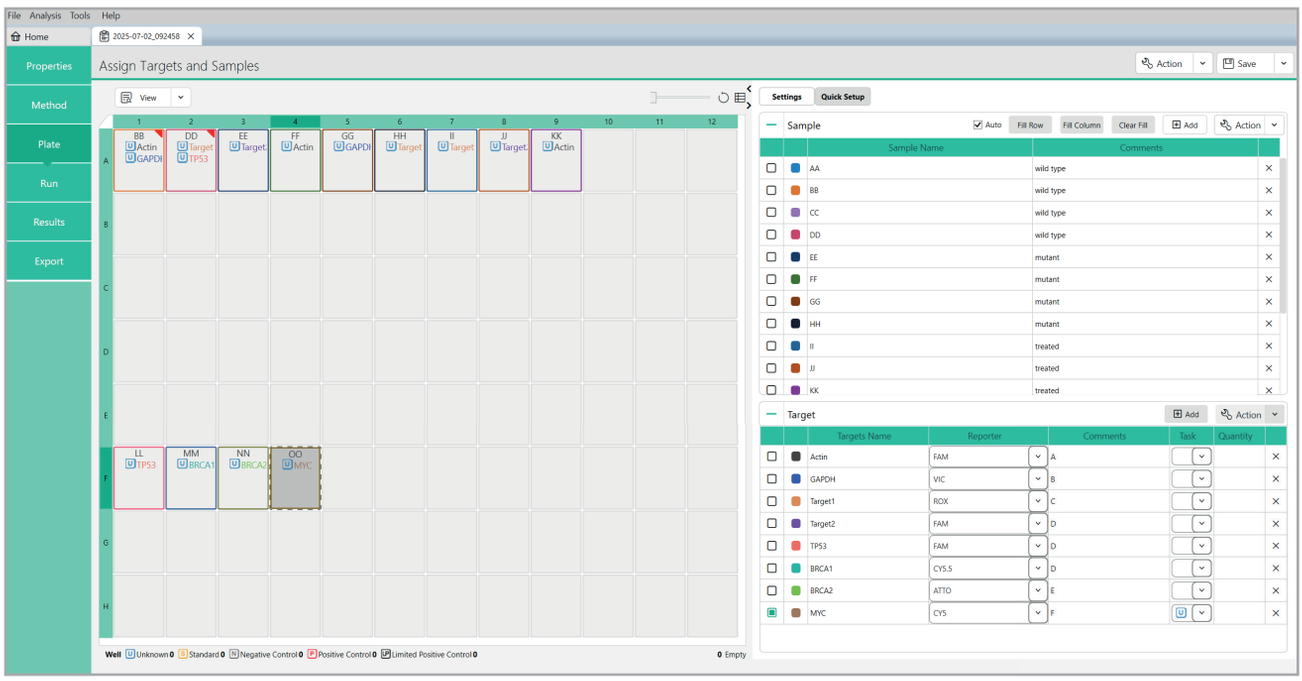
In the qFYR Analysis Studio go to “File” and select “Import Plate”



Select your experiment layout file and click “open”.



The software will load the file and display it as a complete experiment.



Note: Wells A1 and A2 are assigned as multiplex and thus marked with a red sign.

4.6. Run an Experiment

4.6.1. Insert a Plate or Reaction tubes

1. Press the access button on the top of the instrument.
2. The instruments heating block will be ejected.
3. Load the reaction tube or plate onto the 96-well block.

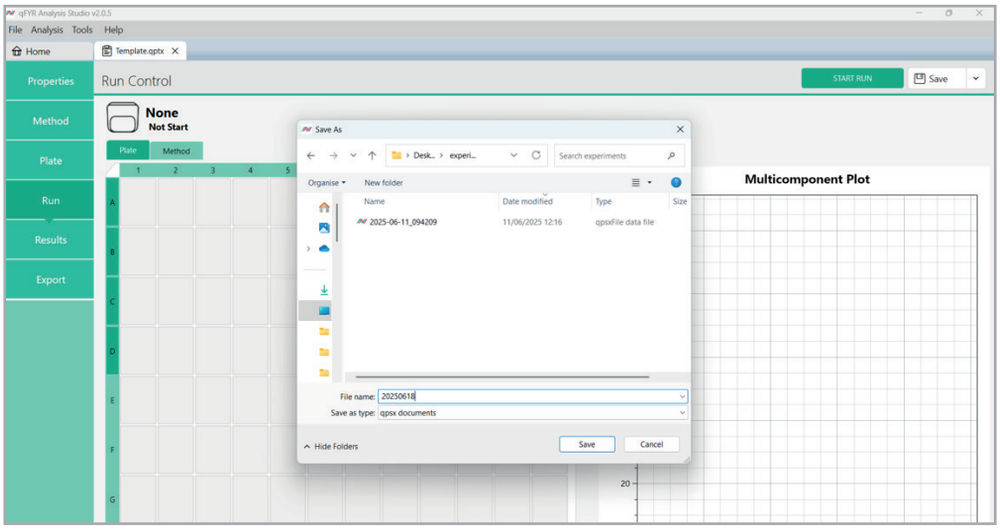
Note: When the drawer is closed, the heating lid will apply the appropriate pressure to securely place the tube band in the module.

4. Press the access button again to close the heating block.
5. After the operation is completed, press the access button to eject the heating block, remove the reaction tube or plate from the heating block, and then close the door by pressing the access button.

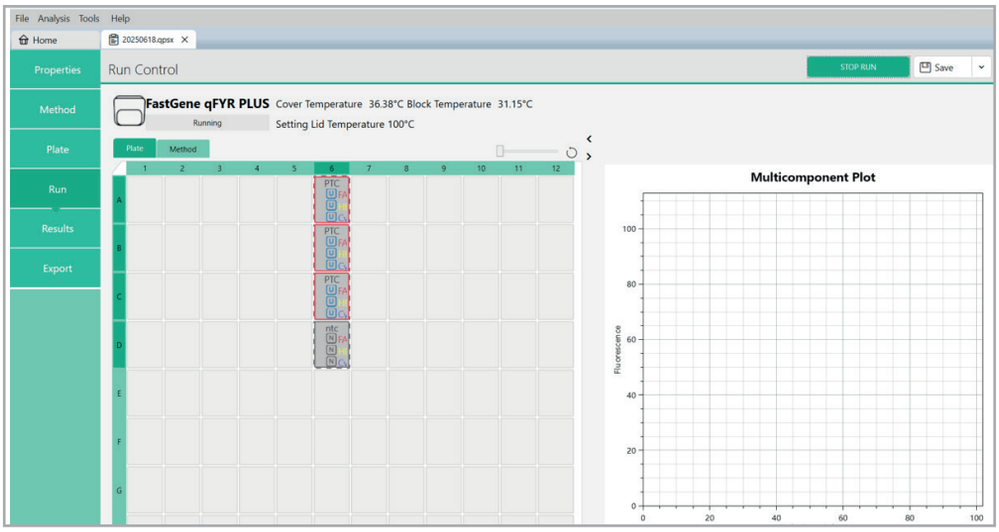
Watch out! Risk of personal injury. During the operation of the instrument, the heating block can reach a temperature of 100 °C. Allow it to cool down to room temperature before processing.

4.6.2. Run the Experimental Program

1. In the „Run“ tab of the software, click „Start Run“. After clicking „Start Run“, a save dialog box appears, select the data file storage location and name.



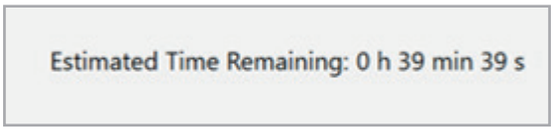
2. After saving, the instrument starts to run the program, and the real-time heating lid temperature and reaction plate temperature will be displayed in the upper left corner of the running interface.

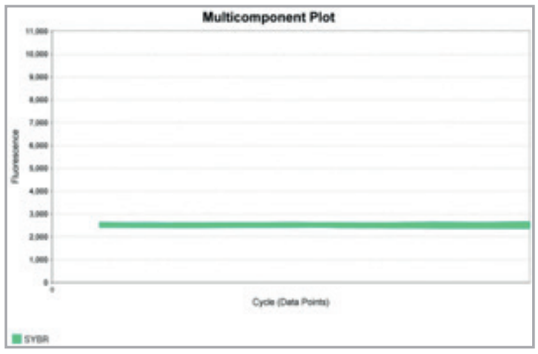




3. When the temperature of the heating lid reaches 100 °C, the real-time heating lids and reaction plates temperature, and the cycle number of the reaction will be displayed in the upper left corner of the running interface.



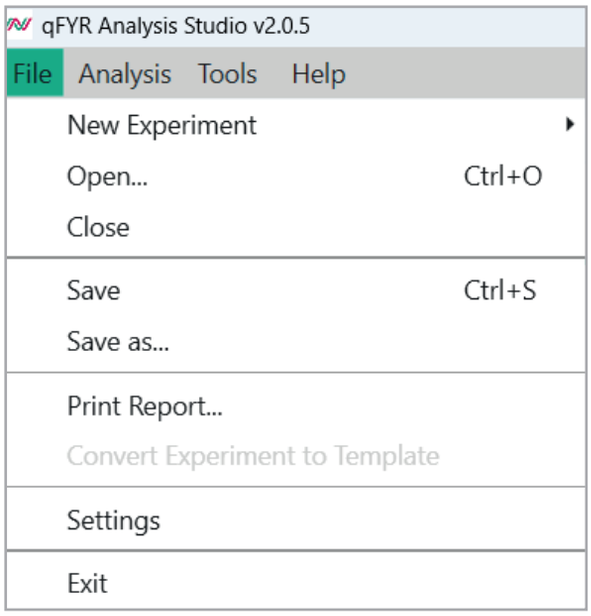


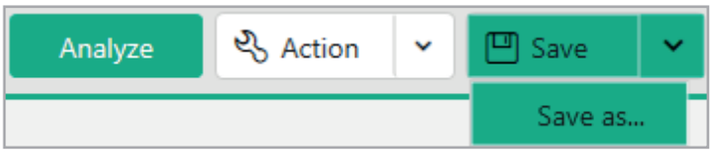


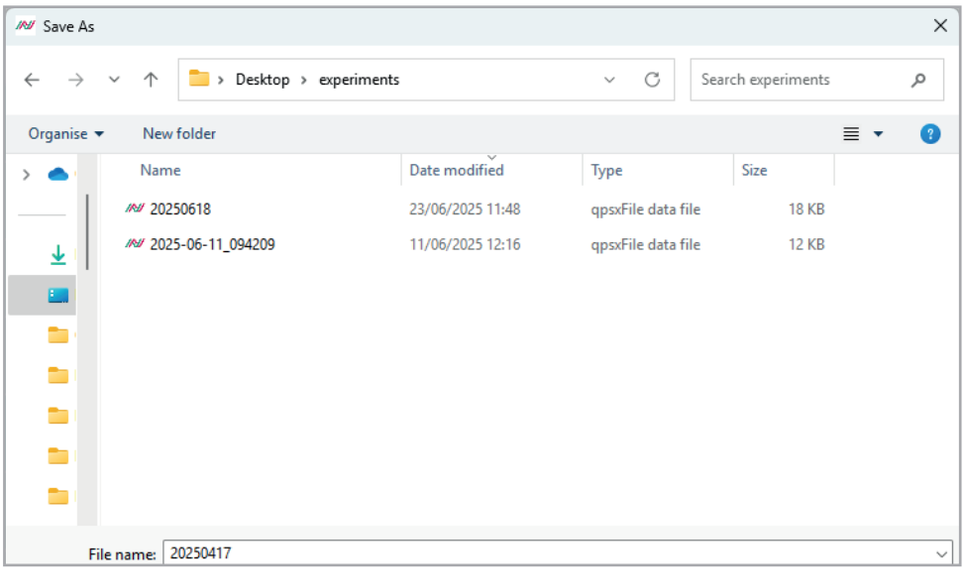
Note: *In the initial reaction cycles, the original data map is most of the background or baseline. When the amplification reaction enters the exponential growth phase, the curve will rise.*

4.7. Save Experiment

Click the „File“ option in the menu bar, choose “Save” or “Save as” to save the original experimental data. At each step of the experiment process, you can choose to save the experiment in the upper right corner of the interface. The extension of the experiment file saved before running is .qptx (experiment running template). The extension of the experiment file saved after the operation is completed, is .qpsx (original data file), which contains all the original operation data and analysis results.







5. Operation of different experiment types

FastGene® qFYR (Plus) Real-Time PCR System experiments are divided into seven steps:

1. Create an experiment
2. Set Experimental Properties
3. Set the operating conditions
4. Set the samples and targets
5. Start the experiment
6. View and analyze the results
7. Export the experimental results

This next chapter explains how to operate the different types of experiments within the software.

5.1. Absolute Quantification (Standard Curve) Experiment

Absolute quantification is a real-time fluorescence quantitative PCR analysis method suitable for researchers who need to determine the actual copy number of targets. To carry out absolute quantification, the target template solution with known concentration should be diluted several times continuously, amplified by real-time fluorescence quantitative PCR, and a standard curve is generated by using the obtained data. A standard curve is drawn with the concentration of each target and the corresponding Cq value. Cq values of unknown samples are then compared with this standard curve to determine their concentration or copy number.

The standard curve method is used for absolute quantification, and the template standard with known concentration or quantity is required, which means that the template should be accurately quantified before the curve is generated. Quantitative accuracy is directly related to the quality of the standard curve. The template used to generate the standard curve and the method used to quantify the template are the basis of the experiment. The accuracy of continuous dilution is very important, and the RT and PCR efficiency of target template and continuous dilution of actual samples are very important.

Although you may need uniform pure templates for initial copy number determination and standard curve generation, it is best to use target templates as similar as possible to experimental samples. Since steps such as nucleic acid extraction and reverse transcription can affect the dynamic range of the reaction, the same processing steps as the experimental samples should be adopted.

In the standard curve experiment:

- Software generates standard curves using data from standard dilution series.
- Software uses standard curves to calculate the number of unknown samples.

Type of response:

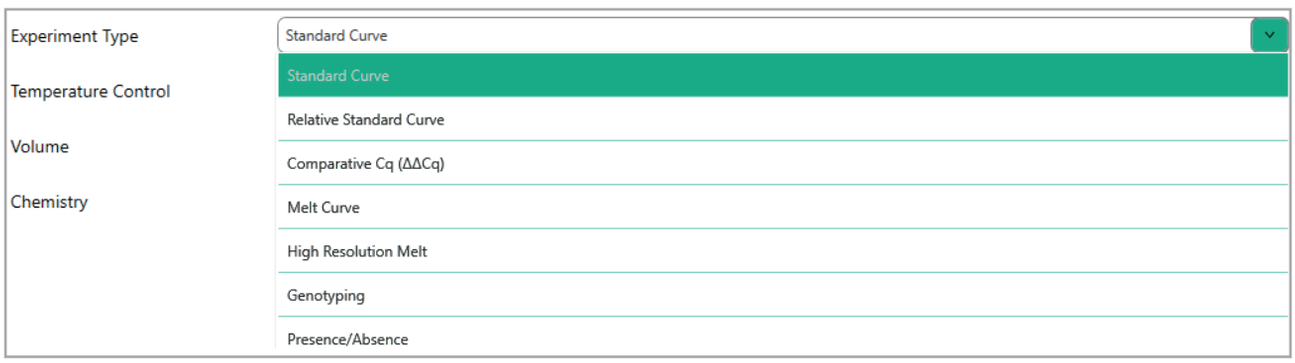
- Many samples can be analyzed in standard curve experiments, but each sample needs its own standard curve.
- The following table lists the types of sample settings involved in the standard curve experiment:

Type Setting	Example Desription
Standard	contains a known number of tergat samples
Unknown	sample to be tested
No template control (NTC)	water or buffer replacement template, no target amplification should occur in NTC well

The accuracy of quantitative experiments increases with the increase of repeated reactions (usually containing at least three repeats). For accurate detection, at least five dilution points in a wide range of standard quantities should be prepared. The range of the standard quantity can be adjusted according to the expected level of presence of the gene.

5.1.1. Setup of a Standard curve

1. Select experiment type “Standard Curve”.



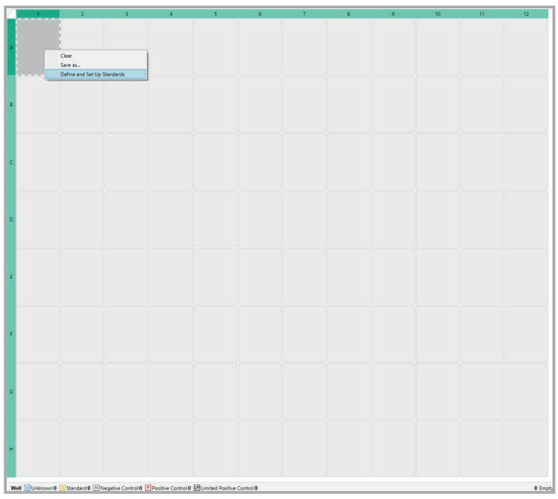
2. Choose the used Plastic type and Chemistry.
3. Add name and optional information.

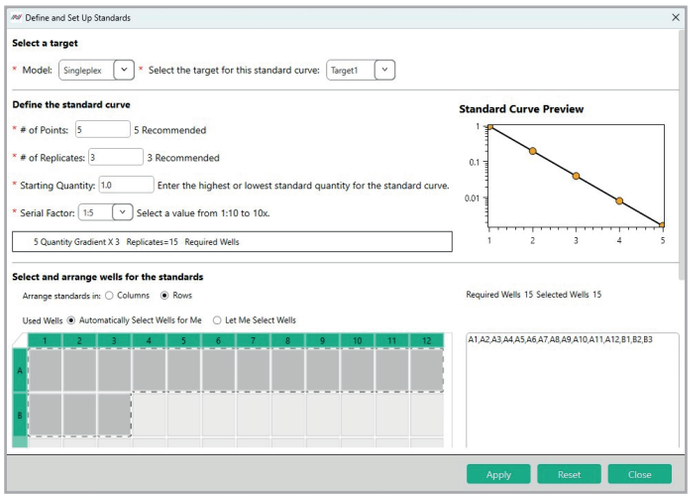


4. After the Method setting is completed, enter the Plate section to enter the well plate setting interface.

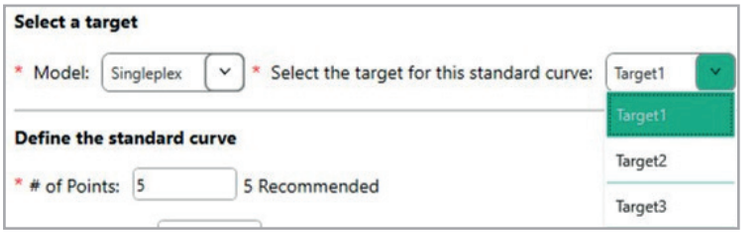
Note: *The plate setup can be set before, during, or after the reaction operation.*

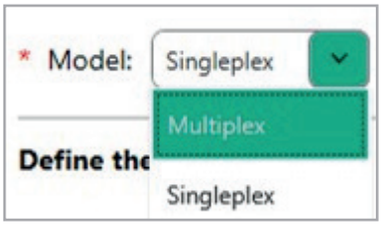
5. Right-click in any area of the plate view and select “Define and Set Up standards”.





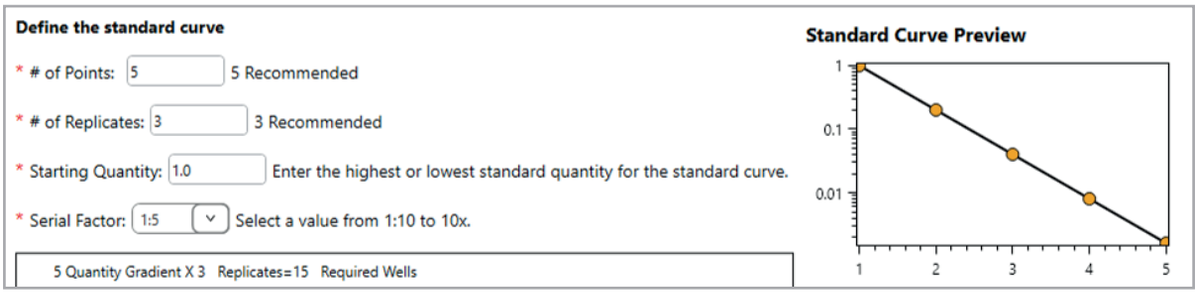
6. Select the target, which should be used for generation of the standard curve.





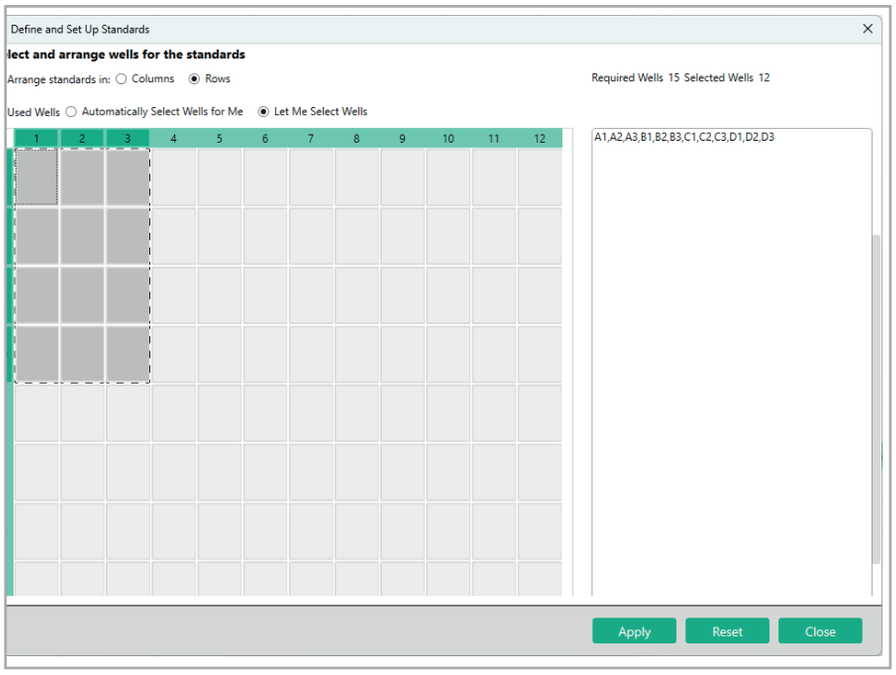
Note: *Singleplex means that there is only one gene to be tested in each reaction well, and Multiplex means that there are two or more genes to be tested in each well.*

7. Set the number of points and replicates, dilution factor and starting quantity according to your experiment.



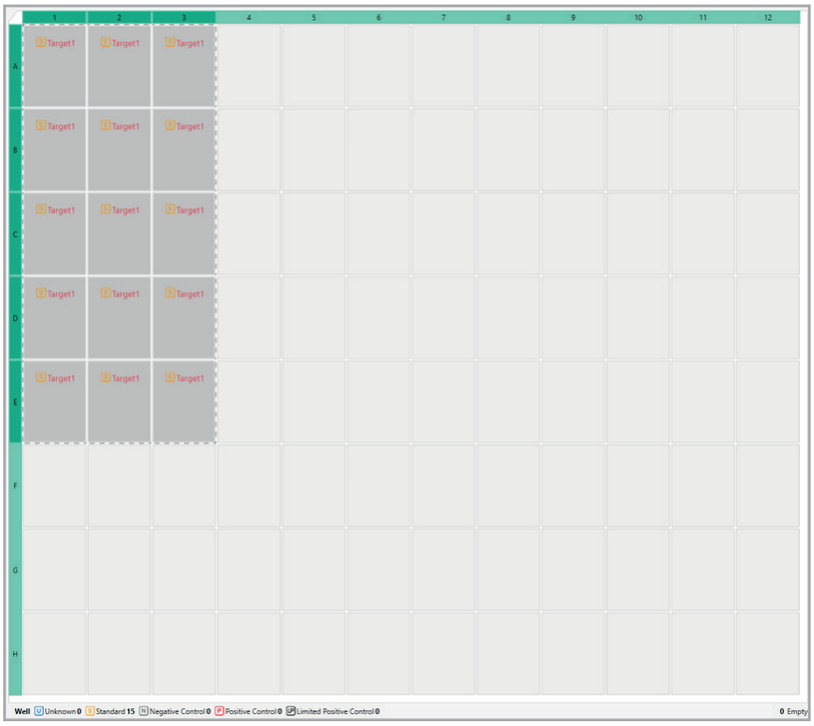
The parameters of the standard curve can be set according to the specific experimental requirements and conditions:

- Standard concentration dilution gradient: recommended at least five different dilutions.
- Three replicates per dilution are recommended.
- Starting Quantity: Enter the maximum or minimum quantity value of the standard, excluding units (units containing copies, copies/µL or ng/µL) and the dilution factor.



- The software can automatically select the reaction well by columns or rows or the operator can select their own reaction well according to the need and arrange the well position of the standard product.
- After the standard well position distribution is set, the software will arrange the standard well position according to the above setting, and display the corresponding well position arrangement and the number of wells.

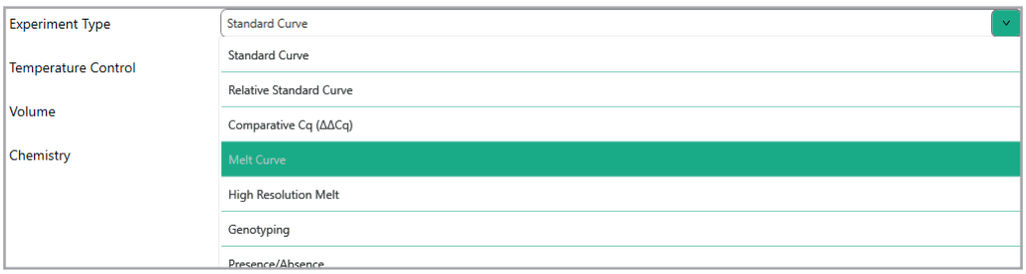
9. The software will automatically apply the above settings to the plate setup, and automatically define the above well position as the standard property (as shown below), so that the plate setup of the standard is completed.



5.2. Experiment of Melting Curve

5.2.1. Setup of Melt Curve Analysis

1. Select experiment type “Melt Curve”.



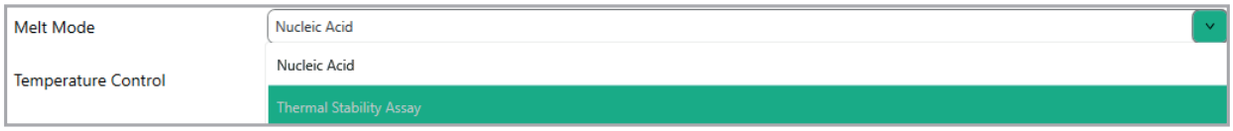
2. Choose the used Plastic type and Chemistry.

3. Add name and optional information.

4. After the Method setting is completed, enter the Plate section to enter the well plate setting interface.

Note: *The plate setup can be set before, during, or after the reaction operation.*

5. Melting Mode:
a. For analysis of nucleic acids, select “Nucleic Acid” here
b. For analysis of protein thermostability, select “Thermal stability assay” here



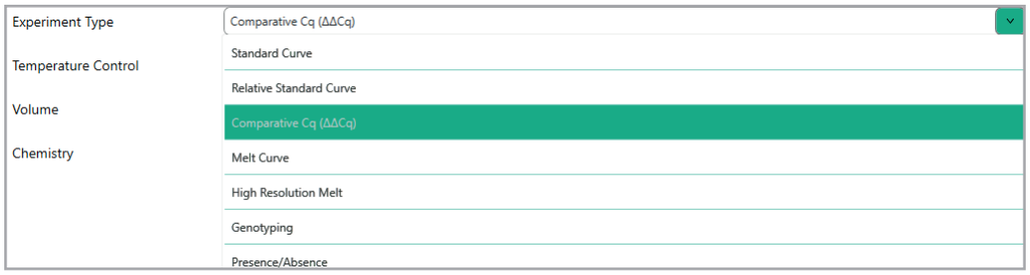
5.3. Relative Quantification Experiment (ΔΔCq)

Relative quantification is suitable for most gene expression studies and can analyze the up- or down-regulation of target gene expression levels in calibrated (normal) samples and one or more experimental samples. Using this technology does not require accurate determination of copy number but focuses on multiple changes compared with calibration samples. ΔΔCq method is a very common method of relative quantitative analysis, which can be used to compare the results of experimental samples, including calibrators (such as untreated or wild-type samples) and standard products (such as housekeeping gene expression), which can be used. Using this method, the target gene in the same two samples can be compared according to the Cq of the standard (normal) gene in the test sample and the calibration sample Cq, and the ΔΔCq value obtained can be used to determine the multiple difference of expression.

*Compare gene expression levels in different tissues.
*Compare the expression level samples of genes in treated and untreated samples.
*Compare the expression levels of genes of interest in different genetic backgrounds.
*Analysis of the conditions of gene expression over time under specific treatment.

5.3.1. Setup of comparative Cq (ΔΔCq) Analysis

1. Select experiment type “Comparative Cq (ΔΔCq)”.



2. Choose the used Plastic type and Chemistry.
3. Add name and optional information.
4. After the Method setting is completed, enter the Plate section to enter the well plate setting interface.

Note: *The plate setup can be set before, during, or after the reaction operation.*

5.4. Genotyping Experiments

In the study of gene variation, in order to obtain a large number of characteristic markers, a large number of individuals must be genotyped. The reasons for the differences in individual genomes at different levels include single nucleotide polymorphisms (SNP) and DNA covalent modification. More than 90% of the gene sequence variants that cause individual differences belong to single nucleotide polymorphisms (SNP). SNPs mainly refer to DNA sequence polymorphisms caused by mutations at the nucleotide level of the genome frequency of at least one allele in the population is not less than 1%, including conversion of single base, transversion and insertion and deletion of single base: in addition, changes in DNA covalent bond modifications, such as DNA methylation.

Real-time fluorescence quantitative PCR can provide different methods for genotyping analysis. For the detection of known mutations, the most commonly used basic method is the end point genotyping using enzyme cleavage hydrolysis probe, and the more advanced method is the fusion curve genotyping using hybrid probe. For the detection of unknown or new mutations, saturated DNA combined with dyes is usually used to genotyping by high resolution melting curve. In this chapter we will introduce the method of allele analysis of known mutation sites using the TaqMan™ probe endpoint method. The method can be easily improved and applied to the detection of various other SNP.

Common genotyping applications:

*Genetic identification of genetic diseases.
*Identification of tumor susceptibility genes and hotspot mutations.
*Identification of pathogen gene mutations and drug resistance.
*Animal and plant forensics, breeding research.

5.4.1. Setup of a Genotyping analysis

1. Select experiment type “Genotyping”.



2. Choose the used Plastic type and Chemistry.

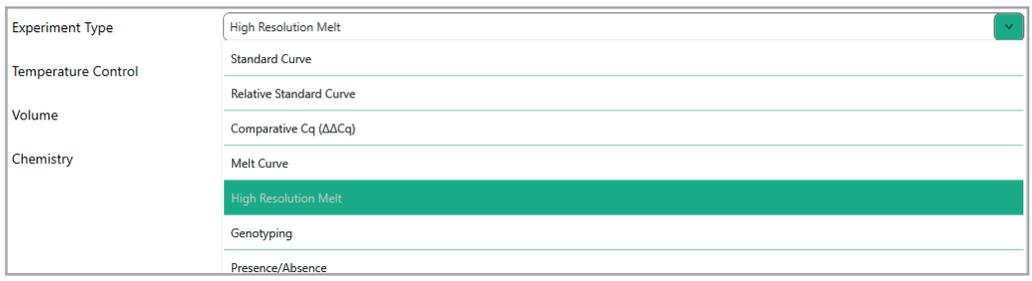
Note: *Genotyping experiments can only be performed using probe-based chemistry.*

3. Add name and optional information.
4. After the Method setting is completed, enter the Plate section to enter the well plate setting interface.

Note: *The plate setup can be set before, during, or after the reaction operation.*

5.4.2. Setup of a a High-Resolution Melt Analysis (HRM)

1. Select experiment type “High-Resolution Melt”.



2. Choose the used Plastic type and Chemistry.

Note: *Genotyping experiments can only be performed using dye-based chemistry. Please use an HRM-specific saturating dye.*

3. Add name and optional information.

4. After the Method setting is completed, enter the Plate section to enter the well plate setting interface.

Note: *The plate setup can be set before, during, or after the reaction operation.*

5. For HRM experiments controls can be defined during the plate setup (optional).

Click Add to add controls and select wells to assign controls similar to assigning samples and targets.



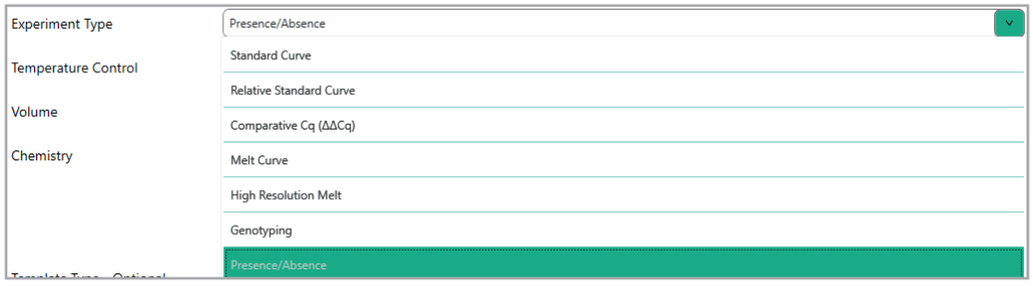
5.5. Presence/Absence Experiments

Presence/Absence experiments are endpoint experiments that can be performed to determine whether a target is present or absent in a sample. This type of experiment can be used for the detection of pathogens.

Presence/Absence experiments can only be performed using a probe-based assay and data is collected at the end. The amount of accumulated target is quantified for each sample. For quantification, the normalized intensity of the reporter dye (Rn) is used and displayed.

5.5.1. Setup of a Presence/Absence analysis

1. Select experiment type “Presence/Absence”.



2. Choose the used Plastic type and Chemistry.

Note: *Presence/Absence experiments can only be performed using probe-based chemistry.*

3. Add name and optional information.

4. After the Method setting is completed, enter the Plate section to enter the well plate setting interface.

Note: *The plate setup can be set before, during, or after the reaction operation.*

5. For Presence / Absence experiments four types of tasks can be applied to the plate setup: Unknown-IPC (well contains unknown sample and IPC (internal positive control)), positive control (well contains a positive control and IPC), NC (negative control)-IPC (well contains no-template control and IPC) and NC-blocked IPC (well contains no-template control and IPC blocking agent).

6. Result Analysis and Data Export

This chapter explains the data overview, analysis, and export.

6.1. Result Tab

6.1.1. Buttons in Toolbar

Toolbar	Name	Function
	Plate view	Plate overview for results
	List	Experiment results by list
	Print...	Print current window image file
	Copy image	Copy current window as image file
	Save to image file	Save current window as file (.jpg, .svg, .png, .emf, .pdf)
	Plot Properties	Plot properties set up
	Show a legend for the plot	Show/hide the legend for the plot
	Lock X Axis	Lock X Axis
	Lock Y Axis	Lock Y Axis
	Reset	Return to last step
	Show Plot Settings	Show Plot Settings

6.1.2. Other Toolbars

Toolbar	Name	Function
	Analyze	Click to analysis result after changing plate or plot settings
	Action	Click to select sample or target
	Save	Save current results as .qps file
	View	View current results by plate or list format
	View roup by	Click to show results by different groups
	Expand/Shrink	Expand/Shrink plate or image view

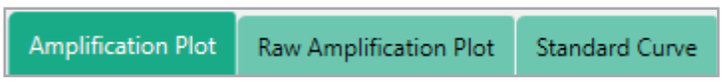
6.2. Result Overview

FastGene® qFYR Analysis Studio will analyze experiment results by default settings after an experiment is finished, and show amplification plot in the Result tab.

Attention: *After omitting wells or changing any settings, please click*  *to obtain new results.*

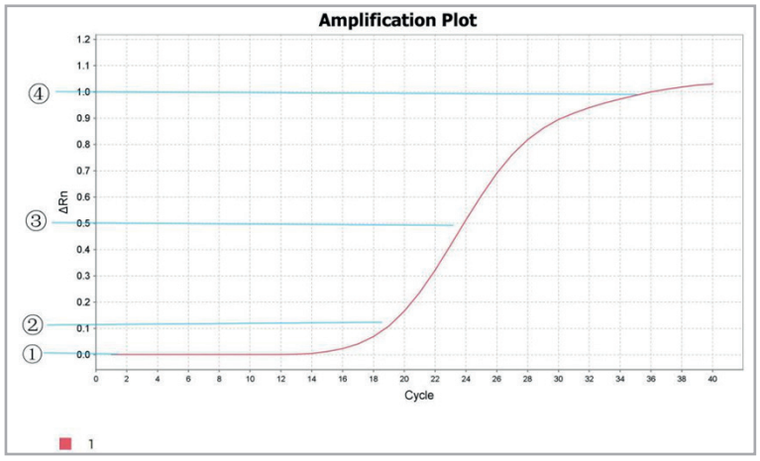
6.2.1. Amplification plot

1. Enter the Amplification plot tab to view the amplification curve.



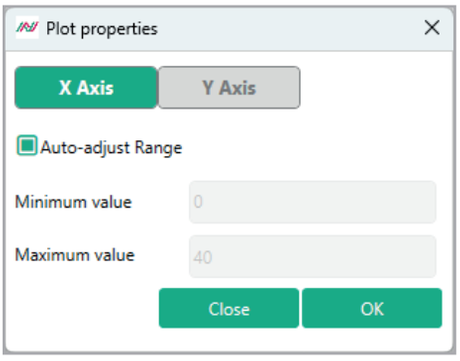
2. Overview amplification curve and evaluate curve shape. A typical amplification curve includes four different parts:

①Baseline stage ②Exponential stage ③Linear stage ④Plateau stage



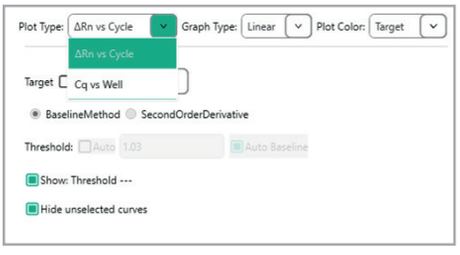
6.2.2. Plot Properties

Select , open the Plot Properties settings. In the ” X Axis” and ” Y Axis” tab, you can adjust the range, as shown below. Select “OK” and „Close“ if any adjustment is made. The adjusted curve will be shown after saving.



6.2.3. Plot Settings

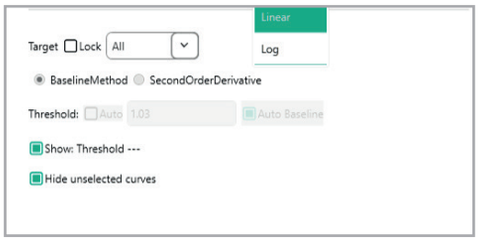
1. Select to enter the Plotting Settings and to adjust plot type, graph type, threshold etc..



2. Use the drop-down menu to choose the Plot Type. The default setting is “Rn vs cycle”, but “ΔRn vs cycle”, “Cq vs well” are available as well.

Plot Type	Description	Purpose
Rn vs cycle	Rn value is the normalized fluorescence value	Recognize and check unnormal amplification well
Cq vs well	Cq value is the cycle number which fluorescence signal achieved threshold	Recognize the unnormal amplification well

3. Graph Type can be changed from “Linear” to “Log” by using the drop-down list. The default setting is “Linear” graph.

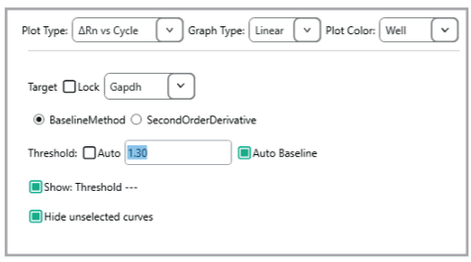


Note: *Plot settings can also be adjusted using the tool bar above the graph.*



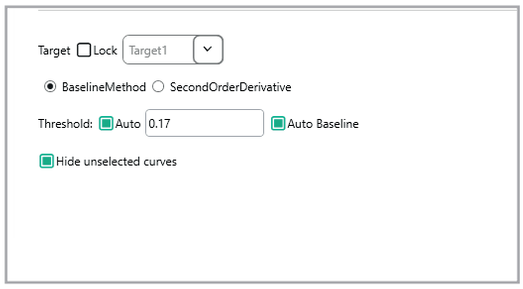
6.2.4 .Threshold Settings

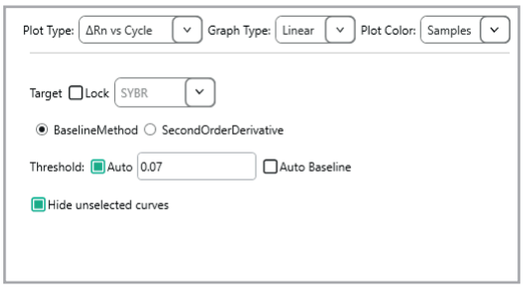
The software will calculate the threshold value by an automatic model. However, it can still be adjusted by unchecking “Auto” and changing the value. Additionally, the threshold line in the graph can be dragged. The new Cq value will be automatically calculated and displayed after the threshold value is adjusted. The threshold line will not be displayed if “Show Threshold” is not selected.

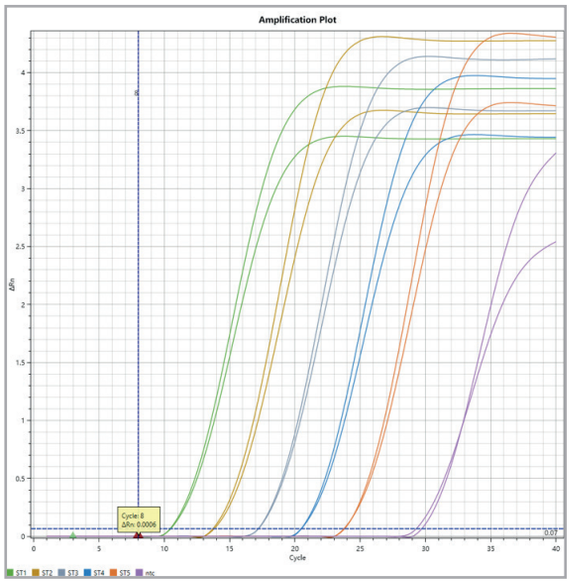


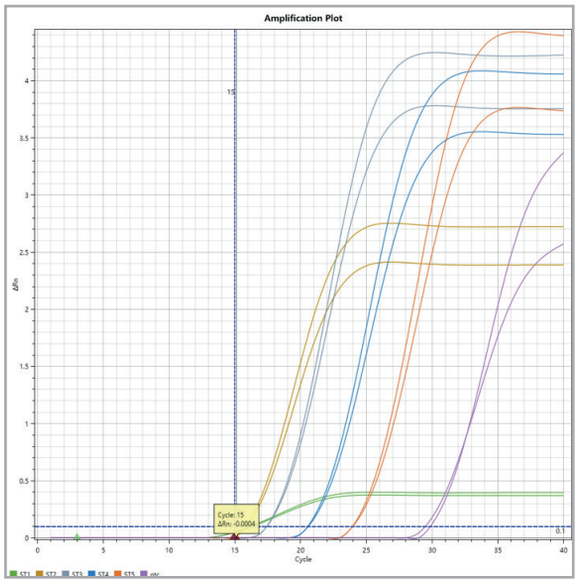
6.2.5. Baseline Settings

The software automatically sets the baseline in the first few cycles of the reaction, usually withing 3 to 15 cycles. If you need to set the baseline manually, remove the automatic threshold check, you can drag and drop the baseline (green triangle) directly on the curve and change it within the 1st to 15th cycle. In case of an early Cq value, it is suggested that the baseline should be set in the straight area of the curve, and when the baseline setting range is too wide, fluorescence can be detected in the 10th cycle, which may cause a curve drop and Cq delay.





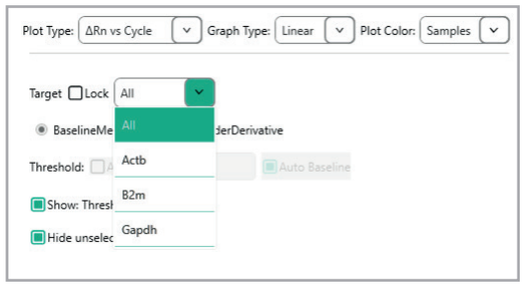


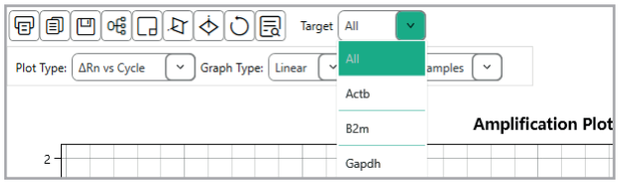


Note: *Please do not forget to click* Analyze *after a baseline setting is changed.*

6.2.6. Target Settings

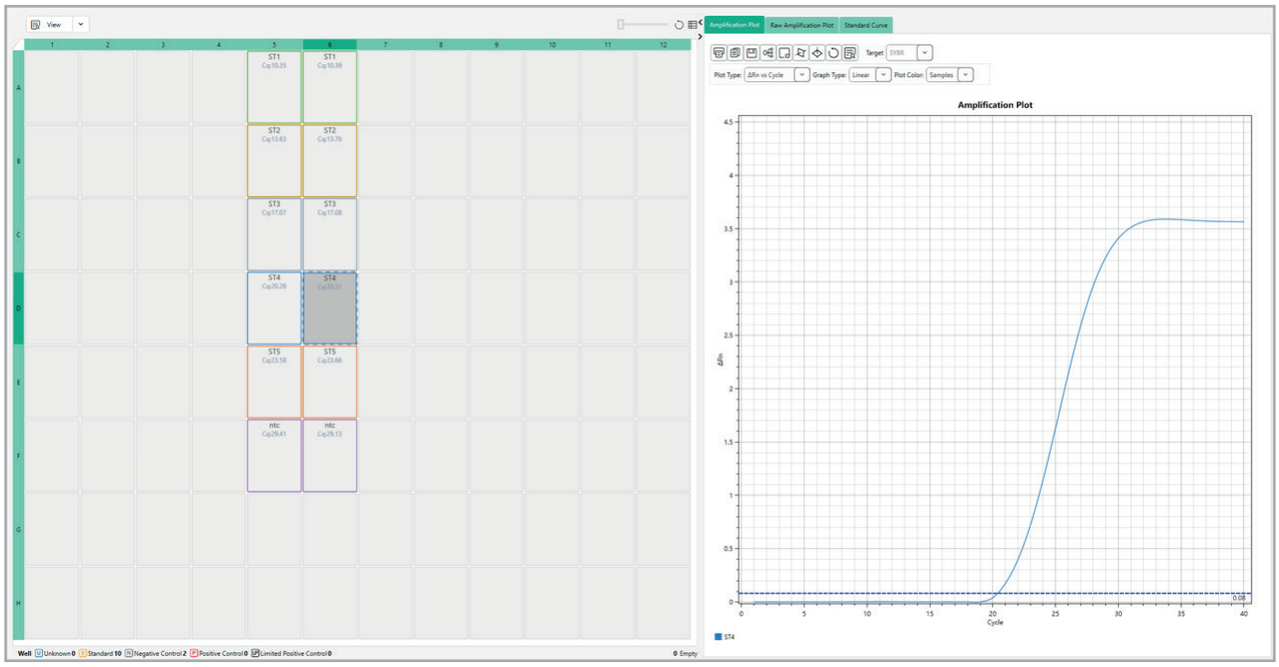
You can select different targets, or all targets to be shown:





6.2.7. Results as Reaction Board

To view the results in plate format, click  .

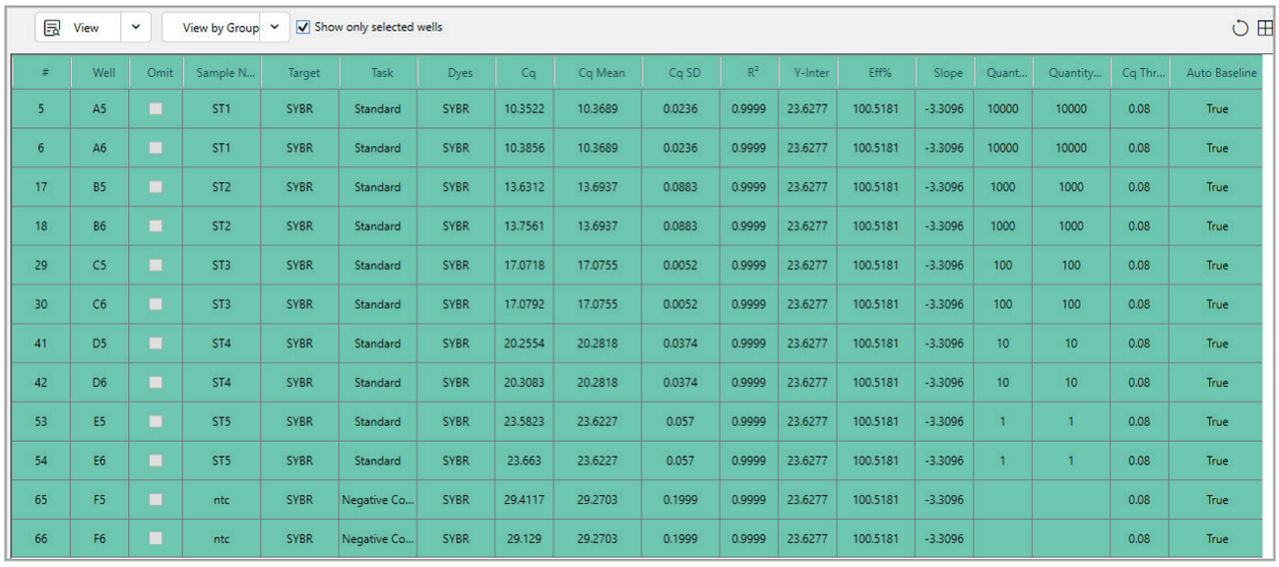


If a specific well is selected, the amplification plot will be shown accordingly. E.g., the amplification curve of well D6 is shown after selecting well D6 in the plate format, as in the figure above.

6.2.8. Results as List

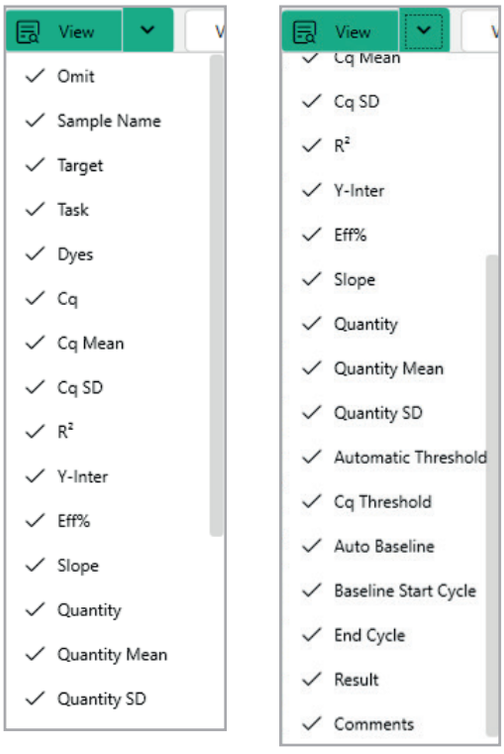
To display the results as list, click .

All the experiment information is listed, including well, sample, target, task, Cq, Cq mean, Cq SD etc.



#	Well	Omit	Sample N...	Target	Task	Dyes	Cq	Cq Mean	Cq SD	R^2	Y-Inter	Eff%	Slope	Quant...	Quantity...	Cq Thr...	Auto Baseline
5	A5		ST1	SYBR	Standard	SYBR	10.3522	10.3689	0.0236	0.9999	23.6277	100.5181	-3.3096	10000	10000	0.08	True
6	A6		ST1	SYBR	Standard	SYBR	10.3856	10.3689	0.0236	0.9999	23.6277	100.5181	-3.3096	10000	10000	0.08	True
17	B5		ST2	SYBR	Standard	SYBR	13.6312	13.6937	0.0883	0.9999	23.6277	100.5181	-3.3096	1000	1000	0.08	True
18	B6		ST2	SYBR	Standard	SYBR	13.7561	13.6937	0.0883	0.9999	23.6277	100.5181	-3.3096	1000	1000	0.08	True
29	C5		ST3	SYBR	Standard	SYBR	17.0718	17.0755	0.0052	0.9999	23.6277	100.5181	-3.3096	100	100	0.08	True
30	C6		ST3	SYBR	Standard	SYBR	17.0792	17.0755	0.0052	0.9999	23.6277	100.5181	-3.3096	100	100	0.08	True
41	D5		ST4	SYBR	Standard	SYBR	20.2554	20.2818	0.0374	0.9999	23.6277	100.5181	-3.3096	10	10	0.08	True
42	D6		ST4	SYBR	Standard	SYBR	20.3083	20.2818	0.0374	0.9999	23.6277	100.5181	-3.3096	10	10	0.08	True
53	E5		ST5	SYBR	Standard	SYBR	23.5823	23.6227	0.057	0.9999	23.6277	100.5181	-3.3096	1	1	0.08	True
54	E6		ST5	SYBR	Standard	SYBR	23.663	23.6227	0.057	0.9999	23.6277	100.5181	-3.3096	1	1	0.08	True
65	F5		ntc	SYBR	Negative Co...	SYBR	29.4117	29.2703	0.1999	0.9999	23.6277	100.5181	-3.3096			0.08	True
66	F6		ntc	SYBR	Negative Co...	SYBR	29.129	29.2703	0.1999	0.9999	23.6277	100.5181	-3.3096			0.08	True

The list can be grouped by the following:

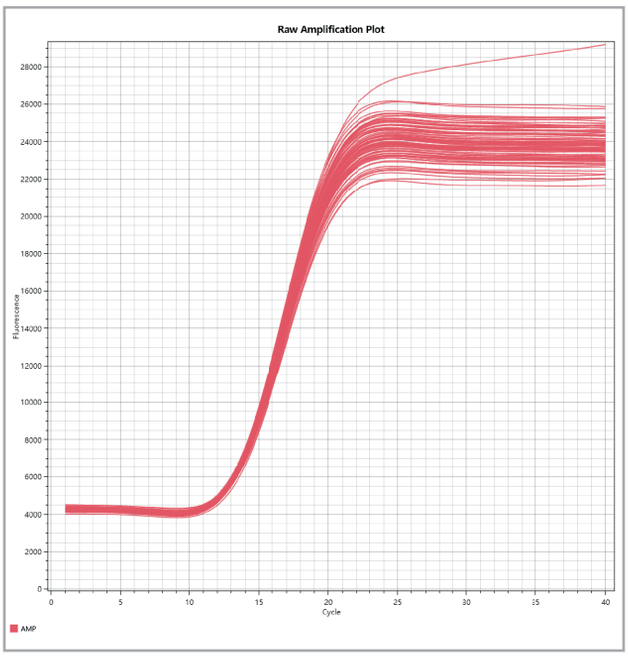


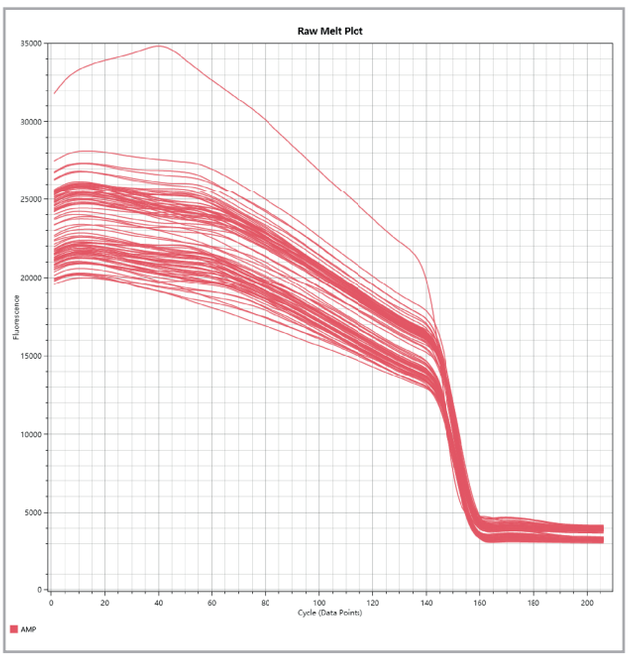
6.2.9. Raw Amplification Plot and Raw Melt Peak Plot

Raw plots present the real time fluorescence signal change. The cycle number is shown on the X-Axis while the fluorescence signal value is plotted on the Y-Axis.



By entering the corresponding tabs multicomponent plots for the amplification stage or for the melt curve step will be shown.





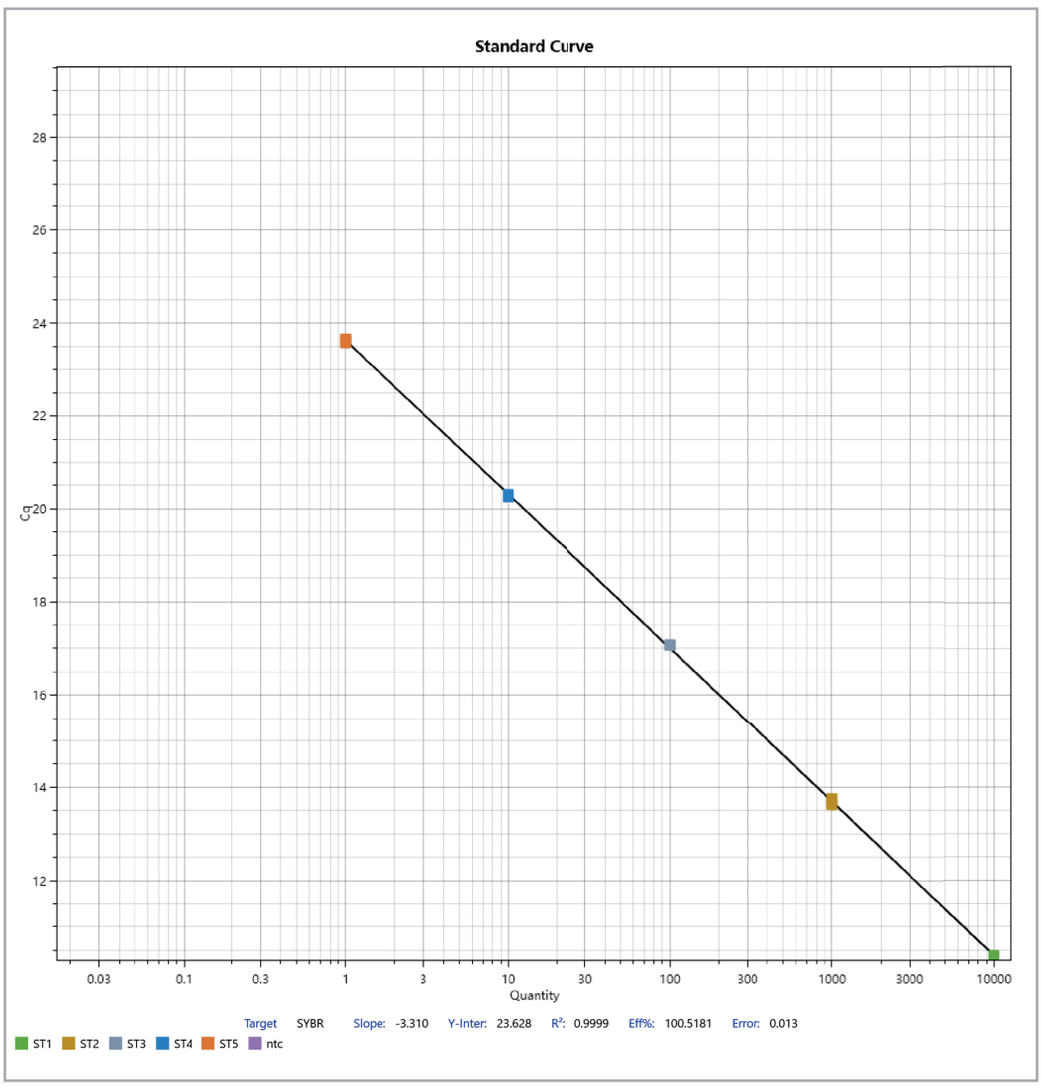
6.2.10. Standard Curve

Standard curves can be used to verify the amplification efficiency of comparative Cq experiments or to perform an absolute quantitative PCR experiment.

1. Click Standard Curve



2. View standard curve



Note: *The amplification efficiency (Eff%), slope, Y-Intercept, R^2 and Error will be shown below the graph. A slope of -3.2~-3.5, a R^2>0.99 and an Efficiency of 90 to 110% are indicators of a reliable experiment.*

3. The default mode shows red dots as standard samples, while blue and green are unknown samples.

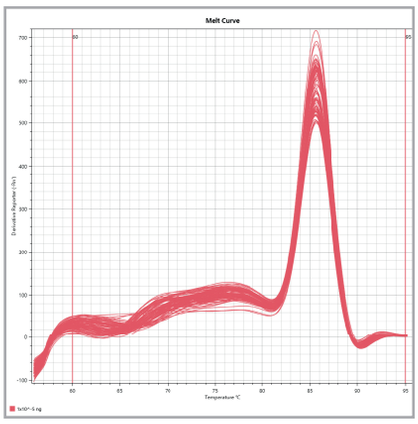
6.2.11. Melt Curve

The melt curve shows the reduction in fluorescence caused by the denaturation of the DNA double helix during the final slow heating step (melt curve analysis).

1. Enter Melt curve plot in the menu.

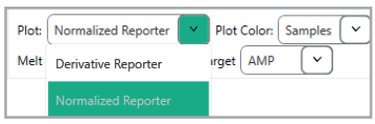


2. View derivative of the melt curve fluorescence plot

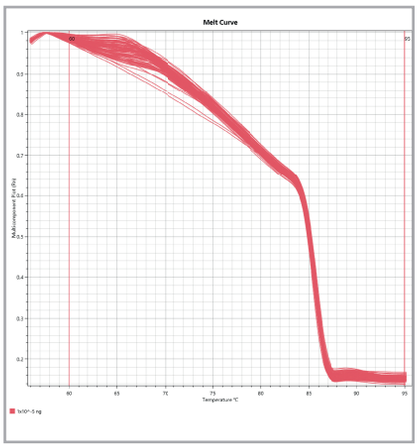


Note: *Melt curve plot is applied to check the specificity of PCR reaction and products: Two or more peaks are indicators of an unspecific amplification or of multiple amplicons.*

3. Change the plot configurations



Note: *The "Derivative reporter" plot displays the first derivative of fluorescence signal change according to the temperature. The Tm (Temperature of melt, the temperature of 50% of dsDNA unwinding to ssDNA) could be used to detect unspecific amplification and primer dimer.*



The "Normalized reporter plot" displays the fluorescence signal value decreasing as temperature rises.

6.2.12. Gene Expression Map

The melt curve shows the reduction in fluorescence caused by the denaturation of the DNA double helix during the final slow heating step (melt curve analysis).

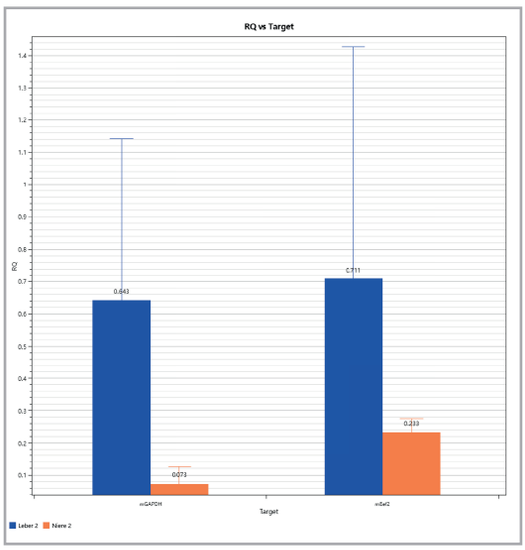
1. Click Gene Expression



2. All sample information is shown on the selected table

#	Sample N...	Target	Cq Mean	ΔCq Mean	ΔCq SE	ΔΔCq	Relative Quan...	RQ min	RQ Max
1	Leber 1	mACTB	28.1398	28.1398					
2	Leber 1	mGAPDH	34.0838	5.944			1		
3	Leber 1	mEef2	26.0509	-2.0889			1		
4	Leber 2	mACTB	27.0563	27.0563	0.1093				
5	Leber 2	mGAPDH	33.6378	6.5814	0.298	0.6374	0.6429	0.3623	1.1408
6	Leber 2	mEef2	25.4601	-1.5963	0.3623	0.4926	0.7107	0.3539	1.4274
7	Milz 1	mACTB	24.907	24.907					
8	Milz 2	mACTB	27.4466	27.4466					
9	Milz 2	mGAPDH	34.4203	6.9737		1.0296	0.4898		
10	Niere 2	mACTB	25.5796	25.5796	0.0878				
11	Niere 2	mGAPDH	35.3001	9.7204	0.2787	3.7764	0.073	0.0427	0.1248
12	Niere 2	mEef2	25.5908	0.0111	0.0882	2.1	0.2333	0.1968	0.2764

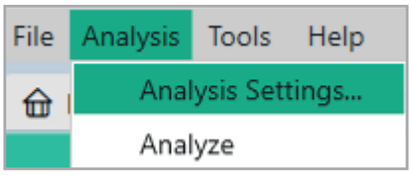
3. Check the gene expression map:



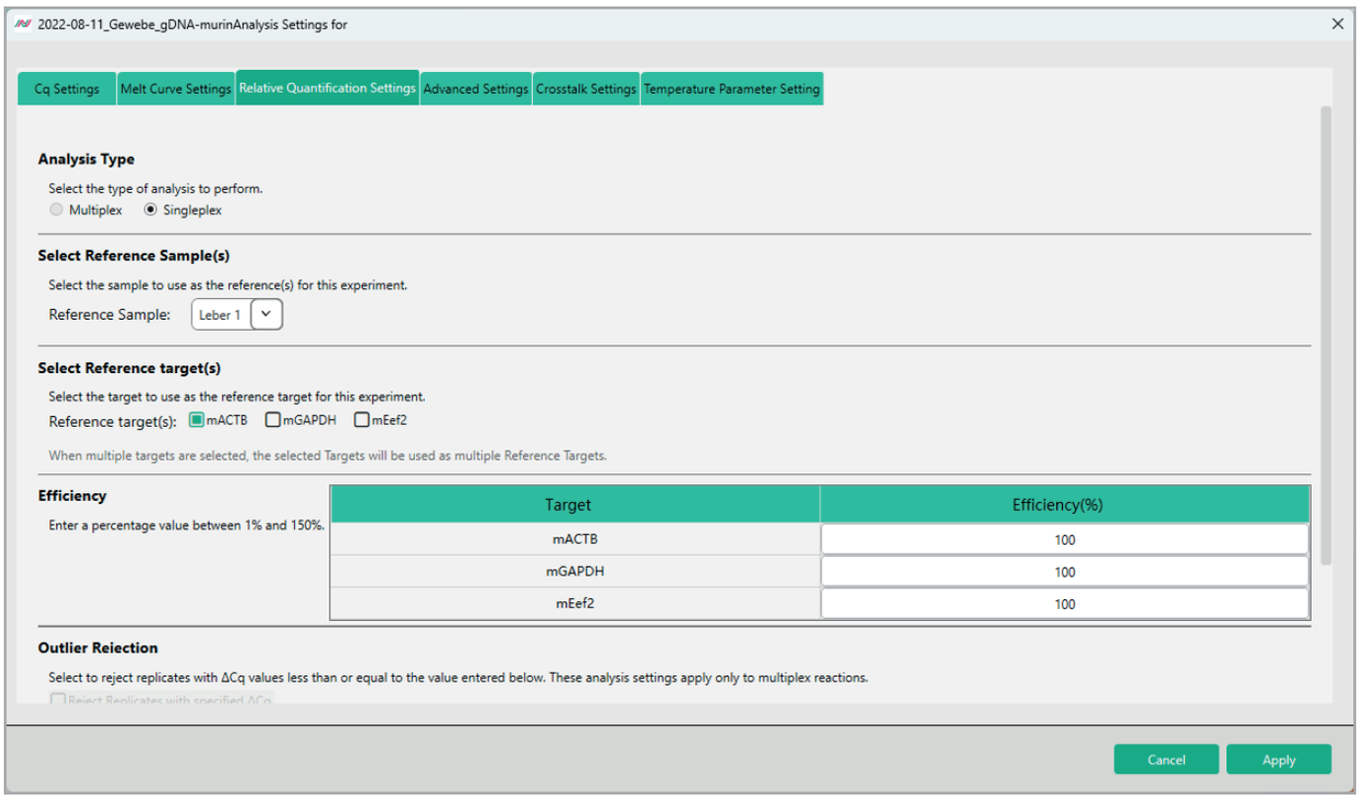
Note: *There are two plot types available: RQ vs Target (relative quantitation (RQ) values are grouped by target), and RQ vs Sample (relative quantitation (RQ) values are grouped by sample).*

6.2.13. Analysis Settings

General Analysis settings can be changed by entering the Analysis settings in the menu toolbar:



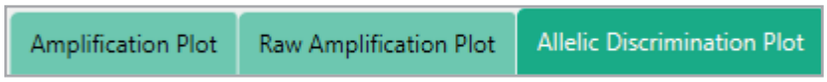
Enter the tab Relative Quantification Settings to edit settings used for relative quantification of data, such as References, Efficiency of used primers, Outlier Rejection and RQ Min/Max calculations.



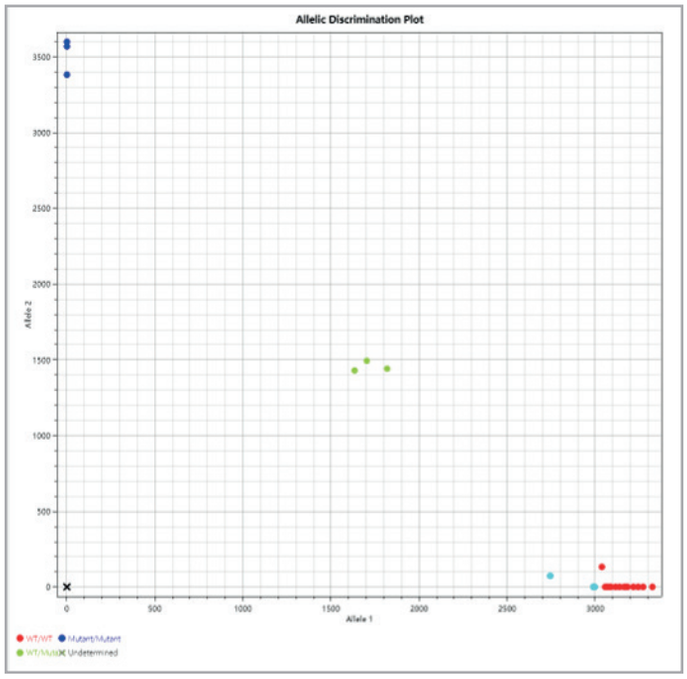
6.2.14. Genotyping Maps

The allelic discrimination plot clusters the normalized reporter fluorescence (Rn) of the allele-specific probes used in the SNP assay for the three possible genotypes (Allele 1 homozygous, Allele 2 homozygous, and Allele 1/2 heterozygous) as well as for negative controls (undetermined)

1. Click the allele identification map.



2. Check the allele identification map.



3. The allele identification map, the colors of different samples and the allele types represented by each color will be displayed.

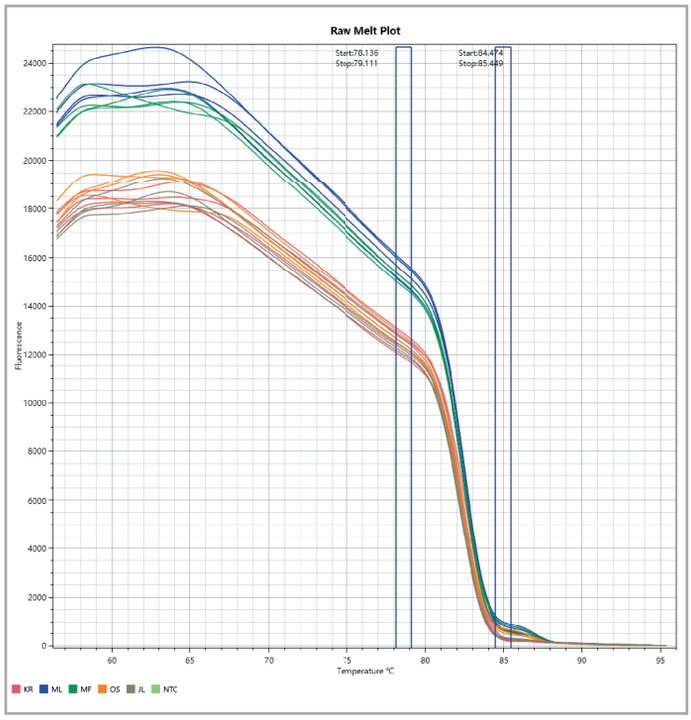
6.2.15. Genotyping Maps

The melt profile of a high-Resolution Melt analysis (HRM) reflects the mix of amplicons present in the sample. Variations in sample characteristics like GC content, length, sequence, and heterozygosity are defining the melt curve characteristics of each amplicon.

1. Click the Raw Melt curves tab.



2. Check the raw melt curves.



3. The software automatically defines the pre- and post-melt regions by pairs of vertical bars before and after the active melt region. If you need to set the regions manually, you can drag and drop the vertical bars of each region directly on the curve. The area between the pair of bars to the left of the active melt region is used to designate 100% fluorescence, where all amplicons are double-stranded. Respectively, the area between the bars to the right of the active melt region is used to determine 0% fluorescence, where all amplicons are single-stranded.

Note: *Please do not forget to click* Analyze *after melt-region setting is changed.*

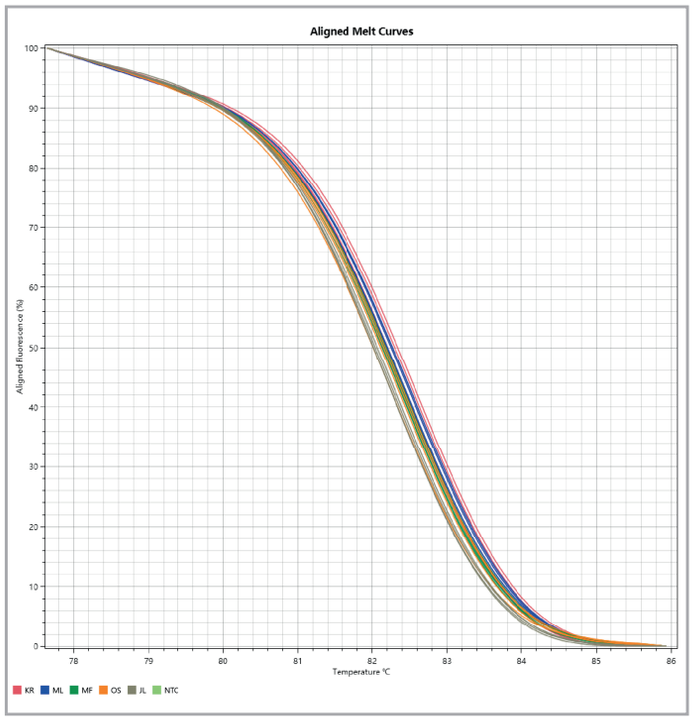
6.2.16. Aligned Melt Curves of the High-Resolution Melt analysis

During HRM two types of melt curves can be observed. On the one hand, melt curves that differ from each other in melting temperature (Tm) of the amplicon, but are similar in curve shape. Usually, this kind of curve is generated by homozygous variant samples. On the other hand, melt curves that differ in shape from homozygote melt curves usually indicate heterozygous samples due to the presence of base-pairing mismatches (heteroduplexes) present in the PCR product mix.

1. Click the Aligned Melt curves tab.



2. Check the aligned melt curves.



Note: *A scaled view of the data is displayed in the aligned plot. Sequence variants with differences in their melt curve (e.g., homozygous vs. heterozygous variants) can be easily discriminated.*

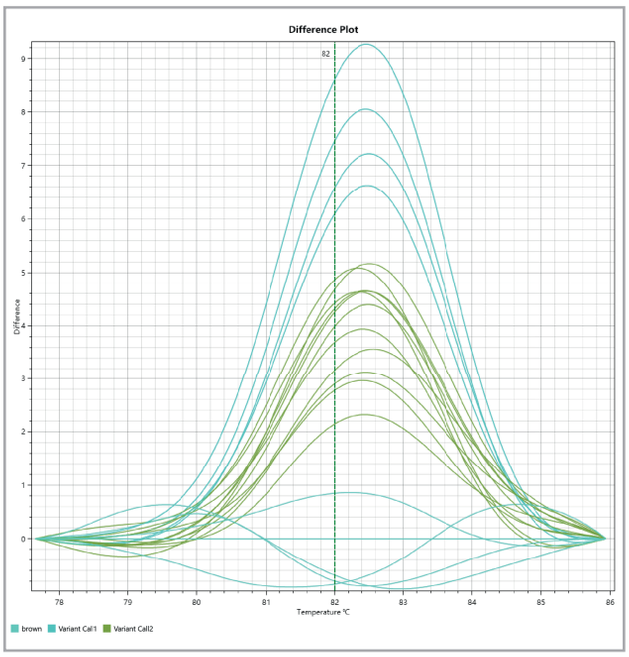
6.2.17. Difference plot of the High-Resolution Melt analysis

The difference plot is a tool to easily visualize small differences in melt curves of HRM analysis. Curves displayed here are generated by the differences between samples and a single reference.

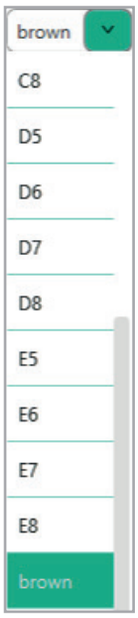
1. Click the Difference Plot tab.



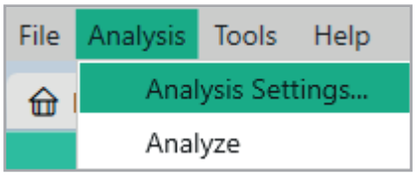
2. Check the difference plot.



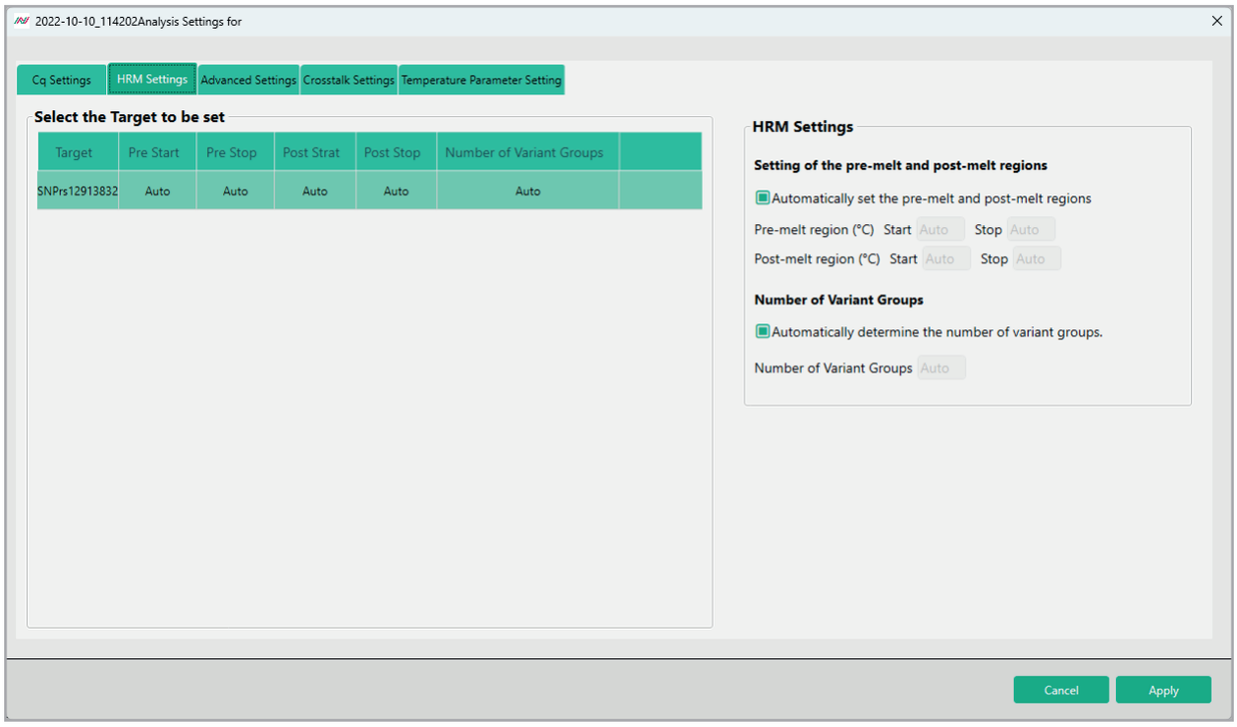
3. References can be changed by using the „Reference“ drop-down menu and can be either pre-defined controls or single wells.



4. The default number of variants in the software pre-set is three. If the actual number of variants is less, any slight difference among the replicates may lead to the replicates of one sample to be divided into different variation groups, even if they have similar curve characteristics. Default settings can be changed by opening the analysis settings in the menu toolbar.



In the tab “HRM Settings” uncheck “Automatically determine the number of variant groups” and enter the desired number. Confirm by clicking Apply .

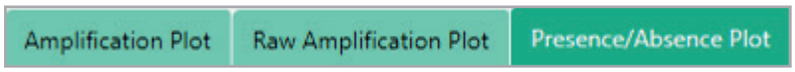


Note: *Please do not forget to click* Analyze *after HRM setting is changed.*

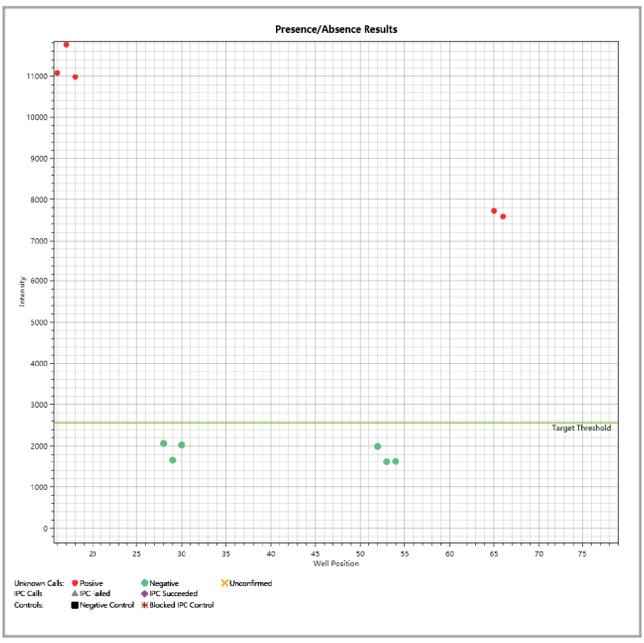
6.2.18. Presence/Absence Plot

The Presence/Absence plot displays the fluorescence intensity measured in each well.

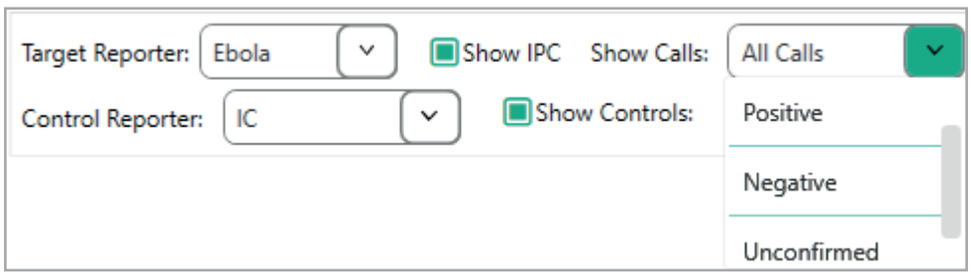
1. Enter the Presence/Absence plot tab.



2. Check the Presence/Absence results.



3. There are 4 types of plot views that can be chosen in the Plot settings: “All calls” – all calls are displayed, “Positive” – only samples where the target is present are displayed, “Negative” - only samples where the target is absent are displayed, and “Unconfirmed”, only unconfirmed samples are displayed. Additionally, you can choose to display fluorescence intensity of the IPC target (“Show IPC”) as well as fluorescence intensity of the IPC target in Negative Control-IPC wells and Negative Control-Blocked IPC wells (“Show Controls”).



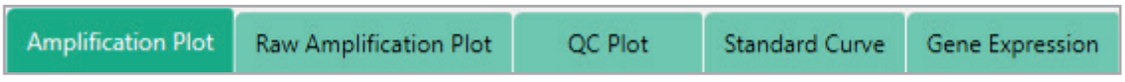
4. The software is calculating two types of thresholds for the Presence/Absence plot that are used for the call:

- The “IPC Threshold” is calculated from Negative Control- Blocked IPC reactions.
- The “Target Threshold” is calculated from Negative Control- IPC reactions.
- Targets are called regarding their fluorescence intensity:
  - Call is present if fluorescence intensity is higher than the target threshold
  - Call is absent if fluorescence intensity is lower than the target threshold, and fluorescence intensity of the IPC is higher than the IPC threshold,
  - Call is unconfirmed if fluorescence intensity is lower than the target threshold, and fluorescence intensity of the IPC is lower than the IPC threshold.

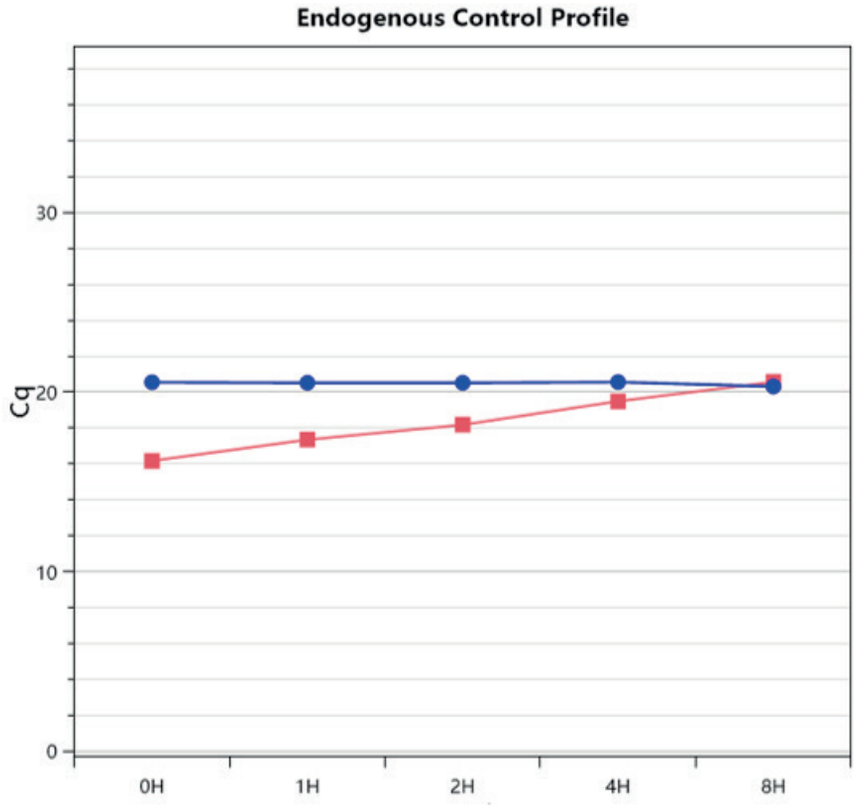
6.2.19. QC Plot

The QC plot displays the Cq levels of genes in different samples and helps users select reference genes for their experiment. All testing items can be displayed in the QC plot. The horizontal axis represents the sample type, and the vertical axis represents the Cq value.

1. Enter the QC Plot tab.



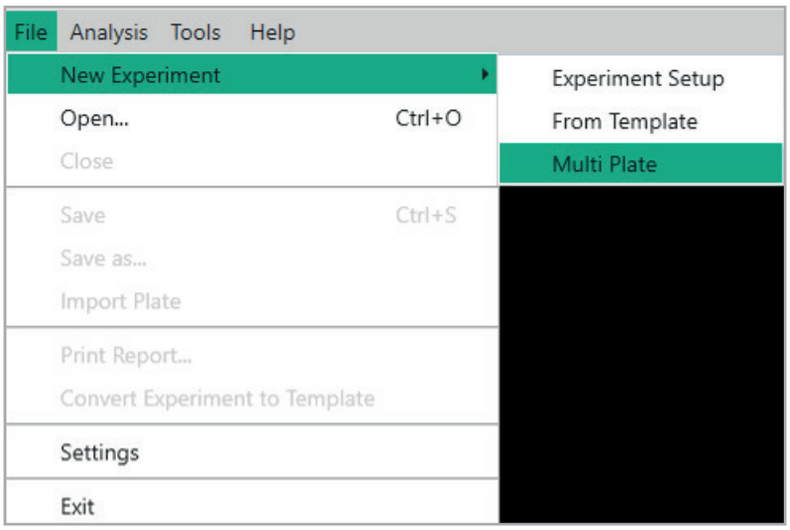
2. Check the Endogenous Control Profile.



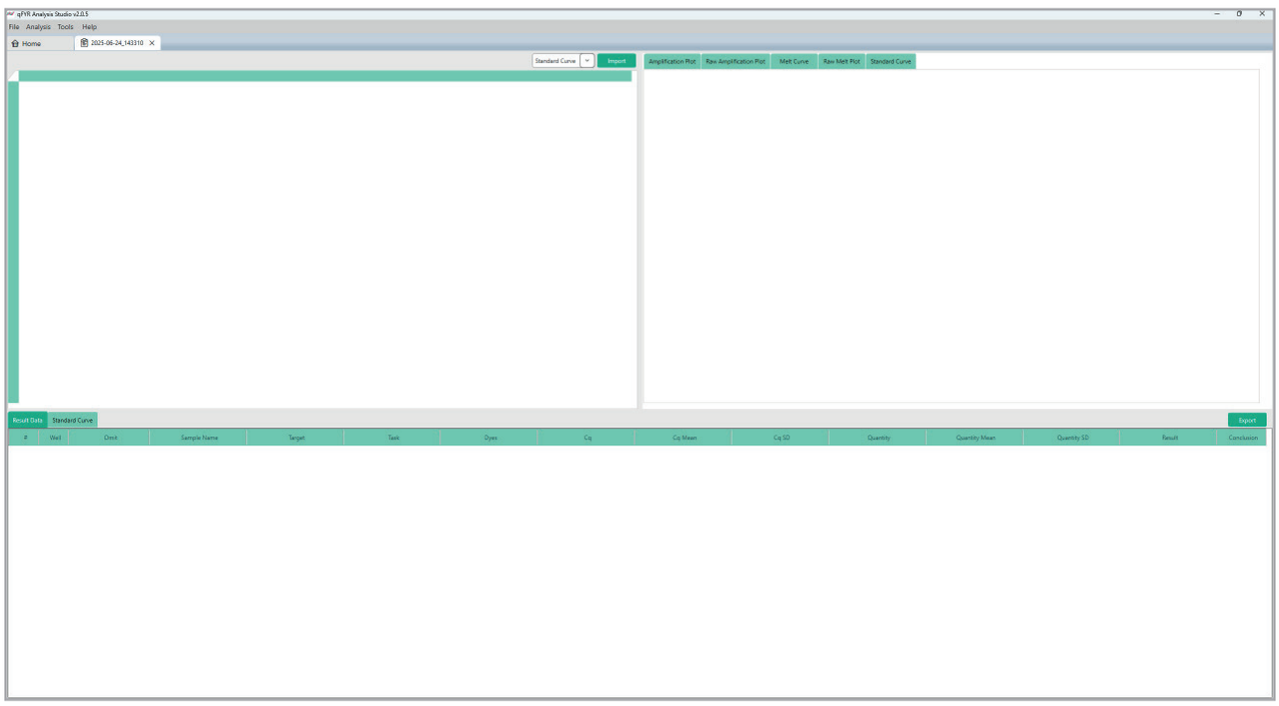
6.2.20. Multi Plate Analysis

It is possible to analyse plates from several experiments simoultanously.

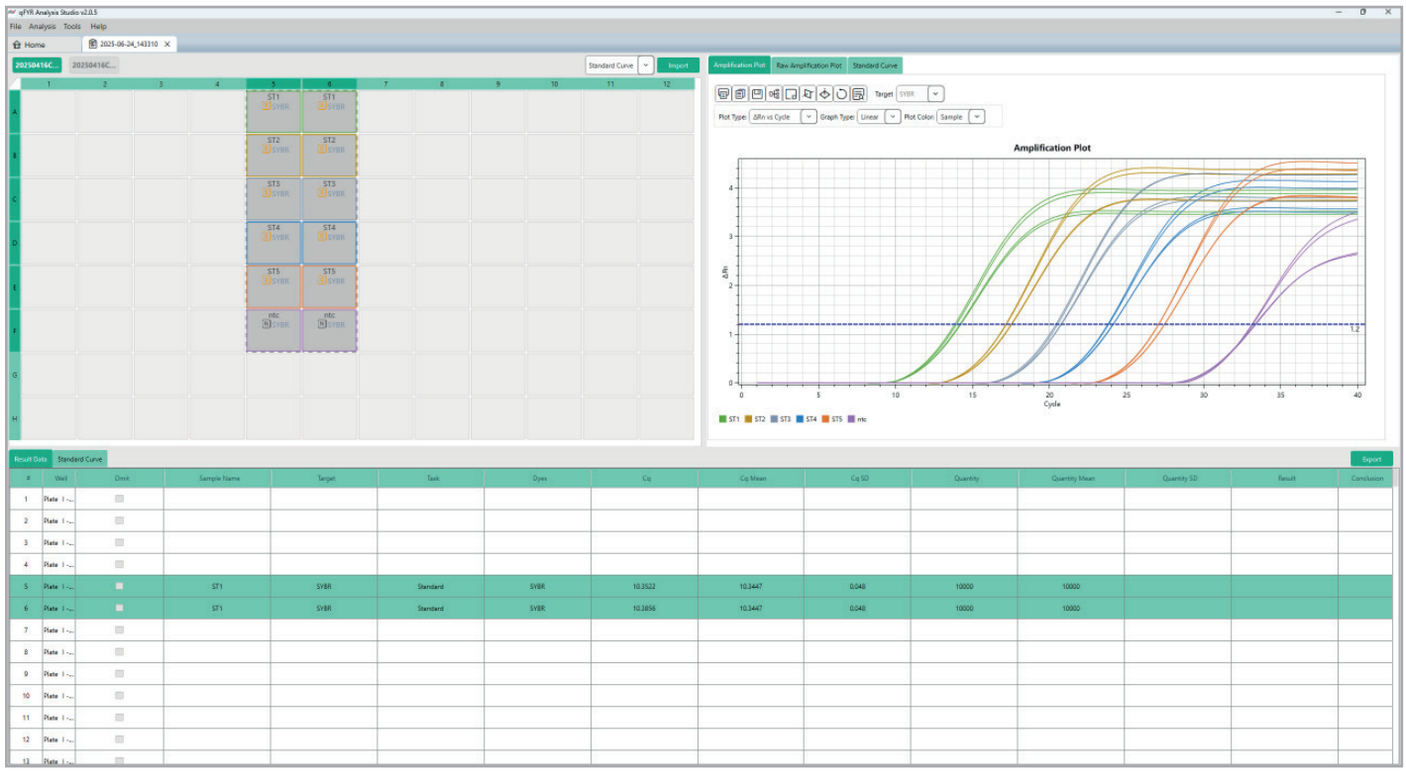
click „File“- „New Experiment“ and „Multi Plate“ button in the toolbar and then start the Multiplate analysis function.



The Multiplate analysis section is divided into three parts: data source (top left), result graph (top right), and result list (bottom).



In the data source section, click the Import button to retrieve the experimental result file from the computer (only support . qpsx format). It supports individual data addition and batch data addition.

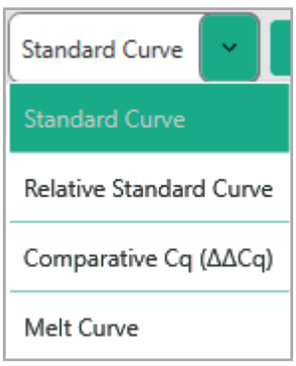


Click the file name to switch and view the well location setting



Check the well location from the data source section, the corresponding result curve will display.

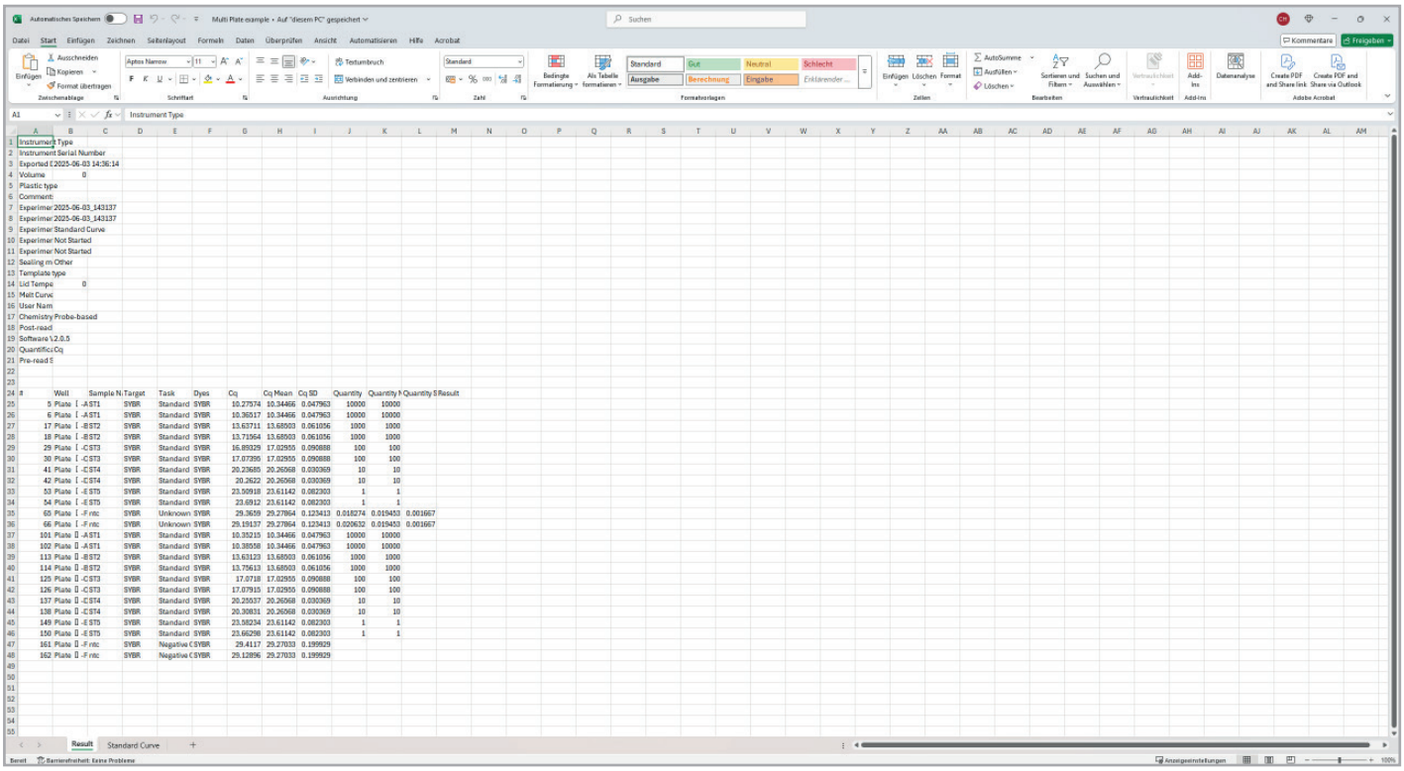
There are four different analysis types: Standard Curve, Relative Standard Curve, Comparative Cq (Δ Δ Cq) and Melt Curve.



Depending on the analysis type selected, the result graph section will display related result data and the Result Data's contents are also related to the analysis type.



Export data: Click on the ‚Export' button on the right side of ‚Result Data' to export the result data to an Excel spreadsheet. If there are multiple results, the data table will be stored in different sheets.



1	Instrument Type													
2	Instrument Serial Number													
3	Exported D	2025-06-03 14:36:14												
4	Volume	0												
5	Plastic type													
6	Comment:													
7	Experimer	2025-06-03_143137												
8	Experimer	2025-06-03_143137												
9	Experimer	Standard Curve												
10	Experimer	Not Started												
11	Experimer	Not Started												
12	Sealing m	Other												
13	Template type													
14	Lid Tempe	0												
15	Melt Curve													
16	User Nam													
17	Chemistry	Probe-based												
18	Post-read													
19	Software \	2.0.5												
20	Quantificc	Cq												
21	Pre-read S													
22														
23														
24	#	Well	Sample N	Target	Task	Dyes	Cq	Cq Mean	Cq SD	Quantity	Quantity M	Quantity S	Result	
25	5	Plate [	A ST1	SYBR	Standard	SYBR	10.27574	10.34466	0.047963	10000	10000			
26	6	Plate [	A ST1	SYBR	Standard	SYBR	10.36517	10.34466	0.047963	10000	10000			
27	17	Plate [	B ST2	SYBR	Standard	SYBR	13.63711	13.68503	0.061056	1000	1000			
28	18	Plate [	B ST2	SYBR	Standard	SYBR	13.71964	13.68503	0.061056	1000	1000			
29	29	Plate [	C ST3	SYBR	Standard	SYBR	16.89329	17.02955	0.090888	100	100			
30	30	Plate [	C ST3	SYBR	Standard	SYBR	17.07395	17.02955	0.090888	100	100			
31	41	Plate [	C ST4	SYBR	Standard	SYBR	20.23685	20.26568	0.030369	10	10			
32	42	Plate [	C ST4	SYBR	Standard	SYBR	20.2622	20.26568	0.030369	10	10			
33	53	Plate [	E ST5	SYBR	Standard	SYBR	23.50918	23.61142	0.082303	1	1			
34	54	Plate [	E ST5	SYBR	Standard	SYBR	23.6912	23.61142	0.082303	1	1			
35	65	Plate [	F ntc	SYBR	Unknown	SYBR	29.3659	29.27864	0.123413	0.018274	0.019453	0.001667		
36	66	Plate [	F ntc	SYBR	Unknown	SYBR	29.19137	29.27864	0.123413	0.020632	0.019453	0.001667		
37	101	Plate [	A ST1	SYBR	Standard	SYBR	10.35215	10.34466	0.047963	10000	10000			
38	102	Plate [	A ST1	SYBR	Standard	SYBR	10.38558	10.34466	0.047963	10000	10000			
39	113	Plate [	B ST2	SYBR	Standard	SYBR	13.63123	13.68503	0.061056	1000	1000			
40	114	Plate [	B ST2	SYBR	Standard	SYBR	13.73613	13.68503	0.061056	1000	1000			
41	125	Plate [	C ST3	SYBR	Standard	SYBR	17.0718	17.02955	0.090888	100	100			
42	126	Plate [	C ST3	SYBR	Standard	SYBR	17.07915	17.02955	0.090888	100	100			
43	137	Plate [	C ST4	SYBR	Standard	SYBR	20.25537	20.26568	0.030369	10	10			
44	138	Plate [	C ST4	SYBR	Standard	SYBR	20.30831	20.26568	0.030369	10	10			
45	149	Plate [	E ST5	SYBR	Standard	SYBR	23.58234	23.61142	0.082303	1	1			
46	150	Plate [	E ST5	SYBR	Standard	SYBR	23.66298	23.61142	0.082303	1	1			
47	161	Plate [	F ntc	SYBR	Negative C	SYBR	29.4117	29.27033	0.199929					
48	162	Plate [	F ntc	SYBR	Negative C	SYBR	29.12896	29.27033	0.199929					
49														
50														
51														
52														
53														
54														
55														

Click ‚File' button - ‚Save' or ‚Save as' in the toolbar, to save the multiplate data as a new file (*.qpsm).

Click ‚File' button, then click ‚OPen' in the toolbar and choose the multiplate data file to display (*.qpsm).

6.3. Data Export

6.3.1. Toolbars in Export Tab

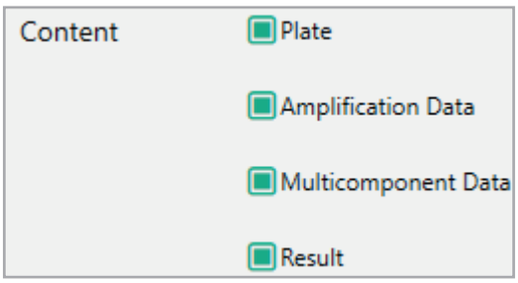
Toolbars	Name	Function
Export	Export	Export data
Save	Save	Save as . qpsx file

6.3.2. Data Export

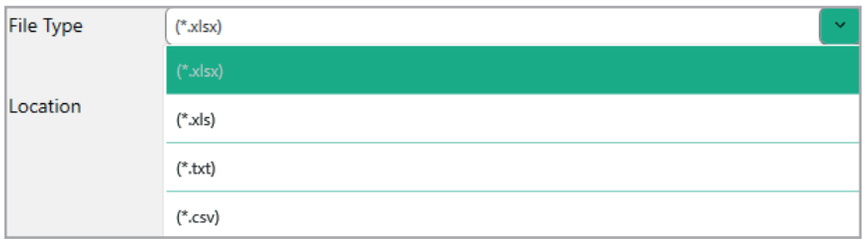
1. File name: The default file name is the name defined in “Plot properties” or could be changed to customized name. English characters are supported, but not special characters.



2. Export content: Include Plate Setup, Amplification Data, Multicomponent Data and Results. More content is available depending on the experiment type. The contents can be selected by box checking.



3. Select: .xlsx, .xsl, .txt or .csv.



4. File Location: The default file location is desktop, which can be changed to other file location by click Browse... .

5. File Export: The experiment result will be saved and opened in Excel automatically after clicking Export .

7. The export is shown in Excel as below.

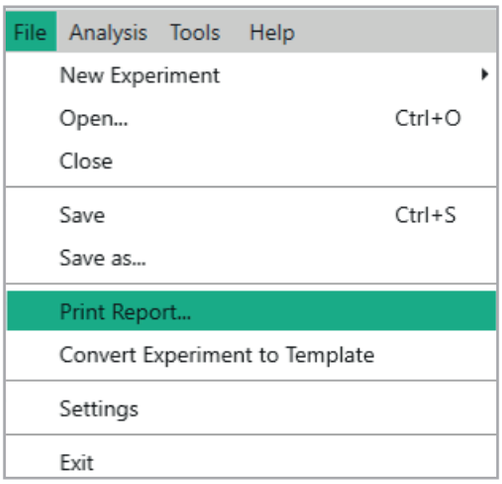
	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U
1	Instrument Type					FastGene qFYR PLUS															
2	Instrument Serial Number					24NGE96DX003															
3	Exported Date					2025-06-17 12:20:51															
4	Volume					20															
5	Stage/ Cycle where Cq Analysis is performed					Stage2,Step2															
6	Plastic type																				
7	Comments																				
8	Experiment File Name					20250416CH.qpsx															
9	Experiment Name					20250416															
10	Experiment Type					Standard Curve															
11	Experiment Run Start Time					2025-04-16 14:30:54															
12	Experiment Run End Time					2025-04-16 15:39:33															
13	Sealing method					Other															
14	Template type					OTHER															
15	Lid Temperature					100															
16	Melt Curve Stage																				
17	User Name																				
18	Chemistry					Dye-based															
19	Post-read Stage/Step																				
20	Software Version					2.0.5															
21	Quantification Cycle Method					Cq															
22	Pre-read Stage/Step																				
23																					
24																					
25	#	Well	Sample N	Target	Task	Dyes	Cq	Cq Mean	Cq SD	R^2	Y-Inter	Eff%	Slope	Quantity	Quantity M	Quantity S	Automatic	Cq Thresh	Auto Base	Baseline S	End Cycle
26	5	A5	ST1	SYBR	Standard	SYBR	10.352152	10.368866	0.0236382	0.9998832	23.627650	100.51814	-3.309574(	10000	10000		True	0.08	True	3	8
27	6	A6	ST1	SYBR	Standard	SYBR	10.385581	10.368866	0.0236382	0.9998832	23.627650	100.51814	-3.309574(	10000	10000		True	0.08	True	3	8
28	17	B5	ST2	SYBR	Standard	SYBR	13.631226	13.693677	0.0883194	0.9998832	23.627650	100.51814	-3.309574(	1000	1000		True	0.08	True	3	8
29	18	B6	ST2	SYBR	Standard	SYBR	13.756129	13.693677	0.0883194	0.9998832	23.627650	100.51814	-3.309574(	1000	1000		True	0.08	True	3	8
30	29	C5	ST3	SYBR	Standard	SYBR	17.071795	17.075473	0.0052016	0.9998832	23.627650	100.51814	-3.309574(	100	100		True	0.08	True	3	8
31	30	C6	ST3	SYBR	Standard	SYBR	17.079151	17.075473	0.0052016	0.9998832	23.627650	100.51814	-3.309574(	100	100		True	0.08	True	3	8
32	41	D5	ST4	SYBR	Standard	SYBR	20.255371	20.281839	0.0374307	0.9998832	23.627650	100.51814	-3.309574(	10	10		True	0.08	True	3	8
33	42	D6	ST4	SYBR	Standard	SYBR	20.308306	20.281839	0.0374307	0.9998832	23.627650	100.51814	-3.309574(	10	10		True	0.08	True	3	8
34	53	E5	ST5	SYBR	Standard	SYBR	23.582338	23.622656	0.0570191	0.9998832	23.627650	100.51814	-3.309574(	1	1		True	0.08	True	3	8
35	54	E6	ST5	SYBR	Standard	SYBR	23.662975	23.622656	0.0570191	0.9998832	23.627650	100.51814	-3.309574(	1	1		True	0.08	True	3	8
36	65	F5	ntc	SYBR	Negative C	SYBR	29.411696	29.270326	0.1999285	0.9998832	23.627650	100.51814	-3.309574(				True	0.08	True	3	8
37	66	F6	ntc	SYBR	Negative C	SYBR	29.128955	29.270326	0.1999285	0.9998832	23.627650	100.51814	-3.309574(				True	0.08	True	3	8

6.3.3. Printed report

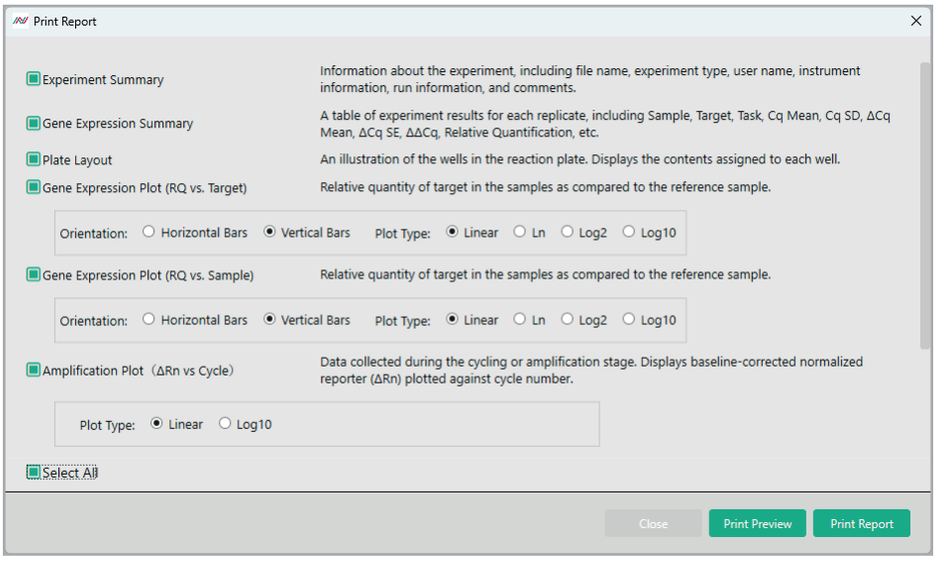
Next to data export, a report of the complete data for an experiment, including graphs, can be saved, and printed.

1. In the result section, select the wells that should be reported.

2. Access the main menu toolbar, enter the “File” drop down and click “Print Report...”

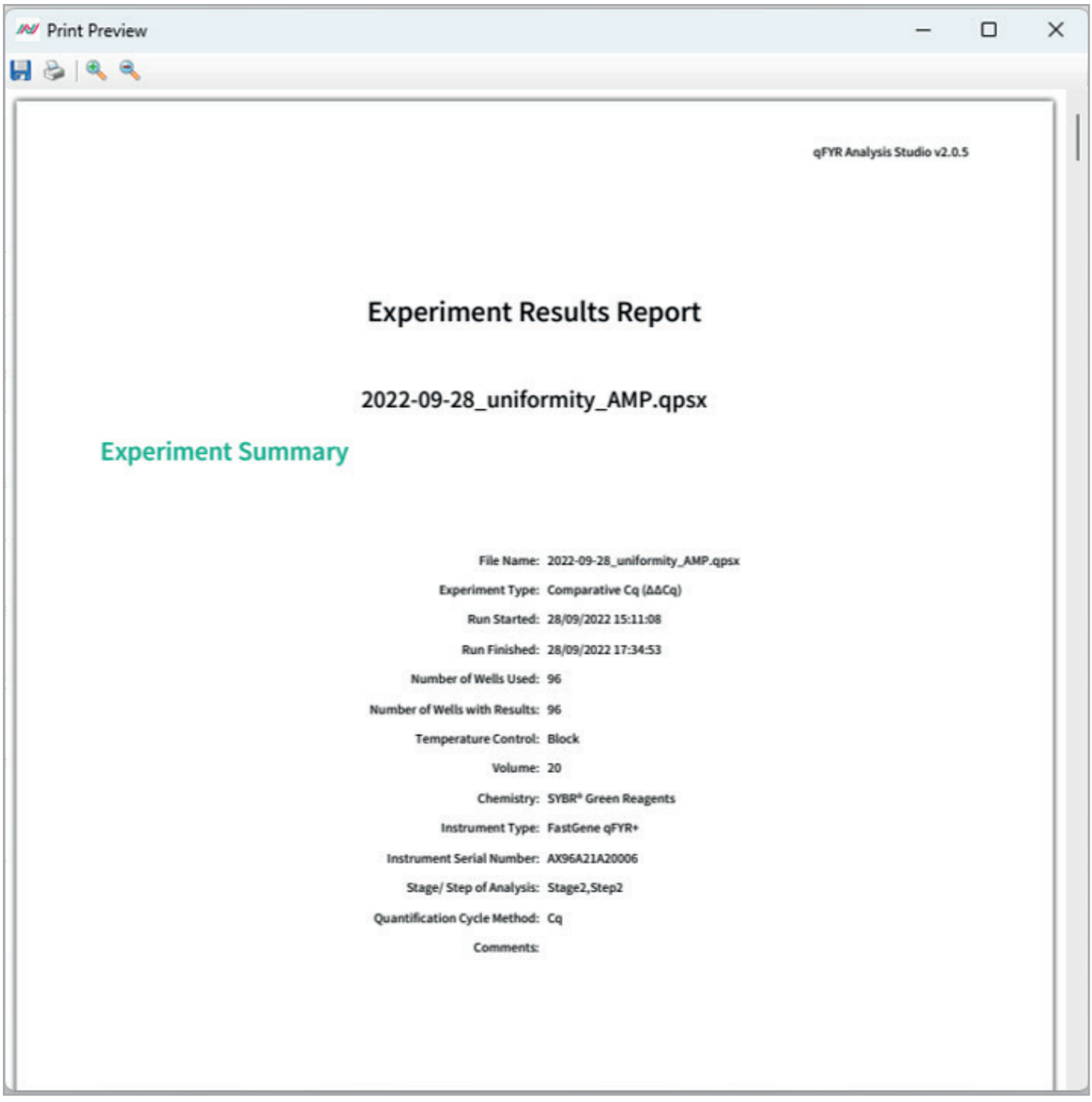


3. A pop-up menu will open to select the data and graphs to be reported



4. Report can either be directly sent to a connected printer by clicking Print Report or previewed and saved by clicking Print Preview .

5. “Print Preview” will open a new pop up and document can be reviewed and saved or printed. File format for saving is *.pdf and file can be password-secured.


Print Preview
qFYR Analysis Studio v2.0.5
Experiment Results Report
2022-09-28_uniformity_AMP.qpsx
Experiment Summary
File Name: 2022-09-28_uniformity_AMP.qpsx
Experiment Type: Comparative Cq (ΔΔCq)
Run Started: 28/09/2022 15:11:08
Run Finished: 28/09/2022 17:34:53
Number of Wells Used: 96
Number of Wells with Results: 96
Temperature Control: Block
Volume: 20
Chemistry: SYBR® Green Reagents
Instrument Type: FastGene qFYR+
Instrument Serial Number: AX96A21A20006
Stage/ Step of Analysis: Stage2,Step2
Quantification Cycle Method: Cq
Comments:

7. Instruments Maintenance

This instrument does not need a lot of maintenance in the case of routine use. If this instrument is used for a long time, it is necessary to clean and maintain the instrument regularly to ensure normal instrument operation. Before cleaning the instrument, please read this chapter carefully. Correct maintenance and cleaning help to prolong the instrument's service life.

If you need to replace any instrument parts during the maintenance, please contact us for professional technical confirmation, and only replace the parts provided by our original factory or our official license agency.

7.1. Instrument Cleaning

Do not clean or maintain the instrument during electrification or operation.

Do not pour water or other solutions into the sample bin, sample module or any instrument component. When the instrument is electrified, the flow of liquid may lead to electric shock!

70% medical disinfectant alcohol is recommended for instrument cleaning and disinfectant. However, ethanol is a volatile liquid. Exposure can irritate the eyes, skin, and respiratory tract, and can lead to central nervous system dysfunction and liver damage. When cleaning with ethanol, please wear suitable protective goggles, clothing, and gloves.

Note: If the instrument user has any questions about the compatibility of disinfectants or cleaning solution with the materials contained in the equipment parts, please consult the manufacturer or official license agency.

During operation, the sample module may have a high temperature. Before cleaning or maintaining, wait for the temperature to drop to room temperature to avoid scalding.

Biological Hazards

Please treat and operate all samples as potential biological hazards. If liquid samples overflow or splatter, please immediately take appropriate disinfectants for disinfection, to avoid contamination or damage to laboratory personnel.

Do not deal with and operate any potential biological hazards without any safety precautions.

7.1.1. Cleaning the Instrument

Turn off the instrument and unplug the power cord.

Use a damp soft cloth to clean the instrument shell, use a mild cleaner to clean if necessary.

Note: Do not spray the cleaner directly on the instrument, which may cause the failure of the electronic instrument.

Please avoid using strong acid and alkali, more than 75% concentrated alcohol, organic solvents or strong detergent and other cleaning disinfectants to clean the instrument without our company‘s permission, it will destroy the surface coating.

7.1.2. Clean Sample-tray and Reaction-holes

Use damp soft cloth to clean the sample tray, use mild detergent to clean it if necessary.

Please clean the reaction hole regularly, to ensure that the consumable test tube is in full contact with the reaction hole and the heat conduction is perfect.

Use cotton swabs dipped in mild detergent to clean the reaction hole.

Please close the heat cover after the sample tray and reaction hole are completely dry. The reaction module with detergent on the heating surface may damage the instrument.

Do not spray the cleaner directly on the instrument, which may cause the failure of the electronic instrument.

7.2 Instrument maintenance

7.2.1. Keep Air-Circulation

The instrument must be positioned with a possibility of free air circulation to ensure accuracy of the target temperature. Please check the placement area regularly to ensure a free air circulation, and no items that can affect the air flow.

7.2.2. Keep Stable Electricity Supply

A stable power supply is required for the normal operation, please check it regularly to ensure that the supply voltage is consistent with the voltage requirement (±10% deviation is allowed). Ensure that the power outlet rated load is not less than the instrument requirement.

7.2.3. Keep Clean

Sample tray, reaction modules and optical components contaminations may interfere with thermal cycling and data collection. Please refer to the following instructions to avoid contamination of this instrument:

Please confirm the seal of the tubes or plates before running the reaction.

Do not put open reaction consumables or damaged lids into the sample tray. The reagent may leak out and cover the hot-lid surface, sample tray and reaction hole.

Do not run PCR reactions containing volatile reagents, which may lead to contamination in the hot-lid, sample tray and reaction hole.

When this instrument is not used for a long time, please unplug and cover with soft cloth or dustproof bag to prevent dust from entering.

7.2.4. Waste Disposal

After each experiment, there is a large number of amplified products in the test tube, which should be treated as soon as possible according to relevant regulations, to avoid contamination of laboratories and instruments.

Do not open the test tube cap after removing it from the reaction module! The high concentration of nucleic acid might pollute the laboratory.

7.2.5. Fuse Replacement

When the circuit fails or is abnormal, current increase or rising of the current may damage important components. If the fuse is installed correctly, the fuse will break when the current rises to a certain height and heat, to protect the instrument or the user.



FastGene® qFYR Plusw Real-Time PCR System are equipped with two 10 A. When the fuse is damaged, the user can replace the fuse as follows:

- Cut off the mainframe power supply and unplug the power cord
- Open the fuse box lid with a screwdriver
- Check the inner fuse tube. If the middle is disconnected (two or one of them), it means that the fuse is broken
- Replace the new fuse tube (fuse type Φ5×20 mm –T10AH250VP)
- Install the replaced fuse seat under the socket.
- **Note:** Power must be turned off and removed before replacing fuse.





7.2.6. Overheating Protection

The instrument heating system is equipped with overheating protection. When the heating system fails and the temperature value exceeds the upper limit of the allowable range, the protection will automatically disconnect and cannot be restored. At this time, the heating system cannot continue to rise and cool normally. After the heating system breaks down, user should stop using the instrument, contact the manufacturer for maintenance in time.

8. Troubleshooting

a)
Observation: Switch on the instrument, but no reaction, and power light off.
Possible cause: Inconsistent connection with power cable
Recommended action: Switch off the power on the rear of instrument, re-plug power cable.

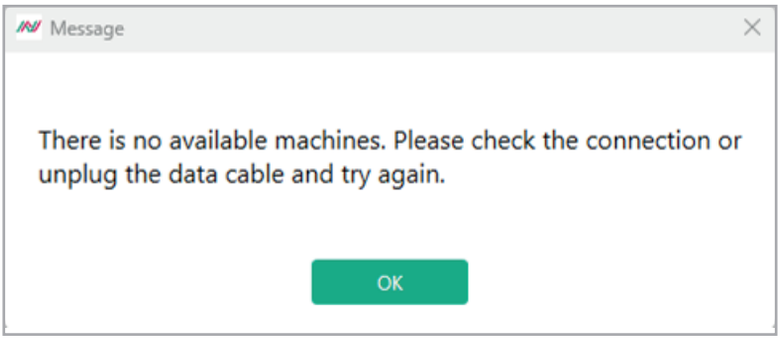


Inconsistent plug



correct plug

b)
Observation: Shows “No Available Instrument” after click “Start Run”.
Possible cause: Inconsistent connection between instrument and computer.
Recommended action: Unplug and plug the USB data cable again, re-start qFYR Analysis Studio.





Inconsistent data cable



Correct connection

c)
Observation: There is no fluorescence signal appearing on “Multicomponent plot”, while the sound of scanning can be heard.
Possible cause: The „sample“ and „target item“ columns on the „well-plate setup“ interface are not filled in. The fluorescence signal is displayed before filled in both. If the user does not fill in „sample“ or „target item „, the instrument will collect and store the fluorescence signal of all channels in 96 wells in the experimental file but not display them on the experimental interface. It can be displayed after the user completes the“ well plate setting „information.
Recommended action: Return to Plate Tab, complete the “Sample” and “Target” information.

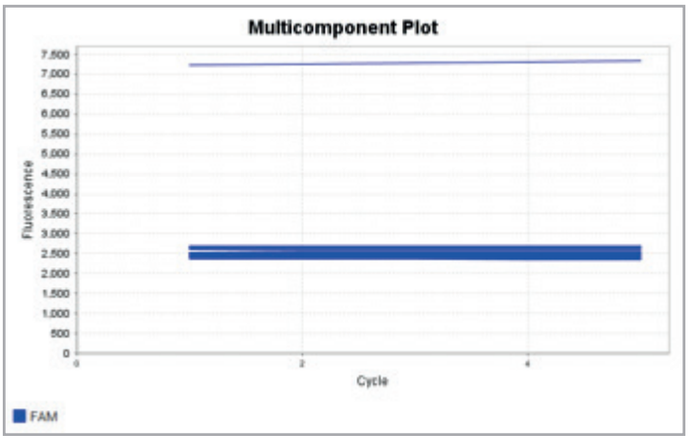
d)
Observation: Error notice of “Not enough memory, please close some experiment”.
Possible cause: Too many experiment files are opened.
Recommended action: Closed some inactive experiments, the error notice will be closed. It is recommended No more than six experiment files are opened simultaneously.

e)
Observation: The ramp rate slowed down obviously.
Possible cause: The outlet of airflow was blocked.
Recommended action: Clean the tamper on inlet and/or outlet of airflow.






f)
Observation: While scanning empty block, background signal of some wells is much higher than average.
Possible cause: There is contamination in these wells.
Recommended action: Clean the contamination.



g)
Observation: Reaction reagent been evaporated.
Possible cause: The tubes or seal on plate are not closed correctly.
Recommended action: Select better air-tight consumables and restart experiment.

Note: If the user cannot judge and troubleshoot the above, contact our company directly.

If the following circumstances occur, the power supply should be cut off immediately, contact the supplier or our company, we will assign qualified maintenance personnel to handle:

- Liquid flows inside the instrument.
- Instruments are rained or watered.
- The instrument has any abnormal sound or smell.
- Damage to the shell caused by instrument dropping down.
- Instrument function changes obviously.

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+49 2421 55496 0
+49 2421 55496 11
info@nippongenetics.eu
www.nippongenetics.eu